

Check for updates

Research

Mycorrhizal symbiosis pathway and edaphic fertility frame root economics space among tree species

Han Yan^{1,2} (b), Grégoire T. Freschet³ (b), Huimin Wang^{1,2}, James Aaron Hogan^{4,5} (b), Shenggong Li^{1,2}, Oscar J. Valverde-Barrantes⁴ (b), Xiaoli Fu^{1,2} (b), Ruili Wang⁶ (b), Xiaoqin Dai¹, Lei Jiang¹, Shengwang Meng¹, Fengting Yang¹, Miaomiao Zhang⁷ and Liang Kou^{1,2} (b)

¹Qianyanzhou Ecological Research Station, Key Laboratory of Ecosystem Network Observation and Modeling, Institute of Geographic Sciences and Natural Resources Research, Chinese Academy of Sciences, Beijing 100101, China; ²College of Resources and Environment, University of Chinese Academy of Sciences, Beijing 100049, China; ³Station d'Ecologie Théorique et Expérimentale, CNRS, 2 route du CNRS, Moulis 09200, France; ⁴Department of Biological Sciences, Institute of Environment, International Center of Tropical Biodiversity, Florida International University, Miami, FL 33199, USA; ⁵Department of Biology, University of Florida, Gainesville, FL 32605, USA; ⁶College of Forestry, Northwest A&F University, Yangling 712100, China; ⁷State Key Laboratory of Tree Genetics and Breeding, Key Laboratory of Tree Breeding and Cultivation of State Forestry Administration, Research Institute of Forestry, Chinese Academy of Forestry, Beijing 100091, China

Author for correspondence: Liang Kou Email: koul@igsnrr.ac.cn

Received: 20 November 2021 Accepted: 18 February 2022

New Phytologist (2022) **doi**: 10.1111/nph.18066

Key words: absorptive root, arbuscular mycorrhizas and ectomycorrhizas, root economics, root trait variation, trait interplay, trait–function linkage.

Summary

• The root economics space (RES) is multidimensional and largely shaped by belowground biotic and abiotic influences. However, how root-fungal symbioses and edaphic fertility drive this complexity remains unclear.

• Here, we measured absorptive root traits of 112 tree species in temperate and subtropical forests of China, including traits linked to functional differences between arbuscular mycor-rhizal (AM) and ectomycorrhizal (ECM) hosts.

• Our data, from known mycorrhizal tree species, revealed a 'fungal-symbiosis' dimension distinguishing AM from ECM species. This divergence likely resulted from the contrasting mycorrhizal evolutionary development of AM vs ECM associations. Increased root tissue cortical space facilitates AM symbiosis, whereas increased root branching favours ECM symbiosis. Irrespective of mycorrhizal type, a 'root-lifespan' dimension reflecting aspects of root construction cost and defence was controlled by variation in specific root length and root tissue density, which was fully independent of root nitrogen content. Within this function-based RES, we observed a substantial covariation of axes with soil phosphorus and nitrate levels, highlighting the role played by these two axes in nutrient acquisition and conservation.

• Overall, our findings demonstrate the importance of evolved mycorrhizal symbiosis pathway and edaphic fertility in framing the RES, and provide theoretical and mechanistic insights into the complexity of root economics.

Introduction

Roots have evolved diverse resource acquisition strategies since the emergence of the first land plants (Ma *et al.*, 2018; Bergmann *et al.*, 2020). Belowground, multiple plant traits, representing aspects of root architecture, morphology, anatomy, physiology or associations with other organisms, concomitantly define the capacity of plants to acquire resources (Freschet *et al.*, 2021b). Although a number of trade-offs define and limit the set of trait values possible to ensure plant viability and success (Weigelt *et al.*, 2021), and these trade-offs are further modulated by environmental constraints (Laughlin *et al.*, 2021), plants show a remarkable diversity of trait values and trait combinations to achieve similar levels of resource acquisition and growth (Freschet *et al.*, 2018; Guerrero-Ramírez *et al.*, 2021). Some of this diversity can nonetheless be summarized using trait-based root

