

# The particular species determining spatial heterogeneity in shady and terrace on the Qinghai-Tibetan Plateau, China



XiaoMing Mou<sup>a,b</sup>, YingWen Yu<sup>b,\*</sup>, Min Zhao<sup>b</sup>, YuQiang Li<sup>a</sup>, Masae Shiyomi<sup>c</sup>

<sup>a</sup> Northwest Institute of Eco-Environment and Resources, Naiman Desertification Research Station, Chinese Academy of Sciences, Lanzhou, Gansu 730000, China

<sup>b</sup> State Key Laboratory of Herbage Improvement and Grassland Agro-ecosystems, College of Pastoral Agriculture Science and Technology, Lanzhou University, Lanzhou, Gansu 730020, China

<sup>c</sup> Ibaraki University, Professor Emeritus, Bunkyo 2-1, Mito 310-8512, Japan

## ARTICLE INFO

### Keywords:

Dominant species

Power law

Qinghai-Tibetan Plateau

Spatial heterogeneity

Vegetation patches

## ABSTRACT

Spatial patterns of plant species and patchy community are important properties in grasslands. However, research regarding spatial patterns of formed patches with various species has not fully advanced until now. Our purpose is to clarify differences in spatial pattern formed by species and community constructed under shady and terrace habitats. The three common *Kobresia-Carex* patches (Size 1, 0.6–0.9 m<sup>2</sup>; Size 2, 3.0–3.8 m<sup>2</sup> and Size 3, 6.5–8.8 m<sup>2</sup>) were selected in shady and terrace on the Qinghai-Tibetan Plateau, and corresponding quadrats of 1m×1m, 2m×2m and 3m×3m were placed for S1, S2 and S3 patches, respectively. The surveyed quadrats were divided into 20cm×20cm large cells (L-cells), and further divided into four 10cm×10cm small cells (S-cells). We used the binary occurrence system (presence/absence data) to record occurrences of all species in S-cells. The analysis shows that the power law model was well able to determine the spatial distribution pattern of species or patchy community in shady and terrace. All species and patches show aggregated distribution in shady and terrace habitats. In the shady habitat, the relative spatial heterogeneity ( $\epsilon$ ) of individual plant species was lowest at presence frequency ( $P$ ) of 0.1–0.3, whereas in the terrace habitat  $\epsilon$  was lowest at  $P < 0.1$ , and  $\epsilon$  increased monotonically with increasing  $P$ . For most dominant species,  $P$  and  $\epsilon$  values were higher in terrace than those in shady. We concluded that the dominant species largely determine spatial heterogeneity of the *Kobresia-Carex* patches, while companion and rare species have weak influence on the community-level heterogeneity in shady and terrace habitats.

## 1. Introduction

In grassland vegetation, species- and community-level spatial heterogeneity synthetically are the main determinants of community dynamics and stability (Mitsunaga and Fujii, 1997; Shaibur et al., 2021), and affect habitat utilization and forage selection by herbivores (Huang et al., 2012). Understanding the processes which determine the spatial pattern of plant species and community can help recognize the mechanisms of grassland degradation and species diversity (Eppinga et al., 2022). The spatial heterogeneity of grasslands is influenced by various factors: (i) species composition and mode of reproduction (Zhang, 2005; Song et al., 2008). For example, Shiyomi et al. (2001) reported that plant species on a temperate mixed-seeding pasture was distributed more

heterogeneously than a random pattern in Japan. (ii) grazing management regime (Wang et al., 2002; Wen et al., 2013) and spatial distribution of excretion by livestock (Islam et al., 2010) or root foraging by burrowing rodents (Li et al., 2009; You et al., 2014); and (iii) habitat microclimate heterogeneity (Song et al., 2008), variability of water resources (Chen et al., 2007; Pueyo et al., 2013) or micro-geomorphology (Song et al., 2010; Casalini and Bisigato, 2017). Selim et al. (2021) found that herbaceous plants reveal a strong aggregated population and accommodates four dominant clusters along with soil salinity gradients in Shyamnagar, Bangladesh.

Patchiness is an important feature in grassland ecosystems (Ludwig et al., 2005; Wintle et al., 2019; Lindenmayer, 2019). Spatial-temporal variability of environments such as fluctuation in temperature, species

\* Corresponding author.

E-mail address: [yuyw@lzu.edu.cn](mailto:yuyw@lzu.edu.cn) (Y. Yu).



composition and mode of reproduction (Zhang, 2005; Song et al., 2008), selective feeding (Adler et al., 2001) and soil nutrient heterogeneity (Collins, 1989) contributes to the formation of vegetation patches with different species characteristics in grasslands. Thus far, studies on vegetation patches have focused on patch connection with grassland degeneration (Zhang et al., 2003), formation mechanisms and dynamics (Emmerson et al., 2012; Smith et al., 2014; Hao et al., 2017), species composition and interspecific association (Wei et al., 2016) and vegetation characteristics and soil features (Yang et al., 2014; Shi et al., 2015). However, few research has focused on spatial heterogeneity of species and patchy community, and how the occurrence of a given species influences the spatial pattern in two common environments in grasslands such as "shady" and "terrace" habitats. In our study, shady and terrace habitats are two typical environments of grasslands on the Qinghai-Tibetan Plateau.

The power law model was first proposed by Taylor (1961) as an empirical method for determining a population statistic with binary variable to indicate the intrinsic level of aggregation in populations. Taylor (1961) observed that when members of a population are randomly distributed in their environment, the sampling variance is equal to the mean. When they are more regularly distributed, the variance is less than the mean; and the aggregate distribution of a population indicates that variance is greater than the mean value. Hughes and Madden (1993) adopted Taylor's (1961) power law of binary variable in studying the number of infected plant individuals in phytopathology (Madden et al., 2018). The power law has since been further developed and applied to study the spatial heterogeneity of individual species and community-level spatial heterogeneity in grassland vegetation (Shiyomi et al., 2001, 2005; Song et al., 2005; Chen and Shiyomi, 2019). Taylor's power law is an empirical formula expressed by statistical regression, and the relationship between mean and variance is affected by the size of plant species and sampling quadrat (Jiménez et al., 2001; Chen et al., 2007). Therefore, whether the power law can be applied to determine the spatial heterogeneity of species and small patches in shady and terrace habitats on the Qinghai-Tibetan Plateau, remains largely unknown.

*Kobresia-Carex* meadows are widely distributed worldwide, and patchy distribution of *Kobresia-Carex* is a common feature on the Qinghai-Tibetan Plateau (Zhao et al., 2021). Patch configuration is viewed as an important factor affecting the distribution of plant species (Hemrová and Münzbergová, 2015). The diameter of larger patches is more than several tens of meters, and small patches are less than 1 m (Wen et al., 2013; Mou et al., 2015, 2020). Spatial distribution pattern is an important property of a plant community, and also is the fundamental parameter to describe a plant community. The actual distribution pattern of species and community reflects the current habitat and the presence of grassland degradation and species diversity. The aim of this paper is the clarification of characteristics of these two habitats based on spatial distribution of species and community. Here, we apply the power law to analyze the species- and community-level spatial heterogeneity on *Kobresia-Carex* patches with different sizes in these two different habitats in alpine meadows, and the occurrence of all species was investigated via binary occurrence counts. Our scientific questions are: (1) whether the power law can be applied to determine the spatial pattern at species- and community-levels in patches on shady and terrace habitats? (2) Do key species determine the community-level spatial heterogeneity in different habitats on the Qinghai-Tibetan Plateau? (3) How different is spatial heterogeneity between the two habitats in shady and terrace environments?

## 2. Methods

### 2.1. Study area

The study site is located at Gansu Agricultural University Experimental Station (37°12'N, 102°45'E; 2,960 m a.s.l.) in Tianzhu County, Gansu Province, China, eastern section of the Qinghai-Tibetan Plateau.

This area has been used for winter grazing by yak (*Bos grunniens*) and Tibetan sheep (*Ovis aries*) from November to the following May for more than 30 years. The climate in this region is influenced by the southeast monsoon and the Siberian high air mass, and is characterized by severe winters and short, cool summers. The mean annual temperature and precipitation are  $-0.1^{\circ}\text{C}$  and 416 mm, respectively. The temperature ranges from mean minimum of  $-18.3^{\circ}\text{C}$  in January to mean maximum  $12.7^{\circ}\text{C}$  in July. The plant growth period is about 120 days (June to September). Soil is classified as Sub-alpine Meadow Soil according to the Chinese Soil Classification system (Wu and Tiessen, 2002).

