

Enhanced soil methane oxidation in both organic layer and topsoil during the succession of subtropical forests

Junhua Liu^{a,b}, Yunjian Xu^{a,b}, Yingmo Zhu^{a,b}, Wen Yin^c, Danhua Fan^d, Guangxuan Yan^e, Syed Turab Raza^{a,b}, Zhiyun Lu^f, Zhe Chen^{a,b,*}

^a Yunnan Key Laboratory of Plant Reproductive Adaptation and Evolutionary Ecology, Institute of Biodiversity, School of Ecology and Environmental Science, Yunnan University, Kunming 650504, Yunnan, China

^b Key Laboratory of Soil Ecology and Health in Universities of Yunnan Province, Yunnan University, Kunming 650500, China

^c Service Center of Science and Technology, Meteorological Bureau of Baoshan, Baoshan 678000, China

^d Meteorological Bureau of Dehong Prefecture, Mangshi 678400, China

^e School of Environment, Henan Normal University, Xinxiang 453007, China

^f Ailaoshan Station of Subtropical Forest Ecosystem Studies, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Jingdong, Yunnan 676209, China

ARTICLE INFO

Keywords:

Methane oxidation
Forest succession
Chronosequence
Nitrogen availability
Methane monooxygenase activity
Subtropical forest

ABSTRACT

Although (sub)tropical forests account for 10–20% of the atmospheric methane (CH₄) uptake by soils, the study of soil CH₄ oxidation rates and the controlling factors during the chronosequences of forest succession remains poorly understood. The objectives of this study were to characterize the vertical distribution patterns and dynamics of CH₄ oxidation among the early-, mid-, and late-successional stages of subtropical forests, and to investigate the main drivers of soil CH₄ fluxes. Three successional forests soils were collected and the ambient and potential CH₄ oxidation rates, the related enzymes as well as the key soil parameters were determined in the laboratory. The soils at mid- and late-successional stages functioned exclusively as a CH₄ sink while the soil at early-successional stage was either a CH₄ source or sink. Soil CH₄ oxidations showed significant vertical distributions along with the successional gradients. The highest rate of ambient CH₄ oxidation was observed in the A-horizon of the mid-successional stage (forest age ~ 100 years), increased by 26-fold compared to the early successional stage, while the highest rate of potential CH₄ oxidation was detected in the O-horizon of the late-successional stages (forest age > 300 years), which increased the CH₄ oxidation by 29% and 21% respectively compared to the early- and mid-successional stages. Soil CH₄ oxidation enhanced with decreasing of soil nitrite and nitrate content but weakened with declining of soil moisture at the successional chronosequence. Collectively, subtropical forests have the potential to increase the soil sink capacity for CH₄ oxidation along the successional gradient, and thereby providing the negative feedbacks to ecosystems under climate changing.

1. Introduction

On the 100-year timescale, the global warming potential of methane (CH₄) is 25 times that of carbon dioxide (CO₂), and CH₄ is the second important greenhouse gas (GHG) in terms of its contribution to climate change, accounting for approximately 20% of the global GHG effect (IPCC, 2013). Over the past 150 years, the methane concentration in atmosphere has increased by 150% (Montzka et al., 2011). The rate of increase in atmospheric methane concentrations is determined by the difference between methane sources and sinks, and currently the terrestrial ecosystem is a net source of atmospheric methane (Tian et al., 2016).

It is well known that well-aerated forest soils are globally important carbon sinks that can mitigate atmospheric methane concentration increase (Basiliako et al., 2009; Chang et al., 2021; Howden et al., 2008). The annual uptake of methane indicating the consumption processes of CH₄ oxidation at both aerobic and anaerobic conditions by global soils ranges from 26 ~ 36 Tg, of which soils account for approximately 10% (Christiansen et al., 2016), and deciduous forests are found to be the strongest sinks of atmospheric CH₄ (Degelmann et al., 2009; Liu and Greaver, 2009; Mazza et al., 2021). Globally, primary forests are at risk of destruction due to human factors (deforestation, man-made fires) and natural factors (such as drought and natural fires). The responses of forests at different succession stages to global changes, such as the

* Correspondence to: Yunnan University, Gewu Building, Kunming 650500, China.

E-mail address: zhechen2019@ynu.edu.cn (Z. Chen).

<https://doi.org/10.1016/j.psep.2022.12.064>

Received 17 September 2022; Received in revised form 4 December 2022; Accepted 21 December 2022

Available online 23 December 2022

0957-5820/© 2022 Institution of Chemical Engineers. Published by Elsevier Ltd. All rights reserved.

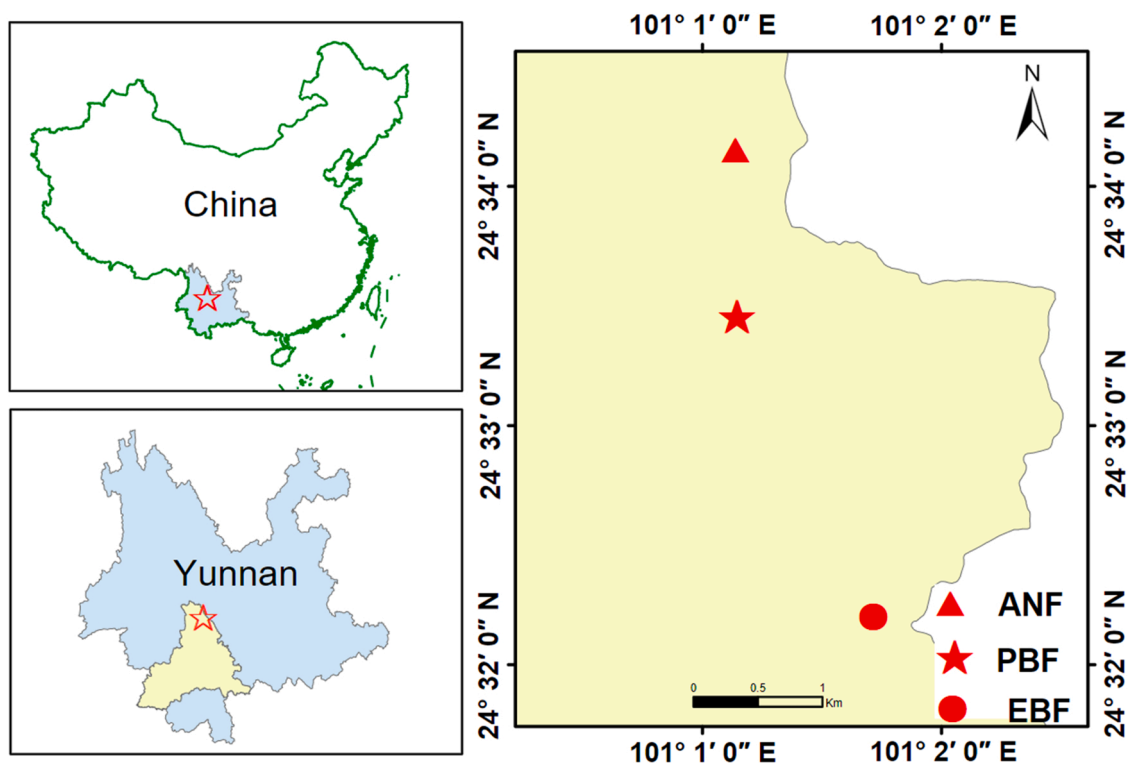


Fig. 1. Location map of the study sites.

oxidation and emission of methane, are still unclear (Tate, 2015). As an important soil biochemical process, methane oxidation is based on the principle that methane is transported to the aerobic area of the soil through the soil pores, and methane oxidizing bacteria can oxidize the surrounding methane into carbon dioxide, thereby effectively reducing methane emissions and alleviating the greenhouse effect (Le Mer and Roger, 2001; Dutaur and Verchot, 2007).

China's forest area is 208 million ha, accounting for 22% of the national area and it contains large methane oxidation potential. The annual uptake of methane by forest soil in China contributed 3.6% of the global methane sink (Cai, 2012), thus the dynamic change of methane oxidation by forest soil is one of the key factors affecting the methane concentration in the atmosphere (Basilikov et al., 2009; Wei et al., 2018). Changes in forest soil CH_4 production and oxidation are affected by various factors such as soil moisture (Sih et al., 2021; Subke et al., 2003; Wang et al., 2013) and soil temperature (Grogan et al., 2004; von Arnold et al., 2005; Zhu et al., 2021) as well as the soil N availability via soil N transformation and plant uptake (Fender et al., 2012; Ren et al., 2022). Soil moisture is a key driver of CH_4 oxidation rates from forest soil because the water-filled porosity limits the transport and diffusion of the oxygen (O_2) and CH_4 in soils (Song et al., 2008; Xiang et al., 2022). As for the temperature effects on CH_4 fluxes, results were inconsistent and tends to be co-affected by the dynamic changes of soil dissolved organic carbon and inorganic nitrogen (Rask et al., 2002; Borken et al., 2006; Liu et al., 2007; Chen et al., 2011; Mills et al., 2013; Li et al., 2021).

Subtropical evergreen broad-leaved forest is the forest with the widest distribution, the largest area and the highest biodiversity in China, accounting for about 1/4 of China's land area (Chen et al., 2019), and this high biodiversity in the subtropics makes it challenging to explore how changes in nutrient status and availability along the forest successional gradients affect the biogeochemical cycling, such as the greenhouse gas emission and uptake (Liu et al., 2020). On both regional and global scale, the rates of primary forest loss and secondary forest generation are increasing in the (sub)tropics (Hansen et al., 2013; Sullivan et al., 2019), creating a requirement for the quantitative assessment of how methane oxidation patterns respond to secondary

succession, and which soil physiochemical factors as well as biotic factors controlling the methane emissions. A recent global meta-analysis showed that primary forest conversion significantly increases soil CH_4 emissions (Han and Zhu, 2020). The methane fluxes from forest soils in successional chronosequences may quite different, due to the shifts in the plant communities, and the age of second forest and the human disturbance and management (Hergoualc'h and Verchot, 2011; Kim and Kirschbaum, 2015), however, the characteristics of methane fluxes or uptake during subtropical forest succession are still unknown.

Similar to other biogeochemical processes, there are vertical distributions in the ability of soils to oxidize CH_4 and this depths difference may affect the CH_4 uptake capacity by soil profiles and is a critical parameter for the CH_4 emission models. Some studies have shown that the highest value of CH_4 oxidation activity in temperate forest soil was located in the subsurface soil (Koschorreck and Conrad, 1993; Plain and Epron, 2021) while other findings illustrated the highest CH_4 oxidation rates was in the surface/topsoil soil (Borken et al., 2003; Sjogersten et al., 2007; Takahashi et al., 2021). For example, Borken et al. (2003) found that the vertical distribution of CH_4 oxidation rates in forest soil varied with the tree species: the highest rate of CH_4 oxidation was located in the subsurface soil of coniferous forest (with spruce and pine as dominant species), but in a broad-leaved forest soil (with beech as the dominant species) the highest value was located in the topsoil. The study by Sjogersten et al. (2007) also showed that the highest methanotrophic activity at a mountain birch forest-tundra system occurred in topsoil. It can be seen that the vertical distribution of CH_4 oxidation in forest soils may vary with climate, biological, soil and other factors. Over the short distances within a subtropical forest, soil abiotic and biotic factors rather than meteorological factors, might control the substantial variations in nutrient supply and the processes of CH_4 production-oxidation. However, compared with the studies of soil methane emissions in boreal and temperate forests, observations for soil methane oxidation in (sub) tropical forests, especially for the forest succession stands and the vertical distribution patterns are still sparse and elusive, implying high uncertainty in estimates of terrestrial ecosystem CH_4 fluxes.