© 2022 The Authors *New Phytologist* © 2022 New Phytologist Foundation economics frameworks (Kong et al., 2014; Valverde-Barrantes et al., 2015; Bergmann et al., 2020). Recent advances have demonstrated the merits of considering a multidimensional root economics space (RES), yet the proposed patterns are highly variable among studies (Kong et al., 2014; Valverde-Barrantes et al., 2015; Weemstra et al., 2016; Liese et al., 2017; Bergmann et al., 2020). This is because roots inhabit in complex soil matrix and interact with both biotic (e.g. mycorrhizal fungi) and abiotic (e.g. nutrient availability) factors that shape root economics in contrasting ways across plant functional types, environments and types of plant-fungi and plant-bacteria symbioses (Bardgett et al., 2014; Valverde-Barrantes et al., 2015; Weemstra et al., 2020). Our conceptual and practical understanding of how such factors, particularly mycorrhizal symbioses and edaphic fertility, drive the complexity of root economics remains largely incomplete (Bergmann et al., 2020; Ding et al., 2020).

Over 90% of woody plant species on Earth have developed symbioses with mycorrhizal fungi (Brundrett, 2009; Bonfante & Genre, 2010). Arbuscular mycorrhizal (AM) and ectomycorrhizal (ECM) roots are the two most common types of symbioses with contrasting evolutionary history and symbiotic (intracellular vs intercellular) pathways (Brundrett, 2002; Tedersoo et al., 2020). For AM species, mycorrhizal associations evolved in early land plants via colonization of primitive roots by fungal hyphae of early glomeromycota and endogonomycetes, where hyphae penetrated cortical cells to form intracellular arbuscules (Brundrett, 2002; Strullu-Derrien et al., 2018; Tedersoo & Bahram, 2019). By contrast, the more recently evolved ECM fungal association forms a sheathing mantle and Hartig net, where fungal hyphae grow around and between epidermal and cortical cells of roots (Smith & Read, 2008; Martin et al., 2016), although for ECM gymnosperms (e.g. Pinaceae) the inner hyphae can reach deeper into cortical cells (Brundrett, 2002; Genre et al., 2020). Ectomycorrhizal fungi have partly evolved from their saprotrophic ancestors (i.e. basidiomycetes and ascomycetes) and exude hormones such as ethylene and auxin (Raudaskoski & Kothe, 2015; Martin et al., 2016; Tedersoo & Brundrett, 2017), which enable roots to proliferate and branch for increased fungal colonization (Fig. 1; Comas et al., 2014; van der Heijden et al., 2015). By contrast, AM species mainly regulate the extent of root cortical tissue to provide the main habitat for AM fungi (Fig. 1). This fundamental divergence implies contrasting influences of mycorrhizal type on root economics. For example, while a trade-off between

mycorrhizal colonization intensity and mean root diameter has been clearly established for AM species, such evidence is scarce for ECM species (Kong *et al.*, 2014; Bergmann *et al.*, 2020; Wambsganss *et al.*, 2021). As such, it has been hypothesized instead that a higher branching intensity (BI, i.e. the number of root tips on a given length unit of parent root) of absorptive roots may be the key trait responsible for an increased probability of colonization by ECM fungi, but this remains to be tested on a comprehensive range of species and traits associated with symbiotic pathways.