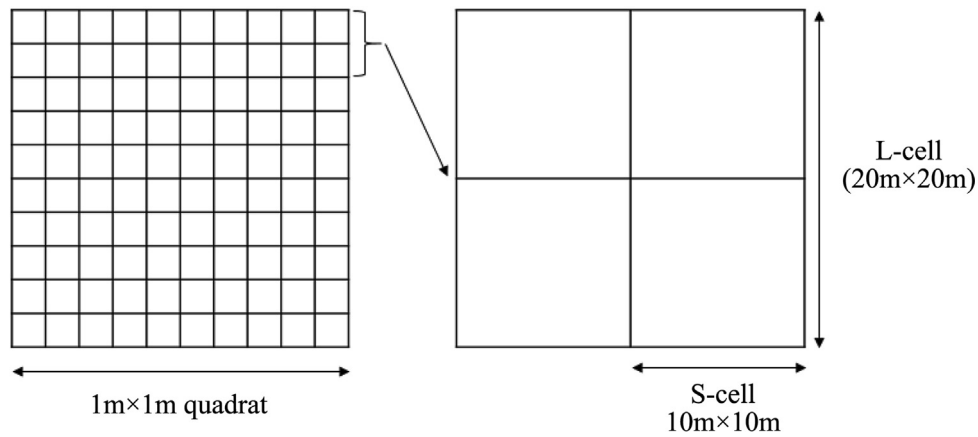
### 2.2. The setting of experimental plot

Our study was conducted on contiguous habitats of shady and terrace, each area about 10 ha. Shady and terrace are two major habitat types and together form unique landscapes on the Qinghai-Tibetan Plateau. The shady habitat has a slope of about  $18^{\circ}$ , and the terrace habitat is relatively flat. Dominant plant species in the shady habitat are *Carex* spp., *Kobresia capillifolia*, *Artemisia smithii* and *Leontopodium nanum*. For the terrace habitat, dominant species are *K. capillifolia*, *Carex* spp., *Melilotids ruthenica*, *Stipa krylovii*, and *L. nanum*. The grazing intensity was higher and 85%–90% of the aboveground biomass was consumed by animals. The grassland is composed of many mosaic patches which are dominated by different species (Wen et al., 2013). The "patch" forms distinct boundaries with other vegetation units within the grassland community and named according to the dominant species (Garbin et al., 2016; Mou et al., 2020; Zhao et al., 2021). It is possible that this mosaic distribution of vegetation on a field scale is a result of mode of reproduction (Zhang, 2005; Song et al., 2008), selective grazing by livestock and changes in micro-topography (Li et al., 2009; Wen et al., 2013). The dominant species of "*Kobresia-Carex* patches" are *K. humilis*, *K. capillifolia* and several *Carex* spp. which together comprise more than 75% of the aboveground biomass.

Based on patch area, three sizes of *Kobresia-Carex* patches (occupying more than 65% area) were selected in shady and terrace land according to the previous survey by Mou et al. (2020) on the Qinghai-Tibetan Plateau: Size 1 (referred to as S1,  $0.6\text{--}0.9\text{ m}^2$ ), Size 2 (S2,  $3.0\text{--}3.8\text{ m}^2$ ) and Size 3 (S3,  $6.5\text{--}8.8\text{ m}^2$ ).

### 2.3. Statistics regarding plant occurrence

Eight replicates of *Kobresia-Carex* patches for three sizes were randomly selected on shady and terrace habitats (2 habitats  $\times$  3 patch sizes  $\times$  8 replicates = 48 patches). According to patch size, quadrats were placed with  $1\text{m}\times 1\text{m}$  quadrats for S1 patch,  $2\text{m}\times 2\text{m}$  quadrats for S2 patch, and  $3\text{m}\times 3\text{m}$  quadrats for S3 patch, and more than 85% of patch area was sampled by corresponding quadrats. The quadrat was divided into  $20\text{cm}\times 20\text{cm}$  large-cells (L-cells), where each L-cell was further subdivided into four  $10\text{cm}\times 10\text{cm}$  small-cells (S-cells) (i.e., a  $1\text{m}\times 1\text{m}$  quadrat was composed of 25 L-cells, a  $2\text{m}\times 2\text{m}$  quadrat composed of 100 L-cells, and a  $3\text{m}\times 3\text{m}$  was composed of 225 L-cells, total 22,400 S-cells in this survey) (Fig. 1). Binary data are easier and less costly to collect in actual vegetation plots, and therefore are more commonly collected by vegetation scientists (Chen et al., 2021; Yamamura et al., 2021). We used a binary occurrence system to record occurrences of all species in S-cells (Shiyomi et al., 2000, 2001). If a specified plant species does not occur in any of the four S-cells, we record 0; if the species occurs in any one of the four S-cells, this was record as 1; if the species occurs in any two S-cells, this was recorded as 2, etc. Therefore, the number of S-cells in an L-cell in which species *i* occurred was 0, 1, 2, 3, or 4 (Fig. 1b). The proportion of occurrence of species *i* in the S-cells for each patch was calculated (as values  $P_i$ ). The dominant, companion and rare species were classified using frequency of occurrence, according to Mariotte's (2014) criterion: (1) "Dominant species", species with  $P_i \geq 0.4$ ; (2) "Companion species", species with  $0.1 \leq P_i < 0.4$ ; and (3) "Rare species", species with  $P_i < 0.1$ .



**Fig. 1.** The quadrats (1m×1m, 2m×2m and 3m×3m), large cells (L-cells) and small-cells (S-cells) were used in vegetation survey. The survey quadrats were divided into 20cm×20cm L-cells; and L-cell was further subdivided into four 10cm×10cm S-cells. We used binary occurrence to record occurrences of species present in the S-cell.

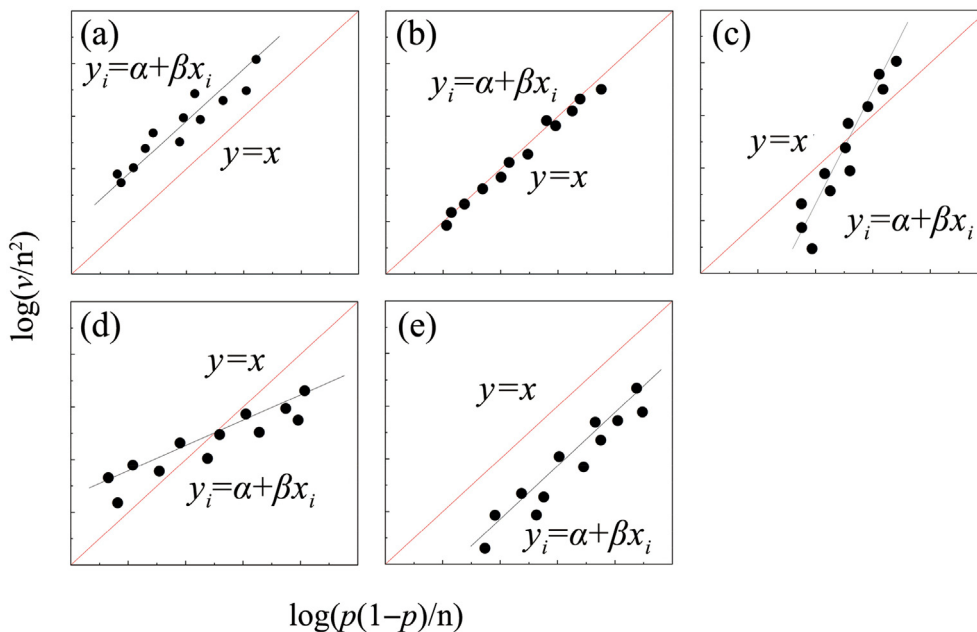
2.4. Model and analysis

2.4.1. The power law

We applied the power law as developed by Shiyomi et al. (2000, 2001) to determine the spatial pattern at species- and community-levels on shady and terrace, where the raw data are the sum of binary occurrences (0 or 1 for absent or present, respectively) among the L-cells of an S-cell (i.e., 0, 1, 2, 3, or 4 in this study). The raw data such as 0, 1, 2, 3, or 4 per L-cell are the sum of binary occurrences, where for an S-cell in an L-cell 0 or 1 for absence or presence for each species is allotted, respectively. In the actual survey, the relative frequency of occurrence of species  $i$  per S-cell is expressed as  $P_i$ , where  $0 \leq P_i \leq 1$ . If it is assumed that occurrence of species  $i$  at an S-cell is random, the probability that the species occurs in  $j$  of 4 S-cells in an L-cell is expressed by  $\binom{4}{j} P_i^j (1 - P_i)^{4-j}$  (binomial distribution), and

the mean and variance are  $4P_i$  and  $4P_i(1-P_i)$ , respectively, where 4 is the number of S-cells in an L-cell,  $j = 0, 1, 2, 3, 4$ . This calculation is performed for  $s$  species;  $i = 1, 2, \dots, s$ .