To address the gap of methane oxidations in subtropical forest soils,

including the vertical distribution patterns of the methane oxidation capacity and dynamics during the forest succession, we adopted the method of “space instead of time” and selected three typical succession stages in a national Natural Reserve: the early-successional stage of *Alnus nepalensis* forest (ANF) dominated by *Alnus nepalensis* with the age around 60 years; the mid-successional stage of *Dianshan poplar* forest (PBF) dominated by *Populus bonatii* with the age around 100 years; and late-successional stage of the evergreen broad-leaved forest (EBF) with the age > 300 years and has been well protected from human disturbance (Li et al., 2016). The early- and mid-successional forests developed through natural succession with the gradual invasion of some pioneer broadleaf species. They are all located in the Ailao Mountains, southwest Yunnan Province, China. The objectives of this study were to (i) explore the effects of forest succession on soil CH₄ oxidation in subtropical areas, including the ambient and potential oxidation capacity; (ii) further characterize the forest succession effects in different soil layers; (iii) reveal the main controlling factors for soil CH₄ oxidation during the forest succession. We hypothesized that the primary forest has the highest capacity for CH₄ oxidation due to the increased activity of methane monooxygenase induced by the increased soil organic carbon accumulation, and the CH₄ oxidation capacity faded with the soil depth extending due to the more air-filled pore space resulting in greater CH₄ diffusing into the O-horizon.

2. Materials and methods

2.1. Study area

This experiment field is located in the Ailaoshan Station of Subtropical Forest which belongs to the Chinese Academy of Sciences, and it is also part of the Ailaoshan Nature Reserve (23°35′–24°44′ N, 100°54′–101°30′ E), which is located in Jingdong County, Yunnan Province, southwest China (Fig. 1). The soils of the three successional forests belong to the Yellow-brown soil and all located in the Nature Reserve with no human disturbance and no cultivation. This area is in a subtropical mountain climate which belongs to the monsoon climate zone. The altitude is 2300 ~ 2600 m and the annual average temperature was 11.3 °C and the average temperature of the growing season was 15.7 °C. The annual precipitation was 1947 mm, of which 85% of the precipitation occurred in the rainy season (from May to October). The three representative succession forests were selected and the detailed information of them was as following:

Alnus nepalensis forest (ANF) is a secondary forest after the deforestation and burning of the evergreen broad-leaved forest, which is located in the northwest corner of the Ailao Mountain Ecological Station, with an area of about 100 hm². It is in the early-successional stage and the recovery period is about 60 years. The ANF is considered to be the primary vegetation/stage after disturbance. The tree layer is composed of a single tree species of *Alnus nepalensis*, and the main species of the shrub layer are *Rhododendron delavayi*, *Lyonia ovalifolia* and *Dchroafebr fuga*. This type of forest canopy is sparse, with the canopy closure of around 75%, and the tree height is around 15 m (Li et al., 2016).

Populus bonatii forest (PBF) is also a secondary forest and formed after the deforestation and burning of the evergreen broad-leaved forest and it is a pioneer tree species for plant community succession in this region. The recovery period of PBF is about 100 years and it is in the mid-successional stage. The canopy closure is 80–85% and the tree height is 15–20 m. The PBF area are scattered as small patches in the primary forest of Evergreen broad-leaved forest (PBF). The dominant species in the tree layer are aspen poplar, accompanied by various Fagaceae plants such as *L. hancei*, *Ternstroemia gymnanthera*, and some species of shrubs and herbaceous vegetation (Li et al., 2016).

Evergreen broad-leaved forest (EBF) is more than 300 years old and is the late-successional stage. It is the largest and most complete native vegetation in the Ailao Mountain, accounting for about 78% of the total vegetation area. The vertical stratification of the community is clear, the

tree layer height is 20–25 m, and the canopy closure is 95%. The dominant species are *Lithocarpus xylocarpus*, *L. crassifolius*, *Castanopsis rufescens*, and *Schima noronhae* (Li et al., 2016).

2.2. Experimental design

In each succession forest, eight plots with 5 m × 5 m at least 10 m distances between each other were established randomly as spatial replicates at the field sites (together there were 24 plots in the three succession forest). The layers of soils used in the experiment were collected in 2021 April. The details of the two layer soils were as following: one is the top O horizon with loose and partly decayed organic matter (approximately 5–8 cm, 6–10 cm and 15–20 cm for ANF, PBF and EBF respectively), the other is the A horizon with organic-rich mineral soil of ~10 cm depth below the O horizon. The two layer soils were sampled from the above mentioned eight replicate plots of each succession forest in field sites, respectively. The five-point sampling method was used to collect 5 soil cores by a steel corer from each plot and was mixed by hand to remove the coarse roots a portion of soil was removed from each sample and packed into a 2 mL centrifuge tube, then stored in a – 80 °C freezer for enzyme activity and microorganisms, and the remaining soil was stored at – 4 °C. Before the experiment started, 25 g of soil was added to the 250 mL flask and then the air in the flask was flushed, followed with the flask was capped. Culture experiments were performed at the temperature of 15.7 °C, which was based on the average temperature of the growing season. In order to maintain the soil water content as in situ, we did not artificially adjust the water content, but only weighed and replenished water every day during the incubation to keep the water content consistent with the initial value of sampling.

Two culture experiments with different initial methane concentration were performed separately: a) monitoring the methane oxidation rates of the three succession forest soils at the ambient methane concentration (approx. 1.85 ppm). The atmospheric methane is mainly consumed by a group of methane-oxidizing bacteria (MOB) that using atmospheric methane as their main energy source and sensitive to low concentration of methane (by high affinity methanotrophs). During the culture process, no exogenous methane was added to the flask and the methane oxidation rates were measured once a day under the atmospheric concentration for three days. After each gas sampling, the flask was blow with a fan for 20 min to fully exchange with the air outside. The flask cap was opened and sieved with a Parafilm (air ventilate but no water escape) after each measurement to maintain the methane status as the ambient condition and to avoid moisture loss; b) monitoring the methane oxidation potentials at the high methane concentration in the surroundings (approx. 1000 ppm CH₄, by low affinity methanotrophs), and the potential assays offer two unique advantages. First, it was capable of measuring the maximum functional capacity of MOB communities because the differences between the successions/sites in potential methane oxidation should reflect changes in MOB community or enzyme activity (Sullivan et al., 2013). Second, potential assays could minimize other important constraints that obscure biogeochemical process, i.e., there are anaerobic micro-zones in the topsoil, and 80% of the endogenous high-concentration methane (18–1000 ppm) produced in the anaerobic zone could be consumed by the low-affinity MOB before escaping into the atmosphere (Cai et al., 2016). Therefore, we observed the ambient and potential CH₄ oxidation rates using the methods described by Cai et al. (2016). For the measurement of methane oxidation potentials, the high-concentration methane was injected into the flasks and the methane concentration was adjusted to approximately 1000 ppm as the reference used (Cai et al., 2016). The soil was pre-cultured at 15.7 °C for 3 days to stabilize soil microbial activity, then the air in the bottle was flushed again, and the methane concentration in the bottle was adjusted to 1000 ppm again to start the measurement. The methane concentration in the flask space was measured every 24 h for 10 days and the cap was not opened, because of the two reasons, on one hand, it is necessary to continuously measure the change of methane

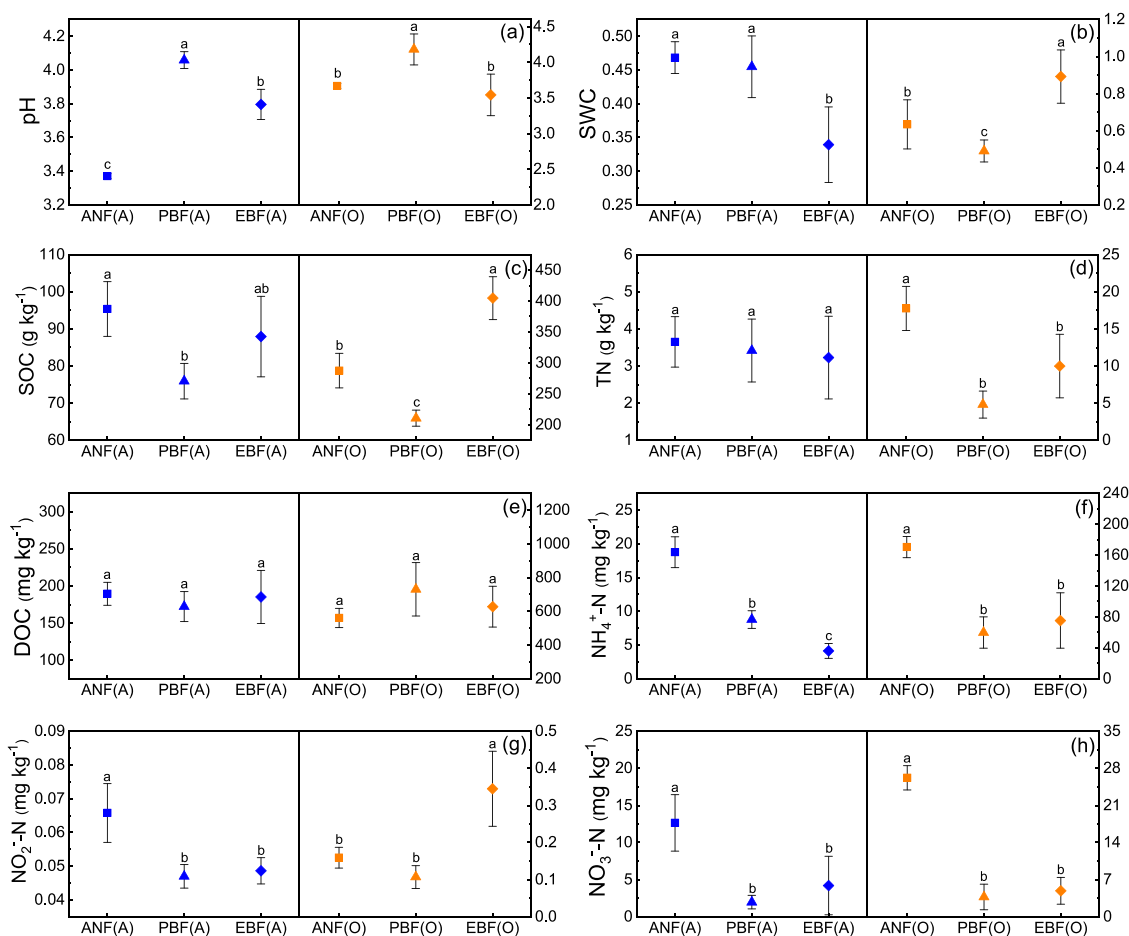


Fig. 2. Soil physicochemical properties in three forest succession stages. Note: Different lowercase letters denote significant differences between the three forests ($P < 0.05$). The blue color represents A-horizon and the orange color represents O-horizon. SWC: soil water content; SOC: soil organic carbon; TN: soil total nitrogen; DOC: dissolved organic carbon. ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

concentration, and on the other hand it is to prevent soil moisture from escaping. For both of the two culture experiment, a 10 mL sample was collected from the headspace by using a 10 mL syringe, then 10 mL of synthetic air (80% N₂, 20% O₂) was injected into the flask to maintain the air pressure in the bottle. The concentration of methane was measured by a gas chromatograph (Shimadzu, Japan, GC-2014).