At the global scale, plant roots differ in their strategies to acquire and conserve edaphic resources along at least two major axes (or functions, Fig. 2a) (Bergmann et al., 2020). A first axis is the 'conservation gradient' ranging from roots with high root tissue density (RTD) that show a slow resource return on investment but are long-lived and well-protected, to roots with a high nitrogen (N) content and metabolic rate for fast resource return on investment but a short life span. A second is the 'collaboration gradient' ranging from a 'do-it-yourself' soil exploration strategy, characterized by high specific root length (SRL) to an 'outsourcing' resource strategy where carbon (C) is allocated to the mycorrhizal partner, typified by greater root diameter and cortical fraction. We propose that this simple view of the 'collaboration' gradient does not adequately reflect the root functional differentiation among woody AM and ECM species, and therefore propose a more refined 'fungal-symbiosis' gradient. Particularly, while AM associations tend to rely on increasing root diameter and cortex



Fig. 1 Schematic depiction of divergent symbiosis pathway of arbuscular mycorrhizal (AM) and ectomycorrhizal (ECM) roots. (a) In AM roots, fungal hyphae (depicted in red) principally from glomeromycota and endogonomycetes grow between and penetrate root cortical cells to form intracellular arbuscules and vesicles. Thus, AM plant roots tend to facilitate mycorrhizal symbiosis by expanding cortical space for increased habitat availability for AM fungi. In ECM roots, fungal hyphae (depicted in blue) principally from basidiomycetes and ascomycetes and stemming from saprotrophic ancestors grow attached to the epidermal cell of lateral roots to form a sheathing mantle and then develop an intercellular 'Hartig net' around root cortical cells (ECM angiosperms) or around and into cortical cells (ECM gymnosperms). (b) For AM species, mycorrhizal colonization intensity can be approximated by the proportion of total absorptive root length colonized. (c) For ECM species, mycorrhizal colonization intensity is rather a fungal occurrence frequency on root tips. Ectomycorrhizal fungal taxa exudate hormones such as ethylene and auxin to stimulate proliferation of lateral roots, thereby ECM species tend to regulate root branching structure hosting mycorrhizal fungi. The left diagram is inspired by Bonfante & Genre (2010).

New Phytologist (2022) www.newphytologist.com





Fig. 2 Conceptual framework for root trait–function linkage and trait interplay. (a) At the root-system level, root economics run on plant-derived carbon ('capital') and soil-derived nutrients ('commodities'), reflecting resource return (nutrient acquisition) on investment (carbon expense). Root diameter (RD) and specific root length (SRL) simultaneously link to two major root functions – root-lifespan and fungal-symbiosis. This multifunctionality connects root tissue density (RTD) and root nitrogen concentration (RN), both related to root lifespan, with symbiotic traits (cortex thickness (CT) and branching intensity (BI), related to fungal symbiosis). Thus, root lifespan is represented by interactions among RD, SRL, RTD and RN, while fungal symbiosis is represented by interactions among RD, SRL, BI and CT. The role of RD and SRL in root economics is determined by the interplay among root traits. (b) By using the term 'trait interplay' in root economics, we here illustrate that root trait trade-offs are not necessarily bivariate but can be more complex, involving multiple traits simultaneously at the root-system level. The mathematical causal trait relationships (solid lines) and expected trait covariations (dotted lines) are displayed by red arrows and blue arrows that denote positive and negative relationships, respectively. Following Freschet *et al.* (2021b), trait position along the vertical axis represents trait hierarchy, with upper levels representing 'basal' traits and lower levels representing 'composite' traits. The chemical root traits are not displayed in root trait interplay for brevity.

space, ECM associations are typically represented by thin, highly branched roots with many root tips (Comas *et al.*, 2014). Therefore, a trade-off in symbiotic root traits can be envisioned to exist between the 'symbiotic traits' of BI and root diameter with opposite trends between symbiotic groups.