The principle of the power law is to compare the theoretical variance above with the variance actually observed for occurrences of species among S-cells. The logarithm of observed variance for each of the  $s$  species is regressed on the logarithm of the theoretical variance expected for a random distribution, as above, any divergence from the regression line can provide information on whether community species is aggregated or regular distribution (Shiyomi et al., 2000). Regression residuals for individual species can reveal species specific properties in the community. Here, we designate the observed variance as  $v_i$  for species  $i$  and transform both the variances divided by 16 into a logarithmic scale:  $x_i = \log(P_i(1-P_i)/4)$ , and  $y_i = \log(v_i/16)$  (Shiyomi et al., 2000, 2001). The  $x_i$  and  $y_i$  could be empirically expressed by a linear-type equation:



**Fig. 2.** Hypothetical spatial heterogeneity of vegetation estimated by the power law, where each dot indicates a species. (a) If the straight (regression) line is located above  $y = x$ , the overall community tendency of spatial pattern is more heterogeneous than would be expected in a random pattern; (b) If the straight line coincides with  $y = x$ , the overall tendency is random; (c) and (d) If the straight line intersects  $y = x$ , the spatial patterns of species below the straight line are less heterogeneous and the spatial patterns of species above the straight line are more heterogeneous than in a random pattern; (e) If the straight line is located below the  $y = x$  line, the pattern is less heterogeneous than would be expected in a random pattern.

$y_i = \alpha + \beta x_i + \varepsilon_i$ , where  $\alpha$  and  $\beta$  are constants for the regression expressing community distribution properties and  $\varepsilon_i$  is a species-specific residual (referred to as "relative spatial heterogeneity" hereafter).

Since  $P_i(1-P_i)/4$  for species  $i$  takes values between 0 and 1/16,  $\log [P_i(1-P_i)/4]$  reaches a maximum at  $P_i = 1/2$ . Therefore, the value of  $\log [P_i(1-P_i)/4]$  increases with increase in  $P_i$  as  $P_i$  increases from 0 to 1/2, and then decreases with further increase of  $P_i$  as  $P_i$  increases from 1/2 to 1. The interpretation of different possible patterns in the regression given by Shiyomi et al. (2001) is as follows: (1) if  $\alpha = 0$  and  $\beta = 1$ , then the community as a whole has a random spatial distribution (Fig. 2); (2) if for any  $\alpha$  and  $\beta \neq 1$ , then the community as a whole exhibits more aggregated or regular than random spatial distribution (Figs. 2a–2e); (3) if  $\beta < 1$ , then species with larger  $\log [P_i(1-P_i)/4]$  (i.e.,  $P_i$  more near to 1/2) are displaying a more regular spatial distribution in the community than ones with smaller  $\log [P_i(1-P_i)/4]$ , with a random distribution indicated by the point where the regression line crosses the  $y = x$  line; (4) The reverse applies where  $\beta > 1$  (Fig. 2).

2.4.2. The spatial heterogeneity indices at species- and community-levels

First, we define the absolute spatial heterogeneity for species  $i$ ,  $\delta_i$ . In the scatter diagram of  $(x_i, y_i)$ , the vertical distance,  $\delta_i$ , between  $y_i$  and the

straight line  $y = x$  can be calculated by  $y_i - x_i$ . When  $\delta_i = 0$ ,  $\delta_i > 0$  or  $\delta_i < 0$ , the spatial pattern of species  $i$  is referred to, respectively, as being a random pattern; as having higher heterogeneity (more aggregation) than would be found in a random pattern; or as having lower heterogeneity (more uniformity) than would be a random pattern (Shiyomi et al., 2001, 2005; Chen and Shiyomi, 2019; Yamamura et al., 2021).

Next, we define the relative spatial heterogeneity for evaluating the status of species  $i$  within a community, that is, the residual of species  $i$ ,  $\varepsilon_i$ , expressed by  $\varepsilon_i = y_i - \alpha - \beta x_i$ , which indicates the degree of difference from the community-level heterogeneity described by the aforementioned linear equation. The following are determined for species  $i$  by  $\varepsilon_i = 0$ ,  $\varepsilon_i > 0$ , or  $\varepsilon_i < 0$ : the relative spatial heterogeneity of species  $i$  is the same as community-level spatial heterogeneity, larger tendency than the spatial heterogeneity of whole grassland community, or smaller tendency than the whole grassland community, respectively (Guan et al., 2016).

The following index,  $\delta_C$ , is a community-level spatial heterogeneity value. The community-level spatial heterogeneity is represented by the following equation (Shiyomi et al., 2001):

$$\delta_C = \frac{\sum_{i=1}^s P_i \delta_i}{\sum_{i=1}^s P_i} \tag{1}$$

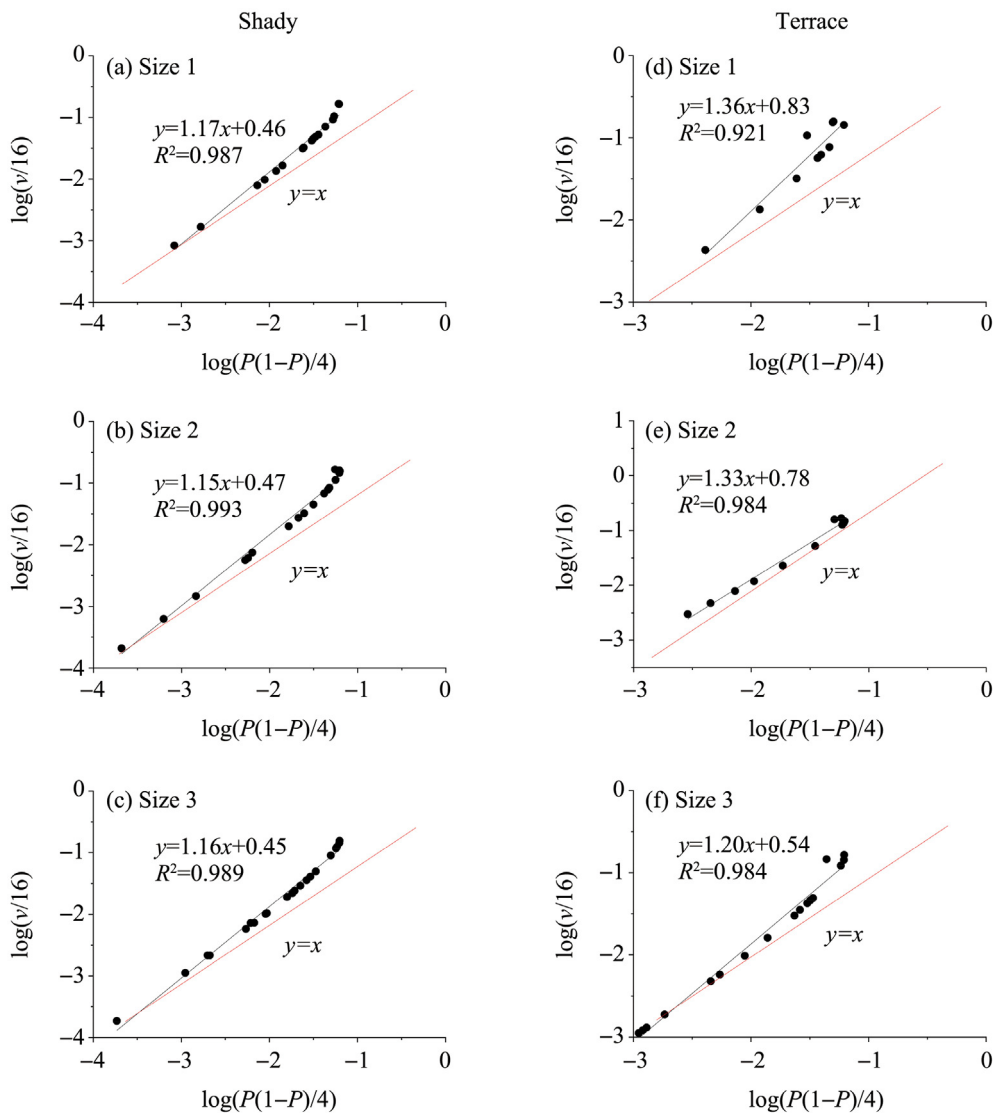


Fig. 3. Application of the power law to examine spatial heterogeneity of *Kobresia-Carex* patches of Size 1, Size 2 and Size 3 in shady (a, b and c) and terrace (d, e and f), the dot represents different plant species. Dots on red lines indicate species which are randomly distributed, and dots above red lines indicate species which distribute more heterogeneity than random pattern.



where  $s$  is the total number of species in the community,  $P_i$  and  $\delta_i$  indicate the relative frequency of occurrences per S-cell and the absolute spatial heterogeneity value for species  $i$ .

2.5. Visualization of plant spatial distribution

In order to visualize the spatial distribution of plant individuals and community, we used photography combined with geographic information software to determine the location of individual plants on shady and terrace habitats. The 10m×10m community segments were randomly selected in shady and terrace habitats, then divided into 50cm×50cm quadrats, respectively. We take the photograph (Nikon D100) on each quadrat from left to right, bottom to top in 10m×10m community segments away from 1 m to ground. The photos (total 800) were imported into ArcGIS 10.8, and determined plant cluster coordinates according to the location of each plant. We established the spatial location for all plant individuals in 10m×10m community on the shady and terrace habitats, and then outputted the distribution map of total species and single species of *Kobresia-Carex*.