2.3. Analysis of soil physicochemical properties

The soil pH was measured via a pH-meter under the condition of soil to water ratio of 1:2.5. The nitrate (NO₃⁻-N), nitrite (NO₂⁻-N), ammonium (NH₄⁺-N) and dissolved organic carbon (DOC) were extracted with 1 mol L⁻¹ KCl solution, and their concentrations in extracts were determined by spectrophotometer method in an Intermittent Chemical Analysis (CleverChem380; DeChem-Tech; Germany). Soil samples were digested by sulfuric acid and then the total nitrogen (TN) was determined by the sodium salicylate method. Soil organic carbon (SOC) was measured by converting total organic carbon into CO₂ through high temperature combustion and catalytic oxidation with an organic carbon analyzer (VarioTOC ; Elementar, Germany).

2.4. The CH₄ emission rate

Methane emission rate was calculated with the following formula:

$$F = \frac{M}{V_m} * (VC_2 - VC_1) * \frac{1}{1000m} * \frac{P}{P_0} * \frac{T_0}{T}$$

Where: F is the CH₄ emission (ng (CH₄)•kg⁻¹soil•d⁻¹); M is the molar mass of CH₄ (16 g•mol⁻¹); V_m is the molar volume of 22.4 (L•mol⁻¹); V is the headspace volume (mL); C₁ is the concentration of the gas to be determined (ppm) in the container; C₂ is the concentration of the gas to be determined (ppm) in the container after one day incubation day; m is the mass of dry soil (g); P is 80.735 kpa and P₀ is 101.325 kpa, which represent the atmospheric pressure in Kunming, China (experimental area) and the standard atmospheric pressure, respectively; T₀ is 273.15 K and T is (273.15 + t (°C)) K, which represent the absolute temperature and the temperature controlled in this experiment under the standard conditions, respectively. Detailed information can be found in Yang et al., 2022.

The cumulative CH₄ emission:

$$E = \sum_{i=1}^n \frac{F_i + F_{i+1}}{2} * (t_{i+1} - t_i)$$

E is the cumulative flux of CH₄ (ng CH₄ Kg⁻¹), F is the instantaneous flux rate of CH₄ (ng CH₄ Kg⁻¹ d⁻¹), i is the ith measurement, and (t_{i+1} - t_i) is the time interval between the two measurements (X. Yang et al., 2022; Z. H. Yang et al., 2022).

2.5. Analysis of enzyme activities

The soil methane monooxygenase (MMO) activities were measured by a colormetric plate assay, and the samples for MMO measurements were from the incubation experiments. In brief, soil slurries (soil: solution = 1:100) were buffered with the 50 mM sodium acetate (pH = 4.5),

Table 1

Means of the determined parameters for the A- and O- horizons in different forest succession stages. ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage), with standard error of the mean. Significant soil depth effects at a given forest are indicated by different indices (a, b). Note that forest succession effects are not illustrated in this table, they are showed in Fig. 1.

	ANF		PBF		EBF	
	A-horizon	O-horizon	A-horizon	O-horizon	A-horizon	O-horizon
CH ₄ cumulation (ng kg ⁻¹ dry soil)	-2.92 ± 0.07 a	-2.56 ± 0.52 a	-3.57 ± 0.20 b	-2.49 ± 0.14 a	-3.15 ± 0.17 a	-4.42 ± 0.43 b
pH	3.37 ± 0.01 b	3.67 ± 0.02 a	4.06 ± 0.03 a	4.18 ± 0.14 a	3.80 ± 0.06 a	3.54 ± 0.19 a
SWC (w/w)	46.84 ± 1.57 a	63.44 ± 8.80 a	45.48 ± 3.05 a	49.10 ± 3.94 a	33.94 ± 3.73 b	89.20 ± 9.56 a
NH ₄ ⁺ -N (mg kg ⁻¹)	18.76 ± 1.53 b	170.34 ± 9.16 a	8.77 ± 0.87 b	59.76 ± 13.53 a	4.12 ± 0.73 b	75.31 ± 23.88 a
NO ₃ ⁻ -N (mg kg ⁻¹)	12.65 ± 2.54 b	26.21 ± 1.53 a	1.97 ± 0.61 a	3.74 ± 1.61 a	4.21 ± 1.92 a	4.91 ± 1.68 a
NO ₂ ⁻ -N (mg kg ⁻¹)	0.066 ± 0.001 b	0.16 ± 0.02 a	0.046 ± 0.002 b	0.11 ± 0.02 a	0.049 ± 0.003 b	0.34 ± 0.07 a
TN (g kg ⁻¹)	3.65 ± 0.45 b	17.76 ± 1.98 a	3.41 ± 0.56 a	4.81 ± 1.05 a	3.23 ± 0.74 b	9.99 ± 2.47 a
DOC (mg kg ⁻¹)	189.48 ± 10.28 b	560.43 ± 38.13 a	172.19 ± 13.45 b	730.27 ± 105.39 a	185.26 ± 23.85 b	627.35 ± 80.84 a
SOC (g kg ⁻¹)	95.36 ± 4.92 b	287.42 ± 18.62 a	75.92 ± 3.20 b	210.71 ± 8.66 a	87.92 ± 7.23 b	404.66 ± 23.11 a

which is close to the mean value of soil pH. All sample microplates were incubated in the dark at 25 °C for 3 h to determine the enzymes. After incubation, we used a 96-well microplate reader to determine the quantity of fluorescence for the enzymes, and the detailed procedure was shown in a previous study (Graham et al., 1992).

2.6. Statistical analysis

The data in this study were firstly analyzed for normal distribution by using the Kolmorov-Smirnov test, and then the variance homogeneity was checked by using the Levene test. One-way ANOVA and Fishers Least Significant Difference (LSD) were used to explore the significant differences of forest succession on each parameter of CH₄ oxidation, soil physicochemical properties and MMO activities. The interacting effects of succession stage and time (incubation date) were examined by using the multi-factor variance analysis. The potential correlations between the CH₄ oxidation and soil physicochemical properties and MMO activities were determined by Pearson correlation analysis. These statistical analysis was conducted by using SPSS (version 19, SPSS Inc., Chicago, USA), and data processing and drawing was used by Origin (version 2021, Origin Lab Inc., Northampton, USA).

3. Results

3.1. Soil physicochemical properties

Soil properties differed significantly among forest succession stages (Fig. 2) and A-horizons (Table 1). The differences of soil organic carbon (SOC), total nitrogen (TN), NH₄⁺, NO₃⁻ and pH existed across the three successional stages, and the effects of forest succession and A-horizons on soil pH, SOC and NH₄⁺ were most pronounced (Fig. 2). The SOC content was lower in the early-stage of *Alnus nepalensis* forest (ANF) and mid-stage of *Populus bonatii* forest (PBF) relative to the late-stage of evergreen broad-leaved forest (EBF) at O-horizon (Fig. 2c), indicating that with the developing process from secondary forest to the primary forest, soil carbon content tends to increasing. In contrast, the EBF induced pronounced decrease of NH₄⁺ and NO₃⁻ content in both the O- and A-horizons compared to the early-successional stage of ANF (Figs. 2f, 2h).

The contents of NH₄⁺, NO₃⁻, NO₂⁻, SOC, TN and DOC in the O-horizon were significantly higher than that of the A-horizon soil, and there was a consistent trend in the three successional forests (Table 1). Soil pH was not different between the two layers across all plots at PBF and EBF forests and was fairly consistent acid (ranging from 3.54 to 4.18,

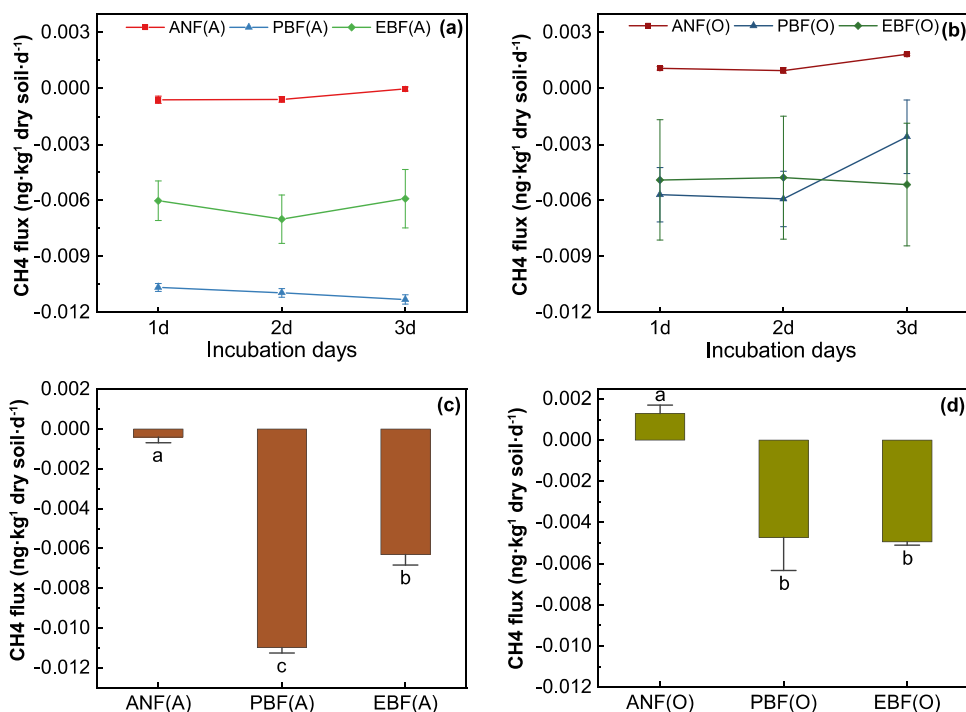


Fig. 3. Effects of forest succession on the rates of ambient CH₄ oxidation (the original CH₄ concentration of 1.85 ppm). Data are standard error ± mean (n = 8). The dynamic changes of the ambient CH₄ oxidation rates in A-horizon (Fig. 3a) and O-horizon (Fig. 3b), and the average rates of CH₄ oxidation in A-horizon (Fig. 3c) and O-horizon (Fig. 3d). Different lowercase letters denote significant differences between the three forests (P < 0.05). ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