Here, we contend that the relevance of certain root traits to multiple functions within the RES (e.g. root diameter and SRL can be conceptually linked to both RES axes) not only limits our capacity to apply a singular interpretation to axes of RES variation but also likely leads to substantial differences in local representations of the RES (e.g. across biomes or plant functional types). Particularly, the degree of variation observed in root trait strategies along axes on the RES seems to depend on the degree of environmental variation studied. Along the conservation gradient, root lifespan has been linked to changing availability of soil nutrients, particularly N (Eissenstat & Yanai, 1997; McCormack & Guo, 2014), suggesting an often ignored but positive role of root lifespan (as well as associated changes towards high RTD, root diameter and C/N, and low SRL) in nutrient acquisition (Ryser, 1996; Eissenstat et al., 2000; Withington et al., 2006; McCormack et al., 2012). Along the proposed 'fungal-symbiosis' gradient, plants would rely on different symbiotic pathways with mycorrhizal fungi to forage immobile soil P (van der Heijden et al., 2015; Liu et al., 2018), via regulating root cortex space for AM species and branching tips for ECM species (Comas et al., 2014). Thus, the diversity of belowground strategies along RES axes depends to some extent on the availability and mobility of limiting nutrients, such as N and P in soil, which also interacts with mycorrhizal association type.

To reveal how root-fungal symbiotic pathways and edaphic fertility drive root economics of woody plants, we sampled absorptive fine roots (the resource-acquiring first- and second-order roots within root systems) of 112 tree species consisting of 77 AM species and 35 ECM species spanning two temperate forests and two subtropical forests in China. We considered seven root economics traits (SRL, RTD, root diameter, cortex thickness (CT), BI, root N concentration (RN) and root carbon concentration (RC)) that are representative of root construction strategy, root-fungal symbiotic pathway and resource return on investment (i.e. soil-derived nutrient acquisition per plant-derived C allocation) at the root system level (Kong et al., 2014; Bergmann et al., 2020). Compared with intraspecific variation, interspecific variation in root traits is often large (Hogan et al., 2020). Moreover, among taxa root traits are highly phylogenetically structured, evidencing how evolution has largely determined root functional differences (Valverde-Barrantes et al., 2013, 2017). Based on phylogenetically informed analyses, we hypothesized that both AM and ECM tree species would fit within a bidimensional RES, represented by:

(1) a resource 'conservation' gradient describing the trade-off between slow vs fast resource return on investment dominated by traits associated with root lifespan, independent of mycorrhizal types;

(2) a 'fungal-symbiosis' dimension that not only accounts for the dichotomy between species adopting a 'do-it-yourself' vs an 'out-sourcing' strategy of soil resource acquisition but also represents differences between mycorrhizal types, with strategies ranging from large root diameters and CTs for AM species to high BIs for ECM species.

Based on current knowledge of tree species root trait distributions along soil resource gradients, we further hypothesized that: (3) interspecific root trait variation along the 'root-lifespan' dimension is more associated with soil N availability, while variation along the 'fungal-symbiosis' dimension is more associated with soil P availability for both mycorrhizal types.

Materials and Methods

Study site and species selection

This study was conducted in four temperate and subtropical forests of China. The four forests were located at: (1) Menglun forestry station (41°52'-42°01'N, 116°49'-116°59'E) in Hebei province; (2) northern slopes of Taibai Mountain (33°49'-34°10'N, 107°19'-107°58'E) in Shaanxi province; (3) Qianyanzhou Ecological Station (26°44'39"N, 115°03'33"E) in Jiangxi province; and (4) Yangming Mountain (25°39'26"N, 114°18'49"E) in Jiangxi Province. The sampling sites range from 102 to 2100 m above sea level in elevation, with mean annual temperature ranging from 2°C to 17.9°C, and mean annual precipitation of 470-1587 mm (Kou et al., 2019; Yan et al., 2019; Jiang et al., 2021). The species we selected are representative of the local plant community at each site. In total, we sampled 366 individuals of 112 woody species and then categorized these species into 77 AM (from 58 genera of 34 families) and 35 ECM species (from 17 genera of seven families) regarding symbiotic guild assignments of Steidinger et al. (2019). Each site has a unique set of species, as described in Supporting Information Table S1.