3. Results

3.1. Application of the power law to the *Kobresia-Carex* patches

Fig. 3 shows the regressions of  $\log(v_i/16)$  on  $\log(P_i(1-P_i)/4)$  for three patch sizes in two habitats. The coefficients of determination ( $R^2$ ) for the power law regressions were high, and most of the species lie above the line of  $y = x$ . Consequently, the power law could determine the spatial distribution of species and patch communities with different size. These results demonstrate that species in these plant communities in both habitats show higher spatial heterogeneity (i.e., spatial aggregation). In addition, for species in the shady habitat, those with high values of  $\log(P_i(1-P_i)/4)$ , generally also exhibited higher  $\delta_i$  values (i.e.,  $\beta > 1$ ) than terrace habitat. The slope  $\beta \approx 1$  in terrace indicates that the spatial heterogeneity of individual plant species is not closely linked with  $P_i$  (Fig. 3e).

3.2.  $P$  and  $\delta$  for each species

In the shady habitat,  $P$  and  $\delta$  values for dominant species of *K. capillifolia* continuously increased with patch size, while *L. nanum* (one of the dominants) increased with patch size and then stabilized (Table 1). In the shady habitat,  $P$  and  $\delta$  values for *Gentiana scabra* and *Astragalus membranaceus* were higher in S1 than S2 and S3. However, in terrace,  $\delta$  values for dominants species *Melilotoids ruthenica* and *S. krylovii* monotonically decreased with patches size, while  $P$  and  $\delta$  values for *K. capillifolia* and *G. scabra* were lower in patch S2 than S1 and S3.

For companion species, in the shady habitat,  $P$  and  $\delta$  values for *Allium sikkimense* and *Kobresia humilis* gradually decreased from S1 to S3, but *Polygonum viviparum* increased from S1 to S2 and then decreased from S2 to S3 (Table 1); In terrace,  $P$  and  $\delta$  values for *A. membranaceus* first increased with patch size and then decreased, while these two values for *K. humilis* monotonically decreased from patch S1 to S3 in terrace. For rare species, in the shady habitat,  $P$  and  $\delta$  values for *Gentiana scabra* increased and then decreased with patch size, but the other rare species fluctuated among patch sizes.

In our surveys, all species show a heterogeneous pattern ( $\delta_i > 0$ ) in two habitats with three patch sizes (Table 1).

3.3.  $P$  and  $\epsilon$  for each species

The  $P$  and relative spatial heterogeneity ( $\epsilon$ ) varied among the three patch sizes both in shady and terrace. *K. capillifolia*, *M. ruthenica* and *Carex humilis* show high  $P$  and  $\epsilon$  values in S3 patch in the terrace habitat;  $\epsilon$  values for *G. scabra* and *A. membranaceus* were high at S1 patch in shady; and  $\epsilon$  for *L. nanum* and *P. viviparum* in S3 patch in the shady

Table 1

The presence frequency ( $P$ ) and absolute spatial heterogeneity ( $\delta$ ) for individual plant species on *Kobresia-Carex* patches of Size 1, Size 2 and Size 3 in shady and terrace.

Plant species and abbreviations under the shady and terrace	Size 1		Size 2		Size 3	
	$P$	$\delta$	$P$	$\delta$	$P$	$\delta$
<b>&lt;Shady habitat&gt;</b>						
<i>Carex</i> sp.	0.577	0.435	0.243	0.232	0.354	0.314
<i>Artemisia smithii</i>	0.560	0.428	0.524	0.400	0.460	0.366
<i>Leontopodium nanum</i>	0.317	0.288	0.668	0.477	0.644	0.458
<i>Polygonum viviparum</i>	0.300	0.247	0.558	0.418	0.527	0.366
<i>Kobresia capillifolia</i>	0.220	0.219	0.498	0.397	0.500	0.398
<i>Melilotoids ruthenica</i>	0.173	0.167	0.687	0.482	0.135	0.147
<i>Oxytropis kansuensis</i>	0.157	0.165	0.148	0.157	0.120	0.131
<i>Stipa krylovii</i>	0.147	0.157	0.257	0.247	0.273	0.260
<i>Kobresia pygmaea</i>	0.140	0.146	0.006	0.007	0.009	0.011
<i>Allium</i> sp.	0.110	0.123	0.094	0.108	0.084	0.097
<i>Kobresia humilis</i>	0.107	0.120	0.338	0.302	0.373	0.326
<i>Astragalus membranaceus</i>	0.060	0.072	0.457	0.371	0.100	0.112
<i>Anaphalis lactea</i>	0.050	0.058	0.212	0.212	0.161	0.167
<i>Veratrum baillonii</i>	0.037	0.045	0.022	0.027	0.038	0.046
<i>Stellera chamaejasme</i>	0.030	0.037	0.111	0.121	0.068	0.080
<i>Agropyron cristatum</i>	0.007	0.008	0.003	0.003	–	–
<i>Stipa purpurea</i>	0.003	0.004	–	–	–	–
<i>Gentiana scabra</i>	0.003	0.004	0.071	0.083	0.022	0.028
<i>Thalictrum aquilegifolium</i>	–	–	0.006	0.007	0.039	0.047
<i>Youngia japonica</i>	–	–	–	–	0.004	0.006
<i>Elymus nutans</i>	–	–	–	–	0.079	0.080
<i>Leymus secalinus</i>	–	–	0.001	0.001	–	–
<i>Koeleria cristata</i>	–	–	0.003	0.003	0.001	0.001
<i>Aster</i> ssp.	–	–	0.023	0.029	0.028	0.034
<i>Dracocephalum heterophyllum</i>	–	–	0.001	0.001	–	–
<i>Poa</i> sp.	–	–	0.001	0.001	–	–
<b>&lt;Terrace&gt;</b>						
<i>Melilotoids ruthenica</i>	0.860	0.551	0.628	0.459	0.551	0.423
<i>Stipa krylovii</i>	0.727	0.495	0.168	0.174	0.150	0.161
<i>Kobresia capillifolia</i>	0.720	0.495	0.714	0.497	0.772	0.520
<i>Carex</i> sp.	0.443	0.366	0.432	0.357	0.443	0.364
<i>Leontopodium nanum</i>	0.247	0.220	0.388	0.332	0.160	0.165
<i>Astragalus membranaceus</i>	0.197	0.198	–	–	0.119	0.132
<i>Artemisia smithii</i>	0.180	0.186	0.473	0.380	0.367	0.321
<i>Kobresia humilis</i>	0.110	0.117	0.044	0.052	0.037	0.044
<i>Allium</i> sp.	0.050	0.054	–	–	–	–
<i>Oxytropis kansuensis</i>	0.017	0.021	0.081	0.091	0.140	0.149
<i>Potentilla bifurca</i>	–	–	0.030	0.035	0.004	0.006
<i>Leymus secalinus</i>	–	–	0.018	0.022	0.105	0.108
<i>Polygonum viviparum</i>	–	–	0.012	0.015	0.019	0.023
<i>Plantago lesngi</i>	–	–	–	–	0.059	0.069
<i>Stellera chamaejasme</i>	–	–	–	–	0.022	0.026
<i>Youngia japonica</i>	–	–	–	–	0.007	0.009
<i>Elymus nutans</i>	–	–	–	–	0.005	0.007
<i>Aster</i> ssp.	–	–	–	–	0.005	0.006

Size 1: 0.6–0.9 m<sup>2</sup>, Size 2: 3.0–3.8 m<sup>2</sup>, and Size 3: 6.5–8.8 m<sup>2</sup>; A dash (–) indicates that the species not found in the four S-cells.

habitat were also large and lay above  $y = 0$  (Fig. 4). This indicates that their relative spatial heterogeneities were higher than the other species in the community. The  $\epsilon$  value for *A. membranaceus* in S1 patch was shady > terrace, and this species has the maximum  $\epsilon$  value (0.159) in S2 patch in terrace. In both habitats,  $\epsilon$  values of most companion species lay below the line  $y = 0$ , indicating a tendency towards more regular distribution than the dominant species of the community. However,  $\epsilon$  values for *S. krylovii* in S2 and S3 patches in terrace lay near  $y = 0$ , indicating that the spatial heterogeneity of *S. krylovii* is more coincident with the community spatial heterogeneity. For rare species,  $\epsilon$  values were various among patch sizes.

A general pattern of  $\epsilon$  values across species and patch size was lowest at  $P_i$  of 0.1–0.3 in shady, whereas in terrace  $\epsilon$  was lowest at  $P < 0.1$ , and  $\epsilon$  increased monotonically with  $P_i$  increasing. For most dominant species,  $P$  and  $\epsilon$  values were higher in terrace than those in shady.