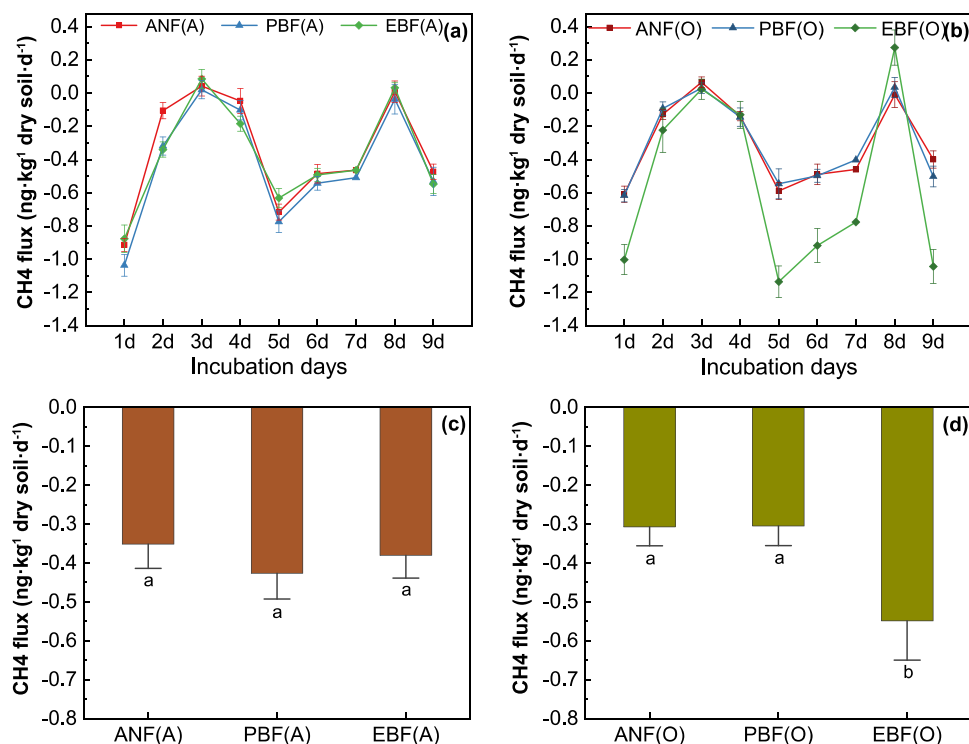


Fig. 4. Effects of forest succession on the rates of potential CH₄ oxidation (the original CH₄ concentration of 1000 ppm). Data are standard error ± mean (n = 8). The dynamic changes of the potential CH₄ oxidation rates in A-horizon (Fig. 4a) and O-horizon (Fig. 4b), and the average rates of CH₄ oxidation in A-horizon (Fig. 4c) and O-horizon (Fig. 4d). Different lowercase letters denote significant differences between the three forests (*P* < 0.05). ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

Table 2
Results of ANOVA testing the effects of the forest succession, incubation time and their interactions on the quantified parameters. Significant effects are indicated in bold.

	Forest succession (ANF vs. PBF vs. EBF)		Time		Forest succession *Time	
	<i>F</i> value	<i>P</i>	<i>F</i> value	<i>P</i>	<i>F</i> value	<i>P</i>
CH ₄ flux in A-horizon	4.024	0.019	103.639	< 0.001	1.026	0.431
CH ₄ flux in O-horizon	34.577	< 0.001	70.064	< 0.001	5.643	< 0.001

Table 1).

3.2. Effects of forest succession on the CH₄ fluxes rates

During the incubation with ambient CH₄ concentration of approx. 1.84 ppm, the methane oxidation rates were monitored in O- and A-horizon soils for the three successional forests. In general, the mid- and late- stages of forests showed significantly stronger CH₄ oxidation than the early stage, and this pattern was found in both O- and A- horizons (Fig. 3). The mean daily rates of CH₄ fluxes in O-horizon were 1.29 ± 0.27, - 4.73 ± 1.07 and - 4.94 ± 0.11 ng kg⁻¹, and in A-horizon were - 0.41 ± 0.19, - 10.97 ± 0.19 and - 6.31 ± 0.35 ng kg⁻¹ for early-, mid-, and late-stages respectively.

During the incubation with high CH₄ concentration of approx. 1000 ppm, the potential rates of methane oxidation were monitored for 9 days, and the CH₄ oxidation rates were about 100 times higher than the rates for ambient CH₄ oxidation (Fig. 4). The mean daily rates of CH₄ fluxes in A-horizon were - 0.351 ± 0.042, - 0.426 ± 0.044 and - 0.380 ± 0.039 ng kg⁻¹ for early-, mid-, and late-stage respectively (Fig. 4c) and there was no significant difference among the three successional forests (Fig. 4c), with the exception that the ANF showed significantly lower CH₄ oxidation than that of the other two forest soils

at 2nd day of incubation (*P* < 0.01, Fig. 4a). As for the O-horizon, the EBF primary forest significantly increased CH₄ oxidation compared with the two secondary forests of ANF and PBF (Figs. 4b, 4d), and the mean CH₄ oxidation rates by EBF was approximately twice that of ANF and PBF (Fig. 4d). Generally, forest succession stages have significant effects on CH₄ oxidation (Table 2, *P* < 0.001) under the high CH₄ concentration surroundings, and the mean daily rates of CH₄ fluxes were - 0.307 ± 0.033, - 0.305 ± 0.034 and - 0.549 ± 0.067 ng kg⁻¹ for early-, mid-, and late-stage respectively (Fig. 4d). On the 1st, 5th, 6th, 7th, and 9th days, the CH₄ oxidation rates were significantly higher in the O-horizon of EBF than that in the secondary forests of ANF and PBF (Fig. 4b).

3.3. Effects of forest succession on the cumulative CH₄ fluxes

Forest succession had a significant impact on the cumulative CH₄ fluxes, for both the ambient and potential rates (Figs. 5, 6). The cumulative ambient CH₄ fluxes of subtropical forest was between 0.00295 and - 0.0402 ng kg⁻¹ after 3 days incubation (Fig. 5) and the cumulative potential CH₄ oxidation was between - 4.50 and - 2.48 ng kg⁻¹ after the 9 days incubation (Fig. 6). As for the A-horizon soil, the cumulative potential CH₄ fluxes in mid stage of PBF was significantly higher than that in the early stage of ANF (-3.57 vs. -2.93 ng kg⁻¹ for PBF and ANF respectively, *P* < 0.01, Figs. 6a, 6d), similar pattern was found for the ambient CH₄ oxidation as showed in Figs. 5a and 5d. As for the O-horizon, the cumulative potential CH₄ fluxes of the three succession stages was between - 5.88 and - 1.94 ng kg⁻¹, and the CH₄ fluxes in the primary forest EBF was significantly lower than that of the two secondary forests (ANF -2.56, PBF -2.49 and EBF -4.42 ng kg⁻¹, Fig. 6e). When accumulating the amount of potential CH₄ oxidation in the O and the A layer, the CH₄ fluxes in EBF was also significantly lower than that of ANF and PBF (Figs. 6e, 6f).

Depth effects on the cumulative methane fluxes in the same succession stage were performed. The results showed that in the early stage of ANF, there was no significant difference of the cumulative potential CH₄ oxidation between the A- and O- horizons (-2.92 vs. -2.56 ng kg⁻¹ for A- and O-horizon respectively, *P* > 0.05, Table 1). However, in the mid stage of PBF, the cumulative potential CH₄ oxidation was significantly

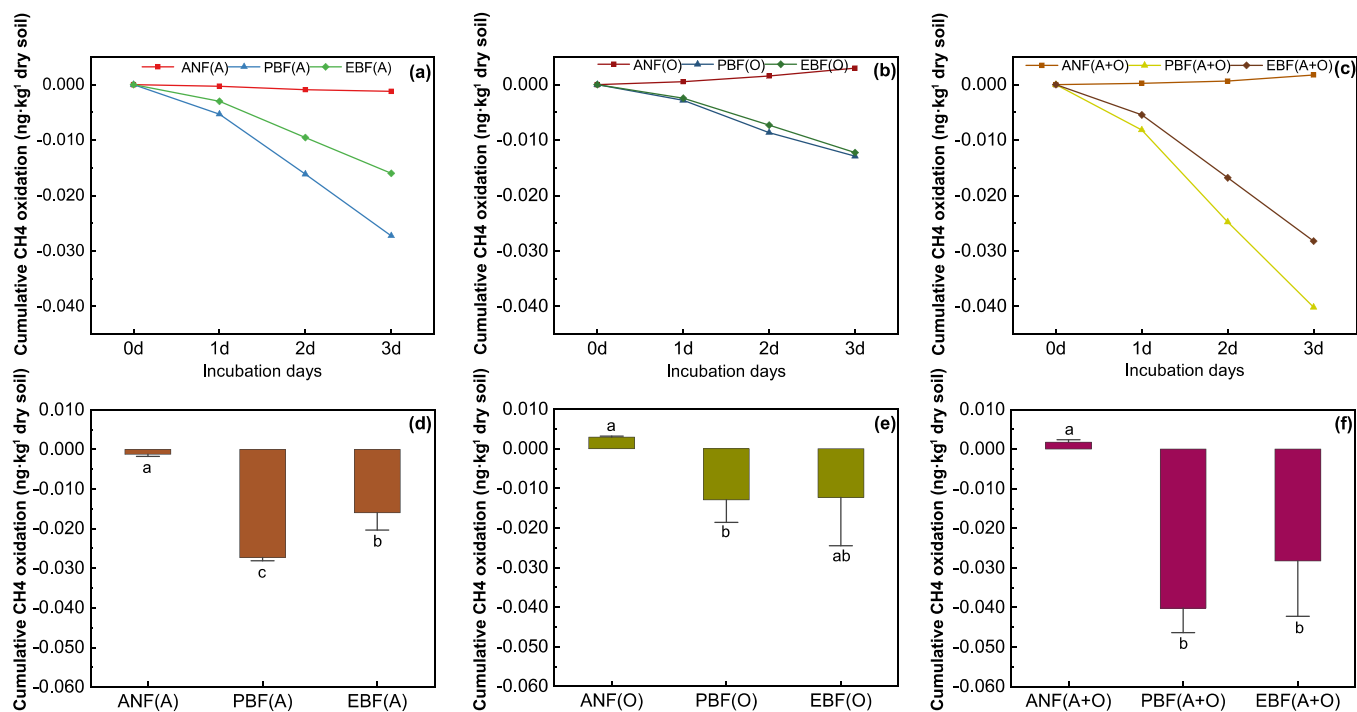


Fig. 5. Effects of forest succession on the cumulative CH₄ oxidation at the original CH₄ concentration of 1.85 ppm. Data are standard error ± mean (n = 8). The dynamic changes of the cumulative ambient CH₄ oxidation in A-horizon (Fig. 5a), O-horizon (Fig. 5b) and the combined horizons of A+O (Fig. 5c), and the average of the cumulative ambient CH₄ oxidation rates in A-horizon (Fig. 5d), O-horizon (Fig. 5e) and the combined horizons of A+O (5 f). Different lowercase letters denote significant differences between the three forests (P < 0.05). ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