Field sampling and processing

The field sampling was carried out in autumn of 2018 and 2020. Samples from each species were collected from at least three mature healthy trees with similar diameter at breast height (DBH) to minimize the potential influences of intraspecific difference (< 5 cm in DBH) on root traits. The individuals for each species were spaced at least 10 m apart to ensure sampling independency. Surface soil (0-20 cm) was excavated around the stems of the trees to expose lateral roots. Intact roots were cut from the main lateral roots, and the attached soil was cleared following the procedure described by Guo et al. (2008b). At least eight intact distal root branches, including more than five root orders, were sampled. Each root sample was then divided into two subsamples. One subsample was gently washed in deionized water to remove adhering organic matter and was immediately fixed in formalin-acetic acid-alcohol solution (90 ml 50% ethanol v/v, 5 ml 100% glacial acetic acid v/v and 5 ml 37% methanol v/v) for measuring anatomical traits. The other subsamples were placed into valve bags, incubated with ice bags and then stored at -20°C for later dissection and analyses of morphological and chemical trait (Guo et al., 2008b). Approximately 1 kg of soil was also collected during root sampling (0-20 cm) from under each tree using a shovel. All soil samples were sieved (2-mm mesh) to remove roots and visible organic particles and stored at 4°C for later laboratory analysis. Soil samples were generally extracted and analysed within 1-2 wk after sampling to minimize the influence of storage time on available nutrients (Bailey et al., 2022).

Root trait measurements

The first set of root subsamples was dissected based on the protocol described by Pregitzer *et al.* (2002) for measuring morphological

root traits. Specifically, three root clusters, including at least five root orders, were randomly chosen and dissected with fine forceps on a plate filled with deionized water as described by Guo et al. (2008). The absorptive roots (1st- and 2nd-order roots) were used in this study because they are belowground resource-acquiring units (Guo et al., 2008b; McCormack et al., 2015; Kou et al., 2018). Root diameter was measured using WINRHIZO ARABIDO v.2012b (Regents Instruments Inc., Quebec City, QC, Canada) by analysing images of root samples scanned on an Epson Expression 10000 XL scanner (Seiko Epson Corp., Suwa, Japan) at a resolution of 400 dpi. Given the difficulties in distinguishing mycorrhizal structures from roots, the mycorrhizal mantle was included in the measurement of root diameter for ECM species. Specific root length was calculated as the ratio of root length to root dry mass, and RTD was calculated as the ratio of root dry mass to root volume. Root volume was recalculated from WINRHIZO outputs as the sum of averages of all diameter classes (Freschet et al., 2021a). Referring to the calculation of BI for the 1st-order roots (Kong et al., 2014), we calculated BI of absorptive roots as the number of their root tips divided by the total root length of their parent roots (i.e. the 3rd-order roots). All absorptive roots were weighed, oven-dried and then ground to fine powders using a Spex 8000-D mixer mill (Spex, Edison, NJ, USA) for measuring the concentrations of C and N using a Vario EL Cube elemental analyser (Elementar, Hanau, Germany). Root trait values were averaged across all sampled replicates for each species.

The second subsample set was also dissected in the same manner as morphology and chemistry for measuring anatomical traits. More than 20 segments were randomly selected from the absorptive roots, stained with safranin-fast green, dehydrated in a set of alcohol solutions and embedded in paraffin. 8-µm-thick root sections were prepared, photographed using a compound microscope (DM2500, DFC450; Leica, Wetzlar, Germany) and measured for CT using IMAGEJ (NIH Image, Bethesda, MD, USA).

Soil measurements

Soil total C and N concentrations were determined using the Vario EL Cube elemental analyser described above. Approximately 13–15 g of soil was used for the extraction (50 ml of 2 mol 1^{-1} KCl) of mineral soil N (NH₄⁺ and NO₃⁻). The concentrations of NH₄⁺ and NO₃⁻ were determined using a Flow Auto Analyser (Autoanalyser III; Bran + Luebbe, Norderstedt, Germany). Available P from acidic and alkaline soil samples was extracted, respectively, using HCl/NH₄F and NaHCO₃, and then determined colorimetrically using an ascorbic acid molybdate analysis on a Flow Auto Analyser. Soil properties were averaged across all soil samples taken from each tree replicates.