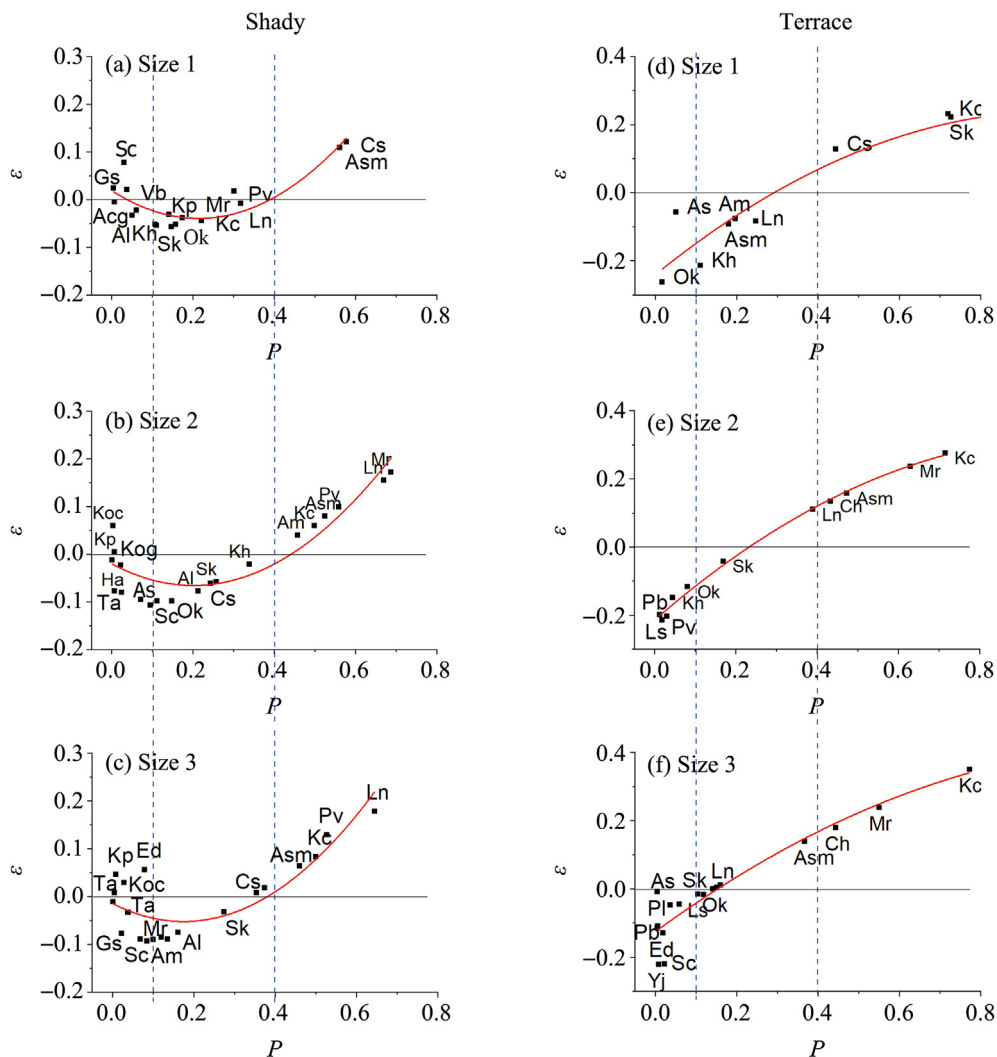
### 4. Discussions

#### 4.1. Evaluation of the spatial pattern by the power law

For biological populations which may consist of a small to large number of individuals, the following indices has been developed for measuring the spatial aggregation of individuals: variance/mean ratio (David and Moore, 1954; Taylor, 1961), Morisita's  $I_{\delta}$ -index index (1961, 1962),  $m-m^*$  method (Iwao, 1968), and  $k$  of negative binomial distribution (Pielou, 1977). However, for biological populations in which the occurrence or incidence is restricted to a binary variable such as infected or uninfected plant, and present or absent in a plant species, these indices are not applicable. Therefore, new indices for binary variables have been developed such as Morisita's (1962)  $I_B$  index, an index of spatial heterogeneity ( $\delta$ ) by Shiyomi et al. (1979). The  $\delta$  in this study is constructed based on the same logical base as the last two measures for the binary variable by power law model. Also, the power law has been applied by many researchers (Shiyomi et al., 2001; Chen et al., 2007; Xie et al., 2009; Chen and Shiyomi, 2019; Mou et al., 2020) to analyze the spatial heterogeneity of plant communities in grasslands. High coefficients of

determination ( $R^2$ ) for linear power law equations show that the spatial heterogeneity of individual species and the community could well fit the power law.

Index of spatial heterogeneity (aggregation) is sensitive to quadrat scale (Shiyomi et al., 2001; Song et al., 2005). The relationship between mean and variance in Taylor's power laws is affected by sample size (Chen et al., 2007). Crown width of species and quadrat scale could affect statistical results when using the binary occurrence system (presence/absence data) to record occurrences of all species, (Jiménez et al., 2001). In previous studies, a 50cm×50cm L-cell is used in analyzing community heterogeneity (Shiyomi et al., 2001, 2005; Song et al., 2005; Chen et al., 2007; Chen and Shiyomi, 2019; Yamamura et al., 2021). In our patch surveys, we used 20cm×20cm L-cells to evaluate the spatial heterogeneity of small patches formed by the small *Kobresia-Carex* community. The current survey not only obtained a measurement precision to the ground species survey, but also meets a high  $R^2$  value in the power law. Thus, the quadrat size (L-cells and S-cells) should be adjusted according to characteristics of a plant community when the power law is applied to measure the spatial heterogeneity of individual species and community in the grassland.



**Fig. 4.** The relationships between presence frequency ( $P$ ) and relative spatial heterogeneity ( $\epsilon$ ) on *Kobresia-Carex* patches of Size 1, Size 2 and Size 3 in shady (a, b and c) and terrace (d, e and f). The black point represents different species. Acg, *Agropyron cristatum*; Al, *Anaphalis lactea*; Am, *Astragalus membranaceus*; As, *Allium sikimense*; Asm, *Artemisia smithii*; Ch, *Carex humilis*; Ed, *Elymus dahuricus*; Gs, *Gentiana scabra*; Ha, *Heteropappus altaicus*; Kc, *Kobresia capillifolia*; Kh, *Kobresia humilis*; Koc, *Koeleria cristata*; Kp, *Kobresia pygmaea*; Ln, *Leontopodium nanum*; Ls, *Leymus secalinus*; Mr, *Melilotoids ruthenica*; Ok, *Oxytropis kansuensis*; Pb, *Potentilla bifurca*; Pc, *Poa crymophila*; Pl, *Plantago lesngi*; Pv, *Polygonum viviparum*; Sc, *Stellera chamaejasme*; Stipa krylovii, *Stipa krylovii*; Sp, *Stipa purpurea*; Ta, *Thalictrum aquilegifolium*; Vb, *Veratrum baillonii*; Yj, *Youngia japonica*. The relative spatial heterogeneity  $\epsilon$ , is effective for analyzing within-structure of communities.

Taylor's power law is not a theoretical formula obtained by mathematical deduction, but an empirical formula obtained by statistical regression. There are problems of significance test and confidence interval estimation of its parameters, especially the estimation and test of  $b$  value (Taylor, 1961; Madden et al., 2018; Chen and Shiyomi, 2019). Therefore, we plotted coordinates of  $\log(v_i/16)$  to  $\log(P_i(1-P_i)/4)$  for each species as scatter diagrams, and then obtained high  $R^2$  values for the linear power law equations (higher than 0.9) in all patch sizes and habitats (Fig. 3). This high degree of fit indicates that the power law well described the spatial pattern, and that the result from our study is consistent with results reported by Chen and Shiyomi (2019). Therefore, the present study confirms that the power law can be expanded to evaluate the spatial heterogeneity of species and community on small vegetation patches in shady and terrace habitats. Our research provides a new application of the power law model to study spatial pattern of species or patchy community in grasslands as reported by Tsuiqi et al. (2001), Song et al. (2005) and Xie et al. (2008). Also, the reasonability of the power law is discussed by Madden et al. (2018), and Chen and Shiyomi (2019). Thus, our first question that power law can be applied to determine the spatial pattern at species- and community-levels in patches on shady and terrace habitats is confirmed.