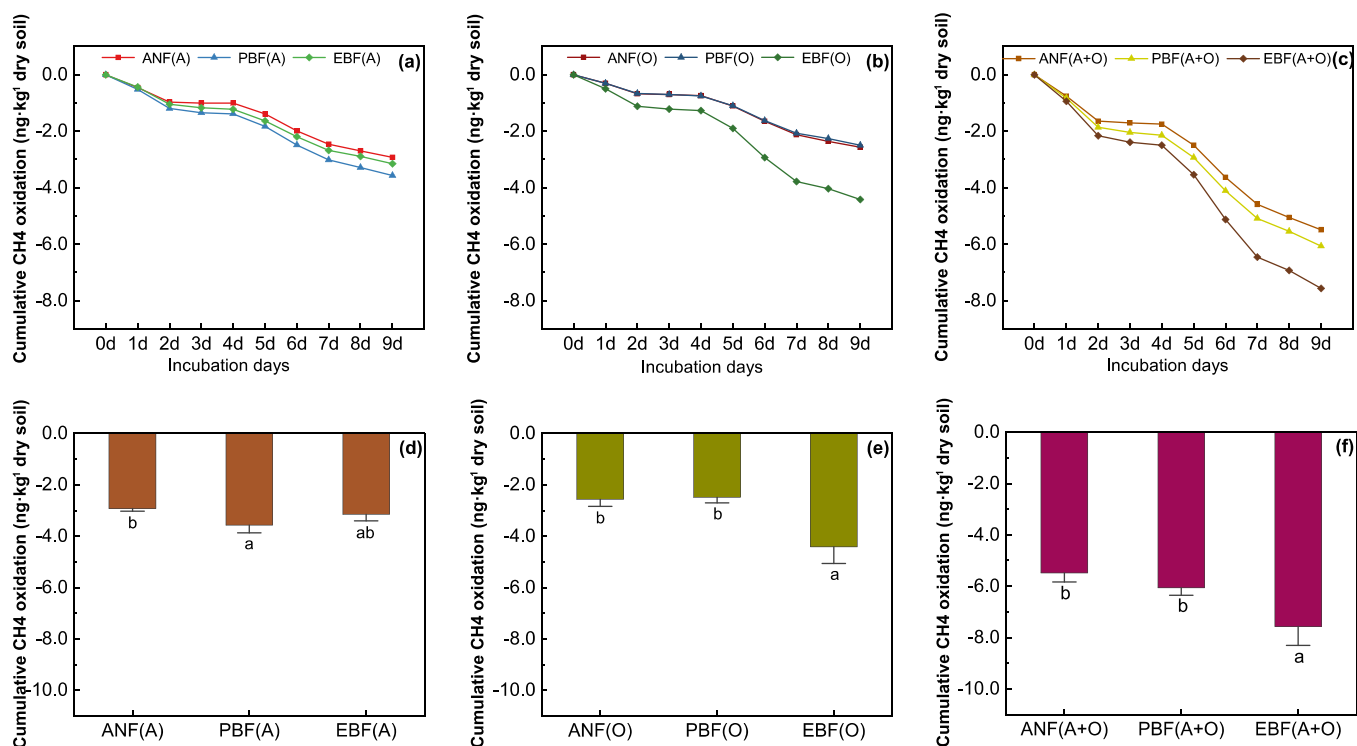


Fig. 6. Effects of forest succession on the cumulative CH₄ oxidation at the original CH₄ concentration of 1000 ppm. Data are standard error ± mean (n = 8). The dynamic changes of the cumulative potential CH₄ oxidation in A-horizon (Fig. 5a), O-horizon (Fig. 5b) and the combined horizons of A+O (Fig. 5c), and the average of the cumulative potential CH₄ oxidation rates in A-horizon (Fig. 5d), O-horizon (Fig. 5e) and the combined horizons of A+O (5 f). Different lowercase letters denote significant differences between the three forests (P < 0.05). ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

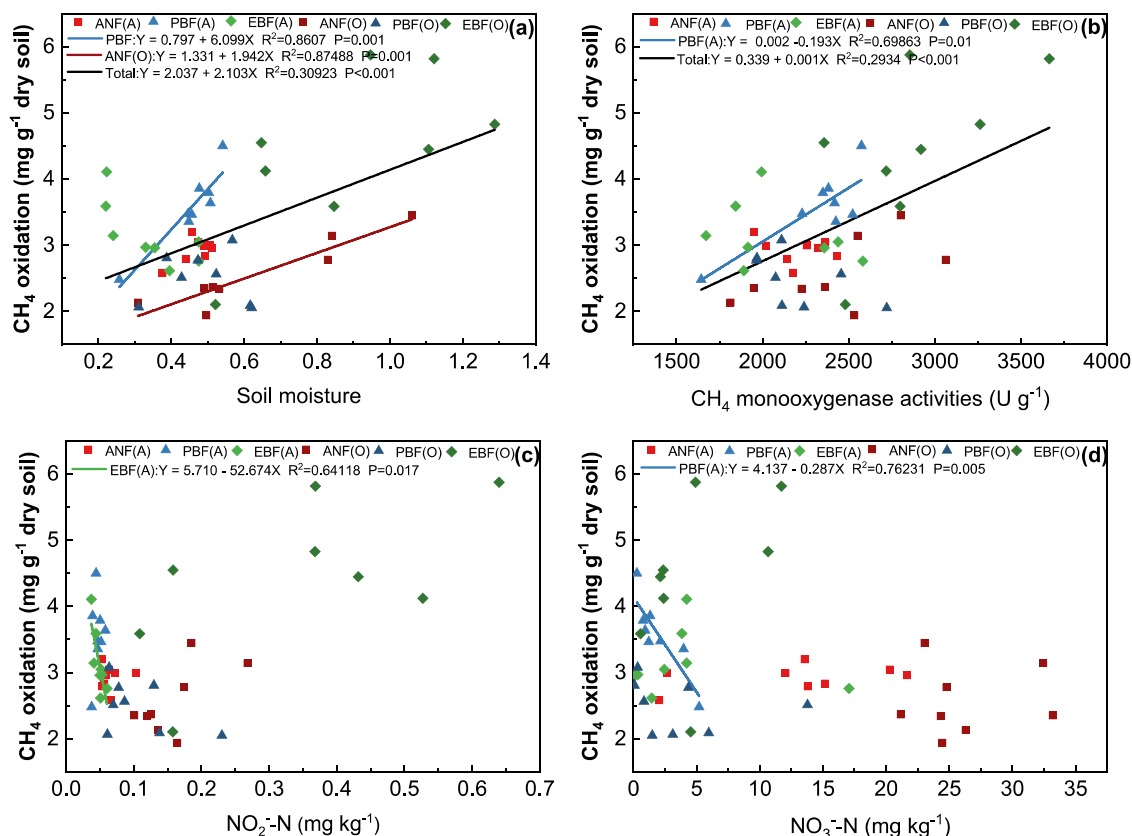


Fig. 7. Relationships between the cumulative CH₄ oxidation and (a) soil moisture, (b) soil methane monooxygenase activities, (c) nitrite content and (d) nitrate content. Regression analyses were either based on the entire dataset from both O- and A-horizons of the three forests of ANF (*Alnus nepalensis* forest, early-stage), PBF (*Populus bonatii* forest, mid-stage) and EBF (Evergreen broad-leaved forest, late-stage), or conducted separately for the three forests and two horizons.

Table 3

Multiple stepwise regression analysis of the dependency of CH₄ oxidation in different forest succession stages to potential control factors.

Explained variable		Regression equation	r ²	P	Factors excluded
CH ₄ oxidation	PBF(A-horizon)	$Y = 1.692 - 31.717NO_2 + 7.409SM$	0.956	< 0.001	NO ₃ , MMO
	EBF(A-horizon)	$Y = 5.71 - 52.674NO_2$	0.641	0.017	SM, NO ₃ , MMO
	ANF(O-horizon)	$Y = 1.331 + 1.942SM$	0.875	0.001	NO ₃ , NO ₂
	ANF(A+O horizons)	$Y = 2.062 + 2.304SM - 5.229NO_2$	0.577	0.004	NO ₃ , MMO
	PBF(A+O horizons)	$Y = 0.764 - 10.209NO_2 + 0.001MMO$	0.602	0.003	SM, NO ₃
	EBF(A+O horizons)	$Y = 2.969 - 4.137NO_2$	0.562	0.001	NO ₃ , MMO, SM
	Total	$Y = 2.037 + 2.103SM$	0.309	< 0.001	NO ₃ , NO ₂ , MMO

NO₂: soil nitrite content (mg N kg⁻¹ dry soil); NO₃: soil nitrate content (mg N kg⁻¹ dry soil); SM: soil moisture; MMO: soil methane monooxygenase activities (U g⁻¹).

lower in the A-horizon than that in the O-horizon (−3.57 vs. −2.49 ng kg⁻¹ for A- and O-horizon respectively, $P < 0.05$, Table 1); while contrast result was found in the late-stage of EBF with the higher cumulative potential CH₄ fluxes in the O-layer (−3.15 vs. −4.42 ng kg⁻¹ for A- and O- respectively, $P < 0.05$, Table 1).

3.4. Relationships between the potential CH₄ oxidation and soil properties

In general, the cumulative potential CH₄ oxidation in different succession stages was positively correlated with soil moisture ($R^2 = 0.860$, $P < 0.01$, Fig. 7a) and MMO activity ($R^2 = 0.699$, $P < 0.05$, Fig. 7b). Likewise, in some given forest succession stage, we obtained similar results. For example, in the ANF O-horizon and the PBF A-horizon, the amount of CH₄ oxidation was positively correlated with soil moisture (Fig. 7a); the amount of CH₄ oxidation in the PBF A-horizon was positively correlated with the MMO activity (Fig. 7b). In contrast, the cumulative CH₄ oxidation was negatively correlated with soil nitrite and nitrate content but only within the specific successional stages (Figs. 7c, 7d).

Stepwise regression analysis was used to identify the most important variables controlling CH₄ oxidation at different succession stages. In the early stage of succession (ANF), CH₄ oxidation was mainly controlled by soil moisture and nitrite content, which together explained 57.7% of the CH₄ oxidation (O- and A- horizons), while if only the O-horizon was considered, moisture explained 87.5% of the CH₄ oxidation (Table 3). In the mid stage of PBF, the results were similar, i.e., CH₄ oxidation at A horizon was mainly affected by soil moisture and nitrite content, which together explained 95.6% of CH₄ oxidation, while if both the two horizons were conducted the CH₄ oxidation was mainly affected by the MMO activity and nitrite content (Table 3). At the late stage of EBF, CH₄ oxidation was mainly affected by soil nitrite content, and the explained ability was 56.2–64.1%. Overall, soil moisture was the main factor controlling the subtropical forest CH₄ oxidation ($R^2 = 0.309$, $P < 0.001$, Table 3).