Data analyses

The taxonomic binomials of all species were verified using *The Plant List* (TPL, http://www.theplantlist.org/). Plant family-level clades were determined according to APG4 (The Angiosperm Phylogeny Group *et al.*, 2016) for angiosperms and TPL for gymnosperms. Phylogenetic trees were constructed, respectively, for all species, AM species and ECM species using V.PHYLOMAKER

(Jin & Qian, 2019), a package designed to generate phylogenies for vascular plants in R software (v.3.6.3; R Core Team, 2020), and V.PHYLOMAKER uses the mega-tree derived primarily from GBOTB (GenBank taxa with backbone provided by Open Tree of Life v.9.1) for seed plants (Smith & Brown, 2018) and Zanne et al.'s (2014) phylogeny for pteridophytes (Jin & Qian, 2019). The phylogenetic tree across all species was then visualized with the annotation of plant phylogeny, mycorrhizal type and distribution of core traits (BI, CT, root diameter, RTD and SRL) using the GGTREE package (Yu et al., 2017) in R. Phylogenetic polytomies were resolved by the 'multi2di' function using the APE package (Paradis & Schliep, 2019) in R. We calculated Blomberg's parametric K to evaluate the phylogenetic conservatism for each absorptive root trait using the PICANTE R package (Blomberg et al., 2003; Münkemüller et al., 2012). A significant phylogenetic signal indicates that the trait is constrained by phylogeny, and a higher value of Blomberg's Kindicates higher phylogenetic conservatism (Blomberg et al., 2003). The variation in each root trait across all species was estimated by calculating the means, minima and maxima and the coefficient of variation (CV). Additionally, bivariate relationships among root traits were assessed using both linear and phylogenetic generalized least square models in the STATS and CAPER R packages (Freckleton et al., 2002; Orme et al., 2013) for all species, and for AM and ECM species, separately. Relationships between root trait and soil nutrient availability were assessed using Pearson's correlations for all species and the two mycorrhizal groups.

Given the influence of phylogeny on traits, we performed phylogenetically informed principal component analysis (pPCA) to account for the dependency in root trait variations for all 112 species across the phylogeny. The pPCA was implemented with Pagel's lambda correlation structure using the 'phyl.pca' function of the PHYTOOLS package (Revell & Collar, 2009; Uyeda et al., 2015) in R. The pPCA was conducted for seven key root traits: BI, CT, root diameter, RTD, SRL, RN and RC that are previously reported as drivers of the trade-offs between fungal dependency and root construction strategies at global levels (Brundrett, 2002; Kong et al., 2014; Bergmann et al., 2020). To check for the effect of potential contamination of roots by soil minerals (potentially different among species, e.g. between ECM and AM species) on the general trends observed in this study, we carried out the same analyses with RTD data corrected for C concentration (i.e. expressed on a root C mass basis rather than root dry mass basis). Because the observed trends were the same, only the noncorrected data were displayed. Factor loadings of pPCA on corrected data can be found in Table S2.

To test for the difference between AM and ECM species on 'fungal-symbiosis' dimension, we first performed an ANOVA on the second axis scores of AM vs ECM species based on the result of pPCA for all species. We then repeated pPCA for AM (n=77) and ECM (n=35) species, respectively. Given the contrasting symbiosis pathways between ECM angiosperms and ECM gymnosperms from the Pinaceae family (Brundrett, 2002), we further performed this analysis for ECM angiosperms (n=28) by removing Pinaceae species (n=7). Lastly, linear regressions were used to examine the relationships of soil nutrients with pPCA loadings on the 'root-lifespan' dimension (PC1) and the 'fungalsymbiosis' dimension (PC2) for all species, AM species and ECM species. To meet the assumption of normality, we log₁₀transformed data for each root trait in bivariate–correlation analyses and pPCAs.