#### 4.2. Effects of species on community heterogeneity ( $\delta_C$ )

All species show aggregated distribution in patches on shady and terrace habitats, and that the results from our study is consistent with results reported by Shiyomi et al. (2001) in a temperate mixed-seeding pasture in Japan. In the present study, we found that dominant species were the key species determining community-level spatial heterogeneity ( $\delta_C$ ) in *Kobresia-Carex* patches, which is consistent with results of Mou et al. (2020). Song et al. (2005) also demonstrated that different species have different effects on spatial heterogeneity in grassland communities. Heterogeneity depends, in part, on the mode of propagation and reproduction (Soons et al., 2004). The dominant species of *Kobresia* and *Carex* reproduce through clonal underground tillering. They can resist grazing stress, and form a dense population "mosaic patch" on overgrazed grasslands. *Leontopodium nanum* can reproduce by the production of clonal ramets or by seeding. It can replace other species with its strong adaptation and resistance when facing harsh environments or heavy grazing (Ren et al., 2009). *A. membranaceus* is a species with thick-diameter roots radiating and branching from the stem base, which confers benefits for nutrient uptake and water storage. These dominants are ordinarily clonal species, and can subsequently attain a relatively

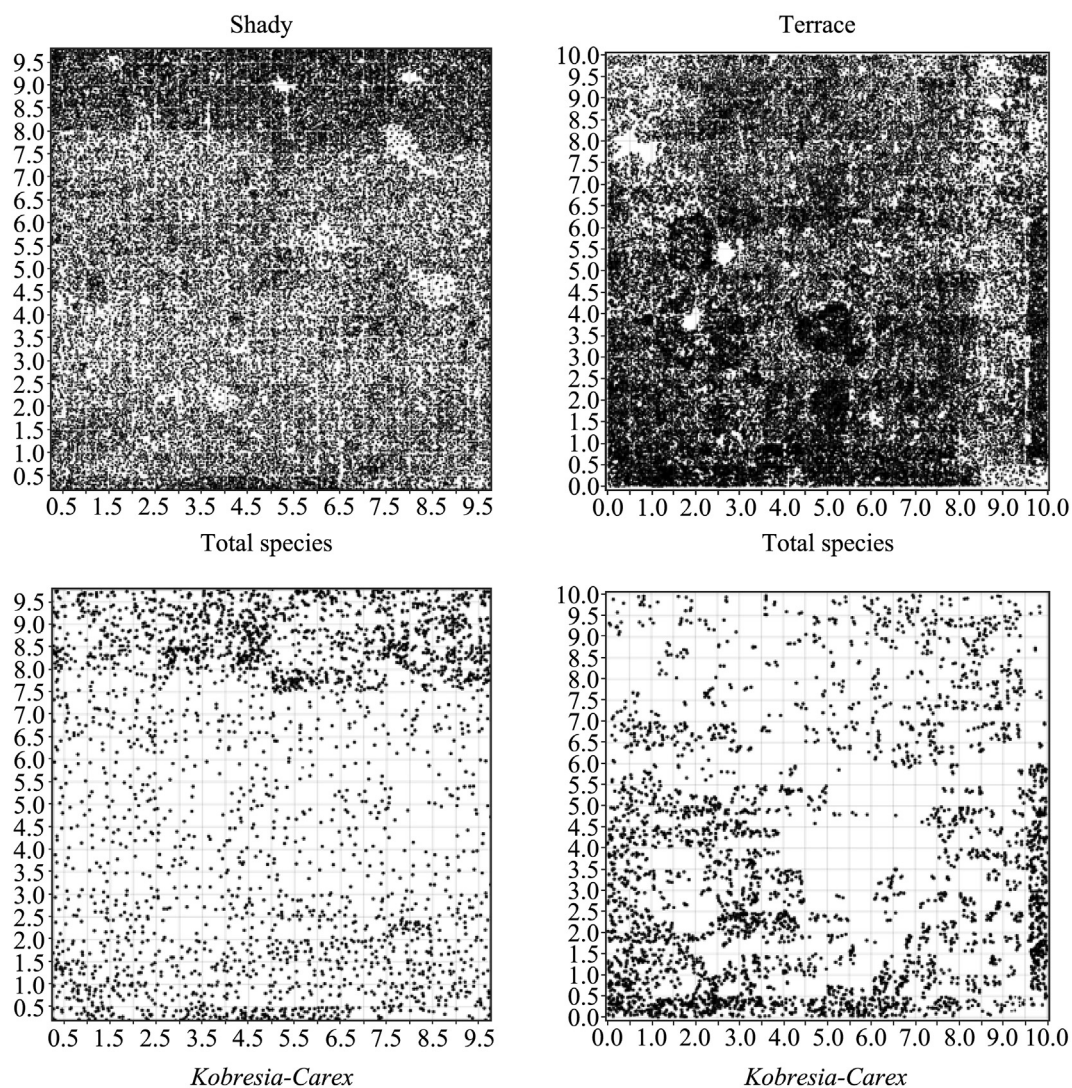


Fig. 5. The spatial distribution of individual plants (total species and *Kobresia-Carex*) in 10m×10m areas on shady and terrace habitats. The black points indicate the location of plant species in 10m×10m areas in different habitats.



high frequency, ground cover, and aboveground biomass, thus leading to a highly heterogeneous spatial distribution in these vegetation patches (Table 1). Thus, the second hypothesis that dominant species determine the community-level spatial heterogeneity in different habitats on the Qinghai-Tibetan Plateau is confirmed.

The companion species of *S. krylovii* and *P. viviparum* maintained a spatial heterogeneity ( $\delta$ ) close to community heterogeneity. *S. chamaejasme* is characterized by intense invasiveness, resource competitiveness and high adaptability to the environment, and then strongly suppresses growth of *S. krylovii* and *P. viviparum* (Ren et al., 2009; Liu et al., 2017). Additionally, *S. krylovii* populations are often overgrazed by cattle and sheep, so that they have difficulty forming a thick sward. The rare species do not have such a strong effect on the spatial aggregation of entire plant communities because these species have a small frequency presence. In addition, the heterogeneity of rare species was probably variable because their spatial pattern can be affected by accidental dispersion and concentration around mother plants or seeds. For example, *A. sikkimense* has a low frequency of occurrence and low spatial heterogeneity value (Table 1). One probable reason is that seeds are readily dispersed by wind or water (Geertsema and Sprangers, 2002; Liu et al., 2013). Therefore, the contribution of rare species to community heterogeneity of patch is weak both in shady and terrace. Consequently, all species show an aggregated distribution at different patch size in shady and terrace habitats, and spatial heterogeneity of the whole community in all patches were strongly affected by the dominant species.

#### 4.3. Relative spatial heterogeneity ( $\epsilon$ ) and habitat

The community-level spatial heterogeneity of *Kobresia-Carex* patches with three sizes differed distinctly in shady and terrace habitats (Fig. 5). Relative spatial heterogeneity ( $\epsilon$ ) of plant species in shady was lowest at  $P$ -values (frequency of occurrences) of 0.1–0.3 in different patch size; by contrast, the  $\epsilon$  value increased monotonically with increases of  $P$  in terrace (Fig. 4). Compared to the shady habitat, the  $\epsilon$  value in terrace was relatively high, and similar results were reported by Mou et al. (2015). The grazing intensity was higher and 85%–90% of the aboveground biomass was consumed by animals. The main cause of higher community-level heterogeneity in terrace than that in the shady habitat might be overgrazing (Adler et al., 2001). The selective feeding of livestock greatly affects the community structure, and the high grazing intensity is an important factor affecting spatial patterns formed by species and community under different habits.

Soil dryness and compaction are caused by excessive animal trampling at heavily grazed sites, resulting in the non-uniform distribution of water and nutrients (Kubo and Isobe, 1975). These altered conditions restrict species growth with lower stress tolerance, and therefore likely enhanced community-level heterogeneity in a terrace habitat (Molla and Huq, 2002). In addition, in a terrace habitat, the remarkably high heterogeneity in dominants, may correlate with facilitating processes linked to intense interspecific competition (Gutierrez-Giron and Gavilan, 2010), further contributing to a higher spatial heterogeneity than that in the shady site (Fig. 5). The new concept of relative spatial heterogeneity is developed in this study to find differences in community structure including spatial pattern. Thus, our third question that the difference in spatial heterogeneity between the two habitats in shady and terrace environments is confirmed. The underlying mechanisms of spatial patterns of individual species and community are formed resulting from interactions between endogenous and exogenous processes. Further studies should be focused on interspecific competition (Berger et al., 2008; Liu et al., 2013; Garbin et al., 2016), species reproduction mode (Liu et al., 2017) and resource heterogeneity, to clarify and explain the spatial heterogeneity of species or patchy community in grasslands.

## 5. Conclusions

The power law was well able to determine the spatial distribution pattern of species or patchy community in shady and terrace environments. Relative spatial heterogeneity ( $\epsilon$ ) of plant species in shady was lowest at  $P$ -values (frequency of occurrences) of 0.1–0.3 in different patch size; by contrast, the  $\epsilon$  value increased monotonically with increases of  $P$  in terrace. Species and patchy communities show aggregated distribution, and dominant, companion and rare species have different influences for spatial heterogeneity of a patch community. The dominant species largely determine spatial heterogeneity of the *Kobresia-Carex* patches, while companion and rare species have weak influence on community-level heterogeneity in shady and terrace habitats. Our research will promote a new application of the power law model in studying community structure including spatial pattern of species or small patchy community in grasslands.