4. Discussion

In this study, experiments on ambient and potential methane

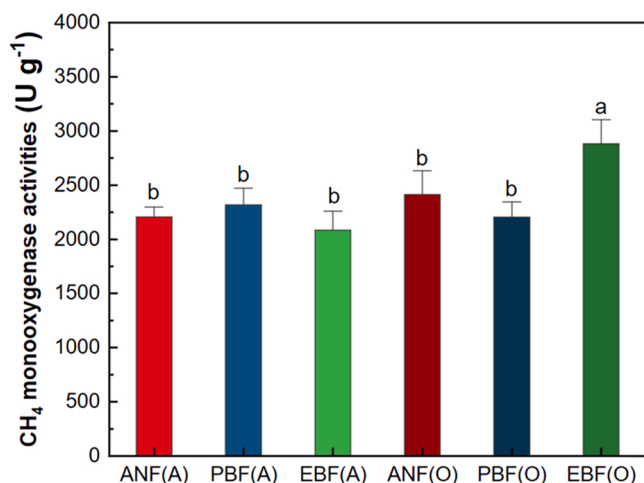


Fig. 8. Soil methane monoxygenase activities in O- and A-horizons of the three forest succession stages. Note: Different lowercase letters denote significant differences between the three forests and two layers ($P < 0.05$). ANF: *Alnus nepalensis* forest (early-stage); PBF: *Populus bonatii* forest (mid-stage), EBF: Evergreen broad-leaved forest (late-stage).

oxidation were conducted by collecting O- and A-horizons from the three successional stages of subtropical forest. We found that O- and A-horizon soils in different successional stages varied in the methane oxidation capacity. For the A- and O-horizons in general, the potential methane oxidation capacity increased with the forest maturity/succession, especially in the O-horizons, where the trend was more pronounced (Fig. 6 f). Our results are consistent with previous reports, for example, the study by Zhou (et al. (2019) illustrated that soil methane oxidation accelerated with the stand age in a secondary subtropical forest of southwest China. In soils of eastern Amazonia tropical forest, the annual CH₄ oxidation in the primary forest was two times higher than that in the secondary forest (2.1 vs 1.0 kg CH₄ ha⁻¹ y⁻¹ for primary and secondary forest respectively) (Verchot et al., 2000). However, there is a study showed that CH₄ oxidation in the oak forest enhanced with stand age but declined with age for soils under Norway spruce (Christiansen and Gundersen, 2011). In the UK Gisburn forest, observations from the shallow organic soils showed that stand age had no significant effect on soil CH₄ oxidation (McNamara et al., 2008), indicating that the effects of stand age on CH₄ oxidation is not always consistent and still is uncertain (Tate, 2015). In the present study, the

variations of soil CH₄ oxidation in the three forest succession stages were found to be related with the soil moisture. Many studies showed that forest soil CH₄ oxidation was negatively correlated with soil moisture, because high moisture states limit the diffusion of atmospheric CH₄ at the soil-atmosphere interface, which led to a decrease in the substrate concentration of methanotrophs in the soil, and accordingly the methane oxidation rates decreased (Bhardwaj and Dubey, 2020; Feng et al., 2020). However, our results were opposite and may be due to the moisture status at the time of sampling. During the dry seasons in subtropical forest as we sampled, soil microbial activity was limited by the relatively low moisture condition (the gravimetric moisture: 30 ~ 65% at most plots), while the high canopy density of the primary forest was not conducive to the evaporation of soil water compared with the secondary forests, and thus the relatively wet conditions in the primary forest during the dry seasons was beneficial to the soil microbial activity, including the activity of methanotrophs (Knief, 2019). This inference was supported by the results of soil respiration as shown in Fig S1, with the similar pattern of cumulative CO₂ fluxes compared to the patterns of the cumulative CH₄ oxidation (Fig S1). It should be noted that our sampling this time was the dry season and if in the rainy season, when the soil moisture was largely increased, the more anaerobic microsites might be created, which was conducive to the production of methane by methanogenesis. Another explanation might be due to the differences between the intact and cultured soils. We did not use the intact soil although we did not sieve the soil after taking back to the laboratory to avoid disturbance, the overall structure of the soil may still be affected, especially the change of porosity, which affects the use of water and the oxidation of methane by soil microorganisms.

In the organic layer, we found that the late-stage of EBF had the strongest potential CH₄ oxidative ability, which was significantly higher than that of the early- and mid-successional stages (Figs. 4b and 4d, Fig. 6). It has been reported that the presence of humus in organic layer reduces the rate of soil methane oxidation (Dong et al., 1998; Saari et al., 1998), but we observed significant methane oxidation in the organic (humus) layers at all three successional stages, indicating that the subtropical forest O-horizon also acts as a methane sink. The study by Dong et al. (1998) and Saari et al. (1998) was conducted in coniferous forests, where the organic compounds of monoterpenes in humic substances can inhibit the oxidation of CH₄, while the inhibitory effect of this organic matter may not exist in broad-leaved forests (Tate, 2015). The methane oxidation of O-horizon in EBF was stronger than that of the other two forests O-horizon (Fig. 4d), which could be due to the higher enzyme activities of methane monoxygenase in the EBF O-horizon (Fig. 8), promoting the CH₄ oxidation.

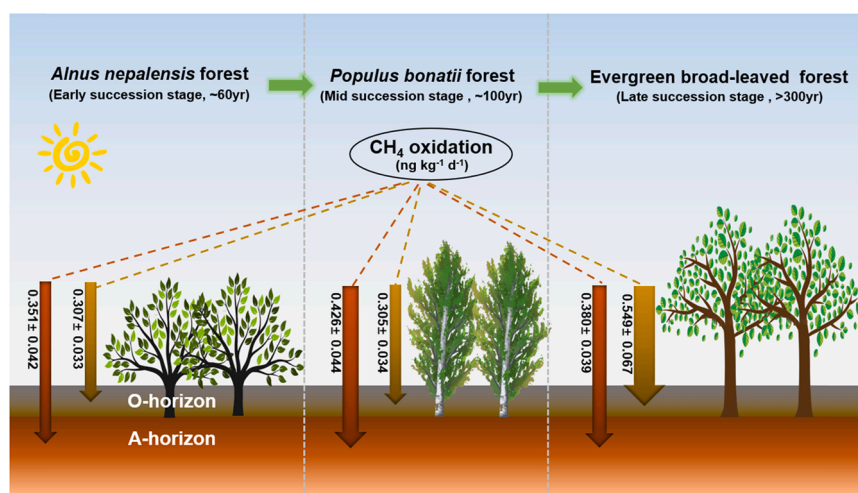


Fig. 9. Scheme of the CH₄ oxidation capacity in O- and A-horizons under the successional stages of subtropical forests. Thickness of the arrows is representative for the respective CH₄ oxidation rates.

In addition to the controlling factor of soil moisture, the higher CH₄ oxidation rate occurring in the late-successional stage of EBF might be attributed to the declines of soil inorganic N (NH₄⁺, NO₂⁻ and NO₃⁻) availabilities in comparison to the early-stage of ANF and mid-stage of PBF. In the early stage of forest succession, the decline of plant communities and the simplification of species will reduce the oxidation of inorganic N by plants, resulting in the increase of soil inorganic nitrogen (Templer et al., 2008). With the development of vegetation succession and the restoration of plant diversity, more soil inorganic nitrogen is retained by the plant and thus causing its decrease (Zhang et al., 2013). Our results of regression analysis revealed that soil NO₂⁻ and NO₃⁻ rather than NH₄⁺ was negatively correlated to the CH₄ oxidation across the forest successions (Table 3, Fig. 7), indicating that NO₂⁻ and NO₃⁻ production by nitrification was strong, especially at the early stage of ANF forest and might inhibit the CH₄ oxidation. In addition, the transformed NO₃⁻ and NO₂⁻ produced via NO₃⁻ reduction are probably toxic to CH₄-oxidizing bacteria (Conrad, 2009). Similarly, previous studies reported that high NO₃⁻ concentrations has been found as a strong inhibition of CH₄ oxidation in unsaturated forest soils, and this inhibition effects varied with the vegetation types (Fender et al., 2012; Jang et al., 2011; Mochizuki et al., 2012). Our results, together with those from other reports, suggest that the decrease in soil NO₂⁻ and NO₃⁻ contents may be as one of the key mechanisms that regulate the increases in soil CH₄ oxidation along the successional gradients of subtropical forests. (Fig. 9).

Beside the above mentioned factors, the increased plant diversity with the forest succession and associated changes in monoterpane release can also affect changes in soil CH₄ oxidation (Dubois et al., 2018; Song et al., 2022; Tate, 2015; Zhu et al., 2021). In the process of forest succession, the role of soil nutrient limitation is strengthened with the development of aboveground vegetation (Li et al 2013; Xu et al., 2022; Yang et al., 2022), which promotes the distribution of plant photosynthesis products to roots to facilitate the absorption of nutrients, resulting in an increase in plant root biomass and exudates (Zhang et al., 2013; Prescott et al., 2020; Maarastawi et al., 2018), thereby increasing the soil microbial biomass and enzymatic activities, including the number of methanotrophs (Wu et al., 2021; Wang et al., 2022) and the shift between oligotrophic and copiotrophic methanotrophic communities (Dorr et al., 2010;). In the same study area, the previous studies showed that the existing amount and carbon and nitrogen components of litter in the three forest succession stages changed significantly, which affected the community characteristics of soil arthropods and nematodes (Zhao et al., 2011, Li et al., 2016). Together with our study, we inferred that the vegetation and litter composition shifts along the forest succession might change nutrient status and thus drive the methanotrophs communities and patterns of CH₄ oxidation. The powerful molecular techniques in the past decade have provided new insights into soil CH₄ emission mechanisms and the characteristics and population functions of Type I and Type II methanotrophs (Tate, 2015; Knief et al., 2015;). In this study, we also attempts to use the quantitative PCR and high-throughput sequencing technologies to reveal the abundance, diversity and gene expression of methanotrophs, but unfortunately we failed to amplify Type I and Type II methanotrophs based on published primers (Cai et al., 2016b). It might be related to the strong inhibition of DNA polymerase in the PCR by high humic acid in forest humus and soil although we have tried several methods for DNA extraction and purification. The researches on soil methanotrophs and methanogenesis were mostly focused on paddy fields and wetlands (Jiang et al., 2019), and the studies on the CH₄-related gene expression in forests was rare, but this is critical to reveal the CH₄ emission mechanisms, and thus it need to be considered in the future.

5. Conclusions

Our study showed the exclusively CH₄ sink across the early-, mid-, and late stages of forest succession and the significant vertical

distributions of CH₄ oxidation. The highest oxidation rate was observed in the organic horizon of the late-successional stages, which increased the CH₄ oxidation by 29% and 21% respectively compared with the early- and mid-successional stages. Furthermore, our study revealed that soil moisture and methane monooxygenase activity were the facilitator of soil CH₄ oxidation while soil nitrite and nitrate availability probably inhibit the CH₄ oxidation along forest successional gradients. Since primary and secondary forests are the main types of forest cover in the global and subtropical regions, our study indicating that the vegetation shifts along the forest succession might change the litterfall composition and nutrient status and consequently drive the patterns of CH₄ oxidation. Understanding the CH₄ oxidation dynamics and the controlling factors in the subtropical forest succession is essential to improve our knowledge of their role in the global carbon cycle and to mitigate and adapt to climate change.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (32260323), Yunnan Science and Technology Planning Project (202002AE090010) and the Major Program for Basic Research Project of Yunnan Province (2018BC007, 202101BC070002). This work was jointly supported by the Ailaoshan Station of Subtropical Forest Ecosystem Studies.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.psep.2022.12.064.