## Acknowledgments

We thank the editors and three anonymous reviewers for very helpful suggestions. We also thank Cory Matthew from Massey University for assistance with embellishment of manuscript. This study was funded by The Second Tibetan Plateau Scientific Expedition and Research (STEP) program (Grant No. 2019QZKK0305), Youth Science and Technology Fund Program of GanSu (Grant No. 22JR5RA083) and the National Natural Science Foundation of China (Grant No. 31971466).

## References

- Adler, P.B., Raff, D.A., Lauenroth, W.K., 2001. The effect of grazing on the spatial heterogeneity of vegetation. *Oecologia* 128 (4), 465–479. <https://doi.org/10.1007/s004420100737>.
- Berger, U., Piou, C., Schifffers, K., et al., 2008. Competition among plants: concepts, individual-based modelling approaches, and a proposal for a future research strategy. *Perspect. Plant Ecol. Evol. Systemat.* 9 (3–4), 121–135. <https://doi.org/10.1016/j.ppees.2007.11.002>.
- Casalini, A.I., Bisigato, A.J., 2017. Geomorphology and soils control vegetation heterogeneity through differential species establishment at an arid ecotone. *J. Arid Environ.* 147, 83–89. <https://doi.org/10.1016/j.jaridenv.2017.08.004>.
- Chen, J., Huang, D.M., Shiyomi, M., et al., 2007. Spatial heterogeneity and diversity of vegetation at the landscape level in Inner Mongolia, China, with special reference to water resources. *Landscape Urban Plann.* 82 (4), 222–232. <https://doi.org/10.1016/j.landurbplan.2007.02.011>.
- Chen, J., Shiyomi, M., 2019. A power law model for analyzing spatial patterns of vegetation abundance in terms of cover, biomass, density, and occurrence: derivation of a common rule. *J. Plant Res.* 132 (4), 481–497. <https://doi.org/10.1007/s12655-019-01116-8>.
- Chen, J., Shiyomi, M., Huang, D., et al., 2021. Quantitative evaluation of species composition dissimilarity within a community and among communities. *Ecol. Res.* 36 (1), 152–160. <https://doi.org/10.1111/1440-1703.12192>.
- Collins, S.L., 1989. Experimental analysis of patch dynamics and community heterogeneity in tallgrass prairie. *Vegetation* 85 (1), 57–66. <https://doi.org/10.1007/BF00042255>.
- David, F., Moore, P.G., 1954. Notes on contagious distributions in plant populations. *Ann. Bot. (London) N.S.* 18 (1), 47–53. <https://doi.org/10.1093/oxfordjournals.aob.a083381>.
- Emmerson, L.M., Facelli, J.M., Chesson, P., et al., 2012. Changes in seed dispersal processes and the potential for between-patch connectivity for an arid land daisy. *Ecology* 93 (3), 544–553. <https://doi.org/10.1890/11-0651.1>.
- Eppinga, M.B., Haber, E.A., Sweeney, L., et al., 2022. *Antigonon leptopus* invasion is associated with plant community disassembly in a Caribbean island ecosystem. *Biol. Invasions* 24 (2), 353–371. <https://doi.org/10.1007/s10530-021-02646-w>.
- Garbin, M.L., Guidoni-Martins, K.G., Hollunder, R.K., et al., 2016. Spatial segregation of subordinate species is not controlled by the dominant species in a tropical coastal plant community. *Perspect. Plant Ecol. Evol. Systemat.* 18, 23–32. <https://doi.org/10.1016/j.ppees.2015.12.002>.
- Geertsema, W., Sprangers, J.T.C.M., 2002. Plant distribution patterns related to species characteristics and spatial and temporal habitat heterogeneity in a network of ditch banks. *Plant Ecol.* 162 (1), 91–108. <https://doi.org/10.1023/A:1020336908907>.
- Guan, Q.Q., Chen, J., Wei, Z.C., et al., 2016. Analyzing the spatial heterogeneity of number of plant individuals in grassland community by using power law model. *Ecol. Model.* 320, 316–321. <https://doi.org/10.1016/j.ecolmodel.2015.10.019>.
- Gutierrez-Giron, A., Gavilan, R.G., 2010. Spatial patterns and interspecific relations analysis help to better understand species distribution patterns in a Mediterranean high mountain grassland. *Plant Ecol.* 210 (1), 137–151. <https://doi.org/10.1007/s11258-010-9745-6>.



- Hao, H.M., Huang, Z., Lu, R., et al., 2017. Patches structure succession based on spatial point pattern features in semi-arid ecosystems of the water-wind erosion crisscross region. *Glob. Ecol. Conserv.* 12, 158–165. <https://doi.org/10.1016/j.gecco.2017.11.001>.
- Hemrová, L., Münzbergová, Z., 2015. The effects of plant traits on species' responses to present and historical patch configurations and patch age. *Oikos* 124 (4), 437–445. <https://doi.org/10.1111/oik.01130>.
- Huang, Y., Wang, L., Wang, D.L., et al., 2012. The effect of plant spatial pattern within a patch on foraging selectivity of grazing sheep. *Landsc. Ecol.* 27 (6), 911–919. <https://doi.org/10.1007/s10980-012-9744-1>.
- Hughes, G., Madden, L.V., 1993. Using the beta-binomial distribution to discrete aggregated patterns of incidence. *Phytopathology* 83 (7), 759–763. <https://doi.org/10.1094/Phyto-83-759>.
- Islam, T., Fukuda, E., Shiyomi, M., et al., 2010. Effects of feces on spatial distribution patterns of grazed grassland communities. *Agric. Sci. China* 9 (1), 121–129. [https://doi.org/10.1016/S1671-2927\(09\)60075-4](https://doi.org/10.1016/S1671-2927(09)60075-4).
- Iwao, S., 1968. A new regression method for analyzing the aggregation pattern of animal populations. *Res. Popul. Ecol.* 10 (1), 1–20. <https://doi.org/10.1007/BF02514729>.
- Jiménez, J.J., Rossi, J.P., Lavelle, P., 2001. Spatial distribution of earthworms in acid-soil savannas of the eastern plains of Colombia. *Appl. Soil Ecol.* 17 (3), 267–278. [https://doi.org/10.1016/S0929-1393\(01\)00133-0](https://doi.org/10.1016/S0929-1393(01)00133-0).
- Kubo, S., Isobe, S., 1975. Treading on grassland, hoof pressure by unstrained walk. *Bull. Natl. Grassl.* 7, 33–38. *Research Institute*.
- Li, X.G., Zhang, M.L., Li, Z.T., et al., 2009. Dynamics of soil properties and organic carbon pool in topsoil of zokor-made mounds at an alpine site on the Qinghai–Tibetan Plateau. *Biol. Fertil. Soils* 45 (8), 865–872. <https://doi.org/10.1007/s00374-009-0398-3>.
- Lindenmayer, D., 2019. Small patches make critical contributions to biodiversity conservation. *Proc. Natl. Acad. Sci. USA* 116 (3), 717–719. <https://doi.org/10.1073/pnas.1820169116>.
- Liu, J., Wu, D., Peng, X., et al., 2013. Exogenous and endogenous determinants of spatial aggregation patterns in Tibetan Plateau meadow vegetation. *J. Plant Ecol.* 6 (4), 277–285. <https://doi.org/10.1093/jpe/rt041>.
- Liu, M.X., Li, L.R., Che, Y.D., et al., 2017. Spatial distribution pattern and association of *Kobresia humilis* and *Polygonum viviparum* under different slope aspects in a subalpine meadow of Gannan. *Chin. J. Ecol.* 36 (10), 2685–2691. <https://doi.org/10.13292/j.1000-4890.201710.024>.
- Ludwig, J.A., Wilcox, B.P., Breshars, D.D., et al., 2005. Vegetation patches and runoff-erosion as interacting ecohydrological processes in semiarid landscapes. *Ecology* 86 (2), 288–297. <https://doi.org/10.1890/03-0569>.
- Madden, L.V., Hughes, G., Moraes, W.B., et al., 2018. Twenty-five years of the binary power law for characterizing heterogeneity of disease incidence. *Phytopathology* 108 (6), 656–680. <https://doi.org/10.1094/PHYTO-07-17-0234-RVW>.
- Mariotte, P., 2014. Do subordinate species punch above their weight? Evidence from above- and below-ground. *New Phytol.* 203 (1), 16–21. <https://doi.org/10.1111/nph.12789>.
- Mitsunaga, T., Fujii, K., 1997. The effects of spatial and temporal environmental heterogeneities on persistence in a laboratory experimental community. *Res. Popul. Ecol.* 39 (2), 249–260. <https://doi.org/10.1007/BF02765271>.
- Molla, S.R., Huq, S.M.I., 2002. Solid waste management: effectiveness of composts on productivity of soils. *Khulna Univ. Stud. Bang.* 4 (1), 671–676.
- Morisita, M., 1961. Measuring of dispersion of individuals and analysis of the distributional patterns. *Jpn. J. Ecol.* 11 (6), 252. [https://doi.org/10.18960/seitai.11.6.252\\_3](https://doi.org/10.18960/seitai.11.6.252_3).
- Morisita, M., 1962. I-index, a measure of dispersion of individuals. *Res. Popul. Ecol.* 4 (1), 1–7. <https://doi.org/10.1007/BF02533903>.
- Mou, X.M., Yu, Y.W., Li, X.G., et al., 2020. Presence frequency of plant species can predict spatial patterns of the species in small patches on the Qinghai-Tibetan Plateau. *Glob. Ecol. Conserv.* 21, e00888. <https://doi.org/10.1016/j.gecco.2019.e00888>.
- Mou, X.M., Yu, Y.W., Wang, X.Z., et al., 2015. Analysis of community spatial pattern of *leontopodium nanum* patches in Qinghai-Tibetan Plateau. *Acta Ecol. Sin.* 35 (16), 5306–5315. <https://doi.org/10.5846/stxb201404190770>.
- Pielou, E.C., 1977. *Mathematical Ecology*. Wiley and Sons, New York, pp. 124–134.
- Pueyo, Y., Moret-Fernandez, D., Saiz, H., et al., 2013. Relationships between plant spatial patterns, water infiltration capacity, and plant community composition in semi-arid Mediterranean ecosystems along stress gradients. *Ecosystems* 16 (3), 452–466. <https://doi.org/10.1007/s10021-012-9620-5>.
- Ren, Q.J., Wu, G.L., Ren, G.H., 2009. Effect of grazing intensity on characteristics of alpine meadow communities in the eastern Qinghai-Tibetan Plateau. *Acta Pratacul. Sinica* 18 (5), 256–261. <https://doi.org/10.11686/cyxb20090533>.
- Selim, A., Bari, E., Rahaman, M.H., et al., 2021. Phytosociology and biodiversity of roadside herbs in a salinity-affected coastal area of Bangladesh. *Heliyon* 7 (8), e07813. <https://doi.org/10.1016/j.heliyon.2021.e07813>.
- Shaibur, M.R., Husain, H., Arpon, S.H., 2021. Utilization of cow dung residues of biogas plant for sustainable development of a rural community. *Curr. Res. Environ. Sustain.* 3, 100026. <https://doi.org/10.1016/j.crsust.2021.100026>.
- Shi, M.M., Zhang, Y.C., Zhang, D.Y., et al., 2015. Plant traits and soil properties jin pasture minni-patches in an alpine meadow. *Acta Pratacul. Sin.* 24 (9), 197–205. <https://doi.org/10.11686/cyxb2015083>.
- Shiyomi, M., Takahashi, S., Yoshimura, J., 2000. A measure for spatial heterogeneity of a grassland vegetation based on the beta-binomial distribution. *J. Veg. Sci.* 11 (5), 627–632. <https://doi.org/10.2307/3236569>.
- Shiyomi, M., Takahashi, S., Yoshimura, J., et al., 2001. Spatial heterogeneity in a grassland community: use of power law. *Ecol. Res.* 16 (3), 487–495. <https://doi.org/10.1046/j.1440-1703.2001.00411.x>.
- Shiyomi, M., Takai, A., 1979. The spatial pattern of infected or infested plants and negative hyper-geometric series. *Jpn. J. Appl. Entomol. Zool.* 23, 224–229. <https://doi.org/10.1303/jjaez.23.224>.
- Shiyomi, M., Yasude, T., Chen, J., 2005. Methods of grazing grassland vegetation survey. *Acta Agrest. Sinica* 13 (2), 149–158. <https://doi.org/10.11733/j.issn.1007-0435.2005.02.014>.
- Smith, A.G., McVinish, R., Pollett, P.K., 2014. A model for a spatially structured metapopulation accounting for within patch dynamics. *Math. Biosci.* 247, 69–79. <https://doi.org/10.1016/j.mbs.2013.11.001>.
- Song, B.M., Huang, D.M., Wang, R.Q., et al., 2008. A measure for spatial heterogeneity of vegetation in the center of Inner Mongolia. *Prog. Nat. Sci.* 18 (3), 289–295. <https://doi.org/10.1016/j.pnsc.2007.07.015>.
- Song, Y.Y., Li, Y.Y., Zhang, W.H., 2010. Analysis of spatial pattern and spatial association of *Haloxylon ammodendron* population in different developmental stages. *Acta Ecol. Sin.* 30 (16), 4317–4327. [https://doi.org/10.1016/S1872-5813\(11\)60001-7](https://doi.org/10.1016/S1872-5813(11)60001-7).
- Song, Z.Y., Huang, D.M., Shiyomi, M., et al., 2005. Spatial heterogeneity and variability of a large-scale vegetation community using a power-law model. *Tsinghua Sci. Technol.* 10 (4), 469–477. [https://doi.org/10.1016/S1007-0214\(05\)70102-4](https://doi.org/10.1016/S1007-0214(05)70102-4).
- Soons, M.B., Heil, G.W., Nathan, R., et al., 2004. Determinants of long-distance seed dispersal by wind in grasslands. *Ecology* 85 (11), 3056–3068. <https://doi.org/10.1890/03-0522>.
- Taylor, L.R., 1961. Aggregation, variance and the mean. *Nature* 189 (4766), 732–735. <https://doi.org/10.1038/189732a0>.
- Wang, Y.S., Shiyomi, M., Tsui, M., et al., 2002. Spatial heterogeneity of vegetation under different grazing intensities in the Northwest Heilongjiang Steppe of China. *Agric. Ecosyst. Environ.* 90 (3), 217–229. [https://doi.org/10.1016/S0167-8809\(01\)00217-1](https://doi.org/10.1016/S0167-8809(01)00217-1).
- Wei, B., Wang, Y., Guan, S.Q., et al., 2016. The vegetation composition and interspecific association of patches distribution in *polygonum viviparum* alpine meadow on northeast Qinghai-Tibetan. *Acta Pratacul. Sin.* 33 (8), 1618–1624. <https://doi.org/10.11829/j.issn.1001-0629.2015-0646>.
- Wen, L., Dong, S.K., Li, Y.Y., et al., 2013. The effects of biotic and abiotic factors on the spatial heterogeneity of alpine grassland vegetation at a small scale on the Qinghai-Tibetan Plateau (QTP), China. *Environ. Monit. Assess.* 185 (10), 8051–8064. <https://doi.org/10.1007/s10661-013-3154-y>.
- Wintle, B.A., Kujala, H., Whitehead, A., et al., 2019. Global synthesis of conservation studies reveals the importance of small habitat patches for biodiversity. *Proc. Natl. Acad. Sci. USA* 116 (3), 909–914. <https://doi.org/10.1073/pnas.1813051115>.
- Wu, R., Tiessen, H., 2002. Effect of land use on soil degradation in alpine grassland soil, China. *Soil Sci. Soc. Am. J.* 66 (5), 1648–1655. <https://doi.org/10.2136/sssaj2002.1648>.
- Xie, X.M., Jian, Y.Z., Wen, X.N., 2009. Spatial and temporal dynamics of the weed community in a seashore *Paspalum* turf. *Weed Sci.* 57 (3), 248–255. <https://doi.org/10.1614/WS-08-124.1>.
- Yamamura, Y., Cheng, J., Yasuda, T., et al., 2021. Livestock-exclusion duration required for restoring grassland in semiarid, loess region in China: estimate based on species composition measured from small-scale vegetation patterns. *Ecol. Res.* 36 (1), 161–176. <https://doi.org/10.1111/1440-1703.12195>.
- Yang, Q., Shi, L.L., Dang, J.J., et al., 2014. Spatial autocorrelation analysis on soil moisture of *Melica przewalskyi* patch in a degraded alpine grassland of Qilian Mountains, Northwest China. *Chin. J. Ecol.* 33 (3), 716–722. <https://doi.org/10.13292/j.1000-4890.2014.0063>.
- You, Q., Xue, X., Peng, F., et al., 2014. Comparison of ecosystem characteristics between degraded and intact alpine meadow in the Qinghai–Tibetan Plateau, China. *Ecol. Eng.* 71, 133–143. <https://doi.org/10.1016/j.ecoleng.2014.07.022>.
- Zhang, J.T., 2005. Two-dimensional pattern analysis for dominant species and community in mountain meadow of Luya Mountain, Shanxi Province, China. *Acta Ecol. Sin.* 25 (6), 1264–1268. <https://doi.org/10.1007/s11515-007-0031-5>.
- Zhang, W.G., Huang, W.B., Yang, Z.Y., 2003. The study on the relationship between mini-patch and degradation of pasture. *Acta Pratacul. Sin.* 12 (3), 44–50.
- Zhao, Y., Mou, X.M., Wei, M., et al., 2021. Effect of vegetation mosaic on spatial heterogeneity of soil organic carbon mineralization and nitrification in an alpine meadow. *Appl. Soil Ecol.* 165, 104007. <https://doi.org/10.1016/j.apsoil.2021.104007>.