References

- von Arnold, K., Nilsson, M., Hanell, B., Weslien, P., Klemmedtsson, L., 2005. Fluxes of CO₂, CH₄ and N₂O from drained organic soils in deciduous forests. *Soil Biol. Biochem.* 37, 1059–1071. <https://doi.org/10.1016/j.soilbio.2004.11.004>.
- Basiliko, N., Khan, A., Prescott, C.E., Roy, R., Grayston, S.J., 2009. Soil greenhouse gas and nutrient dynamics in fertilized western Canadian plantation forests. *Can. J. For. Res.* 39, 1220–1235. <https://doi.org/10.1139/x09-043>.
- Bhardwaj, Y., Dubey, S.K., 2020. Changes in pmoA gene containing methanotrophic population and methane oxidation potential of dry deciduous tropical forest soils. *Curr. Sci.* 118, 750–758. <https://doi.org/10.18520/cs/v118/i5/750-758>.
- Borken, W., Xu, Y.J., Beese, F., 2003. Conversion of hardwood forests to spruce and pine plantations strongly reduced soil methane sink in Germany. *Glob. Change Biol.* 9, 956–966. <https://doi.org/10.1046/j.1365-2486.2003.00631.x>.
- Borken, W., Davidson, E.A., Savage, K., Sundquist, E.T., Steudler, P., 2006. Effect of summer throughfall exclusion, summer drought, and winter snow cover on methane fluxes in a temperate forest soil. *Soil Biol. Biochem.* 38, 1388–1395. <https://doi.org/10.1016/j.soilbio.2005.10.011>.
- Cai, Y., Zheng, Y., Bodelier, P.L.E., Conrad, R., Jia, Z., 2016. Conventional methanotrophs are responsible for atmospheric methane oxidation in paddy soils. *Nat. Commun.* 7 <https://doi.org/10.1038/ncomms11728>.
- Cai, Z., 2012. Greenhouse gas budget for terrestrial ecosystems in China. *Sci. China-Earth Sci.* 55, 173–182. <https://doi.org/10.1007/s11430-011-4309-8>.
- Chang, R., Liu, X., Wang, T., Li, N., Bing, H., 2021. Stimulated or inhibited response of methane flux to nitrogen addition depends on nitrogen levels. *J. Geophys. Res. -Biogeosci.* 126. <https://doi.org/10.1029/2021jg006600>.
- Chen, L., Swenson, N.G., Ji, N.N., Mi, X.C., Ren, H.B., Guo, L.D., Ma, K.P., 2019. Differential soil fungus accumulation and density dependence of trees in a subtropical forest. *Science* 366, 124. <https://doi.org/10.1126/science.aau1361>.
- Chen, W., Wolf, B., Zheng, X., Yao, Z., Butterbach-Bahl, K., Brüggemann, N., Liu, C., Han, S., Han, X., 2011. Annual methane uptake by temperate semiarid steppes as regulated by stocking rates, aboveground plant biomass and topsoil air permeability. *Glob. Change Biol. Bioenergy* 17, 2803–2816. <https://doi.org/10.1111/j.1365-2486.2011.02444.x>.
- Christiansen, J.R., Gundersen, P., 2011. Stand age and tree species affect N₂O and CH₄ exchange from afforested soils. *Biogeosciences* 8, 2535–2546. <https://doi.org/10.5194/bg-8-2535-2011>.

- Christiansen, J.R., Levy-Booth, D., Prescott, C.E., Grayston, S.J., 2016. Microbial and environmental controls of methane fluxes along a soil moisture Gradient in a Pacific coastal temperate rainforest. *Ecosystems* 19, 1255–1270. <https://doi.org/10.1007/s10021-016-0003-1>.
- Conrad, R., 2009. The global methane cycle: recent advances in understanding the microbial processes involved. *Environ. Microbiol. Rep.* 1, 285–292. <https://doi.org/10.1111/j.1758-2229.2009.00038.x>.
- Degelmann, D.M., Borken, W., Kolb, S., 2009. Methane oxidation kinetics differ in European beech and Norway spruce soils. *Eur. J. Soil Sci.* 60, 499–506. <https://doi.org/10.1111/j.1365-2389.2009.01138.x>.
- Dong, Y., Scharffe, D., Lobert, J.M., Crutzen, P.J., Sanhueza, E., 1998. Fluxes of CO₂, CH₄ and N₂O from a temperate forest soil: the effects of leaves and humus layers. *Tellus B Chem Phys Meteorol.* 50, 243–252. <https://doi.org/10.1034/j.1600-0889.1998.t012-00003.x>.
- Dorr, N., Glaser, B., Kolb, S., 2010. Methanotrophic communities in Brazilian ferralsols from naturally forested, afforested, and agricultural sites. *Appl. Environ. Microbiol.* 76, 1307–1310. <https://doi.org/10.1128/aem.02282-09>.
- Dubois, M., Van den Broeck, L., Inze, D., 2018. The pivotal role of ethylene in plant growth. *Trends Plant Sci.* 23, 311–323. <https://doi.org/10.1016/j.tplants.2018.01.003>.
- Dutaur, L., Verchot, L.V., 2007. A global inventory of the soil CH₄ sink. *Glob. Biogeochem. Cycles* 21. <https://doi.org/10.1029/2006gb002734>.
- Fender, A.C., Pfeiffer, B., Gansert, D., Leuschner, C., Daniel, R., Jungkunst, H.F., 2012. The inhibiting effect of nitrate fertilisation on methane uptake of a temperate forest soil is influenced by labile carbon. *Biol. Fertil. Soils* 48, 621–631. <https://doi.org/10.1007/s00374-011-0660-3>.
- Feng, H., Guo, J., Han, M., Wang, W., Peng, C., Jin, J., Song, X., Yu, S., 2020. A review of the mechanisms and controlling factors of methane dynamics in forest ecosystems. *For. Ecol. Manag.* 455. <https://doi.org/10.1016/j.foreco.2019.117702>.
- Graham, D.W., Korich, D.G., Leblanc, R.P., Sinclair, N.A., Arnold, R.G., 1992. Applications of a colorimetric plate assay for soluble methane monooxygenase activity. *Appl. Environ. Microbiol.* 58, 2231–2236. <https://doi.org/10.1128/aem.58.7.2231-2236.1992>.
- Grogan, P., Michelsen, A., Ambus, P., Jonasson, S., 2004. Freeze-thaw regime effects on carbon and nitrogen dynamics in sub-arctic heath tundra mesocosms. *Soil Biol. Biochem.* 36, 641–654. <https://doi.org/10.1016/j.soilbio.2003.12.007>.
- Han, M., Zhu, B., 2020. Changes in soil greenhouse gas fluxes by land use change from primary forest. *Glob. Change Biol.* 26, 2656–2667. <https://doi.org/10.1111/gcb.14993>.
- Hansen, M.C., Potapov, P.V., Moore, R., Hancher, M., Turubanova, S.A., Tyukavina, A., Thau, D., Stehman, S.V., Goetz, S.J., Loveland, T.R., Kommareddy, A., Egorov, A., Chini, L., Justice, C.O., Townshend, J.R.G., 2013. High-Resolution Global Maps of 21st-Century Forest Cover Change. *Science* 342, 850–853. <https://doi.org/10.1126/science.1244693>.
- Hergoualc'h, K., Verchot, L.V., 2011. Stocks and fluxes of carbon associated with land use change in Southeast Asian tropical peatlands: A review. *Glob. Biogeochem. Cycles* 25. <https://doi.org/10.1029/2009gb003718>.
- Howden, S.M., Crimp, S.J., Stokes, C.J., 2008. Climate change and Australian livestock systems: impacts, research and policy issues. *Aust. J. Exp. Agric.* 48, 780–788. <https://doi.org/10.1071/ea08033>.
- IPCC, 2013. The final draft Report, dated 7 June 2013, of the Working Group I contribution to the IPCC 5th Assessment Report. In: *Climate Change 2013: the Physical Science Basis*.
- Jang, I., Lee, S., Zoh, K.D., Kang, H., 2011. Methane concentrations and methanotrophic community structure influence the response of soil methane oxidation to nitrogen content in a temperate forest. *Soil Biol. Biochem.* 43, 620–627. <https://doi.org/10.1016/j.soilbio.2010.11.032>.
- Jiang, Y., Qian, H.Y., Huang, S., Zhang, X.Y., Wang, L., Zhang, L., Shen, M.X., Xiao, X.P., Chen, F., Zhang, H.L., Lu, C.Y., Li, C., Zhang, J., Deng, A.X., van Groenigen, K.J., Zhang, W.J., 2019. Acclimation of methane emissions from rice paddy fields to straw addition. *Sci. Adv.* 5. <https://doi.org/10.1126/sciadv.aau9038>.
- Kim, D.-G., Kirschbaum, M.U.F., 2015. The effect of land-use change on the net exchange rates of greenhouse gases: a compilation of estimates. *Agric. Ecosyst. Environ.* 208, 114–126. <https://doi.org/10.1016/j.agee.2015.04.026>.
- Knief, C., 2015. Diversity and habitat preferences of cultivated and uncultivated aerobic methanotrophic bacteria evaluated based on pmoa as molecular marker. *Front. Microbiol.* 6. <https://doi.org/10.3389/fmicb.2015.01346>.
- Knief, C., 2019. Diversity of methane-cycling microorganisms in soils and their relation to oxygen. *Curr. Issues Mol. Biol.* 33, 23–56. <https://doi.org/10.21775/cimb.033.023>.
- Koschorreck, M., Conrad, R., 1993. Oxidation of atmospheric methane in soil - measurements in the field, in soil cores and in soil samples. *Glob. Biogeochem. Cycles* 7, 109–121. <https://doi.org/10.1029/92gb02814>.
- Le Mer, J., Roger, P., 2001. Production, oxidation, emission and consumption of methane by soils: A review. *Eur. J. Soil Biol.* 37, 25–50. [https://doi.org/10.1016/s1164-5563\(01\)01067-6](https://doi.org/10.1016/s1164-5563(01)01067-6).
- Li, P., Yang, Y.H., Fang, J.Y., 2013. Variations of root and heterotrophic respiration along environmental gradients in China forests. *J. Plant Ecol.* 6, 358–367. <https://doi.org/10.1093/jpe/rtt009>.
- Li, X.Y., Min, X.B., Hu, X.X., Jiang, Z., Li, C.F., Yang, W.C., Zhao, F.P., 2021. In-situ synthesis of highly dispersed Cu-CuxO nanoparticles on porous carbon for the enhanced persulfate activation for phenol degradation. *Sep. Purif. Technol.* 276. <https://doi.org/10.1016/j.seppur.2021.119260>.
- Li, Z., Wei, Z., Yang, X., 2016. Seasonal variations of soil nematode community at different secondary succession stages of evergreen broad-leaved forests in Ailao Mountain. *Chinese J. Ecol.* 35, 3023–3031.
- Li, Z.P., Wei, Z.F., Yang, X.D., 2016. Seasonal variations of soil nematode community at different secondary succession stages of evergreen broad-leaved forests in Ailao Mountain. *Chin. J. Ecol.* 35 (11), 3023–3031. <https://doi.org/10.13292/j.1000-4890.201611.008>.
- Liu, C., Holst, J., Brueggemann, N., Butterbach-Bahl, K., Yao, Z., Yue, J., Han, S., Han, X., Krummelbein, J., Horn, R., Zheng, X., 2007. Winter-grazing reduces methane uptake by soils of a typical semi-arid steppe in Inner Mongolia, China. *Atmos. Environ.* 41, 5948–5958. <https://doi.org/10.1016/j.atmosenv.2007.03.017>.
- Liu, L., Greaver, T.L., 2009. A review of nitrogen enrichment effects on three biogenic GHGs: the CO₂ sink may be largely offset by stimulated N₂O and CH₄ emission. *Ecol. Lett.* 12, 1103–1117. <https://doi.org/10.1111/j.1461-0248.2009.01351.x>.
- Liu, S., Luo, D., Cheng, R., Yang, H., Wu, J., Shi, Z., 2020. Soil-atmosphere exchange of greenhouse gases from typical subalpine forests on the eastern Qinghai-Tibetan Plateau: Effects of forest regeneration patterns. *Land Degrad. Dev.* 31, 2019–2032. <https://doi.org/10.1002/ldr.3586>.
- Maarastawi, S.A., Frindte, K., Geer, R., Krober, E., Knief, C., 2018. Temporal dynamics and compartment specific rice straw degradation in bulk soil and the rhizosphere of maize. *Soil Biol. Biochem.* 127, 200–212. <https://doi.org/10.1016/j.soilbio.2018.09.028>.
- Mazza, G., Agnelli, A.E., Lagomarsino, A., 2021. The effect of tree species composition on soil C and N pools and greenhouse gas fluxes in a mediterranean reforestation. *J. Soil Sci. Plant Nutr.* 21, 1339–1352. <https://doi.org/10.1007/s42729-021-00444-w>.
- McNamara, N.P., Black, H.L.J., Pearce, T.G., Reay, D.S., Ineson, P., 2008. The influence of afforestation and tree species on soil methane fluxes from shallow organic soils at the UK Gisburn Forest Experiment. *Soil Use Manag.* 24, 1–7. <https://doi.org/10.1111/j.1475-2743.2008.00147.x>.
- Mills, R.T.E., Dewhurst, N., Sowerby, A., Emmett, B.A., Jones, D.L., 2013. Interactive effects of depth and temperature on CH₄ and N₂O flux in a shallow podzol. *Soil Biol. Biochem.* 62, 1–4. <https://doi.org/10.1016/j.soilbio.2013.03.003>.
- Mochizuki, Y., Koba, K., Yoh, M., 2012. Strong inhibitory effect of nitrate on atmospheric methane oxidation in forest soils. *Soil Biol. Biochem.* 50, 164–166. <https://doi.org/10.1016/j.soilbio.2012.03.013>.
- Montzka, S.A., Dlugokencky, E.J., Butler, J.H., 2011. Non-CO₂ greenhouse gases and climate change. *Nature* 476, 43–50. <https://doi.org/10.1038/nature10322>.
- Plain, C., Epron, D., 2021. Pulse labelling of deep A-horizons in forest with (CH₄)-C-13: testing a new method for tracing methane in the upper horizons, understorey vegetation and tree stems using laser-based spectrometry. *Biogeochemistry* 153, 215–222. <https://doi.org/10.1007/s10533-021-00775-x>.
- Prescott, C.E., Grayston, S.J., Helmisaari, H.S., Kustovska, E., Korner, C., Lambers, H., Meier, I.C., Millard, P., Ostonen, I., 2020. Surplus Carbon Drives Allocation and Plant-Soil Interactions. *Trends Ecol. Evol.* 35, 1110–1118. <https://doi.org/10.1016/j.tree.2020.08.007>.
- Rask, H., Schoenau, J., Anderson, D., 2002. Factors influencing methane flux from a boreal forest wetland in Saskatchewan, Canada. *Soil Biol. Biochem.* 34, 435–443. [https://doi.org/10.1016/s0038-0717\(01\)00197-3](https://doi.org/10.1016/s0038-0717(01)00197-3).
- Ren, L., Wu, X.W., Ma, D.L., Liu, L., Li, X., Song, D.D., 2022. Nitrite-dependent anaerobic methane oxidation bacteria and potential in permafrost region of Daxing'an Mountains. *Appl. Microbiol. Biotechnol.* 106, 743–754. <https://doi.org/10.1007/s00253-021-11739-4>.
- Saari, A., Heiskanen, J., Martikainen, P.J., 1998. Effect of the organic horizon on methane oxidation and uptake in soil of a boreal Scots pine forest. *FEMS microbiology ecology* 26, 245–255. <https://doi.org/10.1111/j.1574-6941.1998.tb00509.x>.
- Sihl, D., Xu, X., Ortiz, M.S., O'Connell, C.S., Silver, W.L., Lopez-Lloreda, C., Brenner, J.M., Quinn, R.K., Phillips, J.R., Newman, B.D., Mayes, M.A., 2021. Representing methane emissions from wet tropical forest soils using microbial functional groups constrained by soil diffusivity. *Biogeosciences* 18, 1769–1786. <https://doi.org/10.5194/bg-18-1769-2021>.
- Sjogersten, S., Melander, E., Wookey, P.A., 2007. Depth distribution of net methanotrophic activity at a mountain birch forest-tundra heath ecotone, northern Sweden. *Arct. Antarct. Alp. Res.* 39, 477–480. [https://doi.org/10.1657/1523-0430\(06-0099\)](https://doi.org/10.1657/1523-0430(06-0099)).
- Song, C., Zhang, J., Wang, Y., Wang, Y., Zhao, Z., 2008. Emission of CO₂, CH₄ and N₂O from freshwater marsh in northeast of China. *J. Environ. Manag.* 88, 428–436. <https://doi.org/10.1016/j.jenvman.2007.03.030>.
- Subke, J.A., Reichstein, M., Tenhunen, J.D., 2003. Explaining temporal variation in soil CO₂ efflux in a mature spruce forest in Southern Germany. *Soil Biol. Biochem.* 35, 1467–1483. [https://doi.org/10.1016/s0038-0717\(03\)00241-4](https://doi.org/10.1016/s0038-0717(03)00241-4).
- Sullivan, B.W., Selman, P.C., Hart, S.C., 2013. Does dissolved organic carbon regulate biological methane oxidation in semiarid soils. *Glob. Change Biol.* 19, 2149–2157. <https://doi.org/10.1111/gcb.12201>.
- Sullivan, B.W., Nifong, R.L., Nasto, M.K., Alvarez-Clare, S., Dencker, C.M., Soper, F.M., Shoemaker, K.T., Ishida, F.Y., Zaragoza-Castells, J., Davidson, E.A., Cleveland, C.C., 2019. Biogeochemical recuperation of lowland tropical forest during succession. *Ecology* 100. <https://doi.org/10.1002/ecy.2641>.
- Takahashi, K., Sakabe, A., Kanazawa, A., Kosugi, Y., 2021. Vertical profiles of methane concentration above and within the canopy of a temperate Japanese cypress forest. *Atmos. Environ.* -X 12. <https://doi.org/10.1016/j.aeoaa.2021.100143>.
- Tate, K.R., 2015. Soil methane oxidation and land-use change - from process to mitigation. *Soil Biol. Biochem.* 80, 260–272. <https://doi.org/10.1016/j.soilbio.2014.10.010>.
- Templer, P.H., Silver, W.L., Pett-Ridge, J., DeAngelis, K.M., Firestone, M.K., 2008. Plant and microbial controls on nitrogen retention and loss in a humid tropical forest. *Ecology* 89, 3030–3040. <https://doi.org/10.1890/07-1631.1>.
- Tian, H., Lu, C., Ciais, P., Michalak, A.M., Canadell, J.G., Saikawa, E., Huntzinger, D.N., Gurney, K.R., Stich, S., Zhang, B., Yang, J., Bousquet, P., Bruhwiler, L., Chen, G.,

- Drugokencky, E., Friedlingstein, P., Melillo, J., Pan, S., Poulter, B., Prinn, R., Saunois, M., Schwalm, C.R., Wofsy, S.C., 2016. The terrestrial biosphere as a net source of greenhouse gases to the atmosphere. *Nature* 531, 225. <https://doi.org/10.1038/nature16946>.
- Wang, D.Y., Repo, E., He, F.S., Zhang, X.W., Xiang, H.R., Yang, W.C., Min, X.B., Zhao, F.P., 2022. Dual functional sites strategies toward enhanced heavy metal remediation: Interlayer expanded Mg-Al layered double hydroxide by intercalation with L-cysteine. *J. Hazard. Mater.* 439. <https://doi.org/10.1016/j.jhazmat.2022.129693>.
- Wang, J.M., Murphy, J.G., Geddes, J.A., Winsborough, C.L., Basiliko, N., Thomas, S.C., 2013. Methane fluxes measured by eddy covariance and static chamber techniques at a temperate forest in central Ontario, Canada. *Biogeosciences* 10, 4371–4382. <https://doi.org/10.5194/bg-10-4371-2013>.
- Wei, H., Peng, C., Liu, S., Liu, X., Li, P., Song, H., Yuan, M., Wang, M., 2018. Variation in soil methane fluxes and comparison between two forests in China. *Forests* 9. <https://doi.org/10.3390/f9040204>.
- Wu, Y., Chen, W.J., Li, Q., Guo, Z.Q., Li, Y.Z., Zhao, Z.W., Zhai, J.Y., Liu, G.B., Xue, S., 2021. Ecoenzymatic stoichiometry and nutrient limitation under a natural secondary succession of vegetation on the Loess Plateau, China. *Land Degrad. Dev.* 32, 399–409. <https://doi.org/10.1002/ldr.3723>.
- Xiang, H.R., Min, X.B., Tang, C.J., Sillanpaa, M., Zhao, F.P., 2022. Recent advances in membrane filtration for heavy metal removal from wastewater: A mini review. *J. Water Process Eng.* 49. <https://doi.org/10.1016/j.jwpe.2022.103023>.
- Xu, M.P., Li, W.J., Wang, J.Y., Zhu, Y.F., Feng, Y.Z., Yang, G.H., Zhang, W., Han, X.H., 2022. Soil ecoenzymatic stoichiometry reveals microbial phosphorus limitation after vegetation restoration on the Loess Plateau, China. *Sci. Total Environ.* 815. <https://doi.org/10.1016/j.scitotenv.2022.152918>.
- Yang, X., Zhu, Y., Xu, Y., Li, X., Zhang, S., Qian, Q., et al., 2022. Simulated warming and low O₂ promote N₂O and N₂ emissions in subtropical montane forest soil. *J. Soils Sediment.* 1–14. <https://doi.org/10.1007/s11368-022-03234-8>.
- Yang, Z.H., Gong, H.Y., He, F.S., Repo, E., Yang, W.C., Liao, Q., Zhao, F.P., 2022. Iron-doped hydroxyapatite for the simultaneous remediation of lead-, cadmium- and arsenic-co-contaminated soil. *Environ. Pollut.* 312. <https://doi.org/10.1016/j.envpol.2022.119953>.
- Zhang, K.R., Cheng, X.L., Dang, H.S., Ye, C., Zhang, Y.L., Zhang, Q.F., 2013. Linking litter production, quality and decomposition to vegetation succession following agricultural abandonment. *Soil Biol. Biochem.* 57, 803–813. <https://doi.org/10.1016/j.soilbio.2012.08.005>.
- Zhou, M., Wang, X., Ren, X., Zhu, B., 2019. Afforestation and deforestation enhanced soil CH₄ uptake in a subtropical agricultural landscape: Evidence from multi-year and multi-site field experiments. *Sci. Total Environ.* 662, 313–323. <https://doi.org/10.1016/j.scitotenv.2019.01.247>.
- Zhu, D., Wu, N., Bhattarai, N., Oli, K.P., Chen, H., Rawat, G.S., Rashid, I., Dhakal, M., Joshi, S., Tian, J., Zhu, Qa, Chaudhary, S., Tshering, K., 2021. Methane emissions respond to soil temperature in convergent patterns but divergent sensitivities across wetlands along altitude. *Glob. Change Biol.* 27, 941–955. <https://doi.org/10.1111/gcb.15454>.