Results

Variations in root traits and plant phylogeny influences

Across 112 tree species (Table S1), we found patterns of variation in absorptive root traits (Table 1) that differ from previously published global patterns of trait variation, as available from the GRooT database (Guerrero-Ramírez et al., 2021) (Table S3). The symbiotic root traits (BI and CT) varied considerably, with BI ranging from a minimum of 0.77 cm⁻¹ of *Cinnamomum por*rectum (AM species) to a maximum of 12.80 cm⁻¹ of Castanea mollissima (ECM species) (coefficient of variation (CV) = 63.9% vs 60.2% in the GRooT database). There was an eightfold variation in CT, ranging from 0.04 mm of C. mollissima (ECM species) to 0.32 mm of Diospyros lotus (AM species) with a CV of 36.8% vs 59.5% in the GRooT database. For morphological root traits, there were greater variations in RTD (CV=80.2% vs 68.5% in the GRooT database) and SRL (CV=77.6% vs 181.5% in the GRooT database) than RD (CV=16.0% in our data set vs 45.1% in the GRooT database). For root tissue C and N, the variation in RC (CV=10.4% vs 16.6% in the GRooT database) was smaller than that in RN (CV = 25.3% vs 37.3% in the GRooT database). The very low value of RC observed in some species suggests that roots may have been contaminated

Table 1 Summary of absorptive root traits and phylogenetic signals (Blomberg's K) for 112 tree species.

Trait	Abbreviation	Units	Min	Max	Mean	CV%	K value
Branching intensity	BI	branches cm ⁻¹	0.77	12.80	4.25	63.9	0.029
Cortex thickness	СТ	mm	0.04	0.32	0.18	36.8	0.044
Root diameter	RD	mm	0.31	0.62	0.38	16.0	0.080
Root tissue density	RTD	g cm ⁻³	0.04	0.97	0.25	80.2	0.018
Specific root length	SRL	$m g^{-1}$	7.78	303.96	69.27	77.6	0.014
Root nitrogen concentration	RN	%	0.81	2.63	1.78	25.3	0.080
Root carbon concentration	RC	%	28.28	50.73	42.30	10.4	0.016

Max, maximum value of the trait; Min, minimum value of the trait.

Values (mean, n = 112); the significant K values are in bold (P < 0.05).

with soil minerals, especially in ECM roots that are harder to clean, with potential effects on associated values of those mass-related traits.

There were no significant phylogenetic signals for the two symbiotic traits that are most indicative of root construction tradeoffs with fungal association (CT for AM trees vs BI for ECM trees) when all species were analysed as well as when analyses were run within mycorrhizal types based on Blomberg's *K* values (P > 0.05; Table 1). Root diameter and RN were influenced by plant phylogeny and exhibited higher phylogenetic conservatism than other root traits (Table 1). For instance, gymnosperms and species in the Magnoliids tended to have thicker roots with higher RN, while species belonging to the more recently evolved clades like the Rosids showed the opposite patterns (Fig. 3). Also, similar patterns of phylogenetic influence were observed when separating all species into different mycorrhizal groups, except for root diameter that had a marginally significant phylogenetic signals for AM species (P = 0.073, Table S4).

Bivariate relationships among root traits

Our results from the phylogenetically corrected bivariate trait relationships, particularly for those related to symbiotic interactions and RN, showed striking differences when performed on all species vs on separate mycorrhizal groups (Figs 4, S1, S2). Statistically significant correlations were found among BI, CT and root diameter, with BI decreasing linearly with both CT and root diameter for all species $(r^2 = 0.04, P < 0.05 \text{ and } r^2 = 0.40, P < 0.001, \text{ respectively; Fig. 4})$ and for AM species only ($r^2 = 0.07$, P < 0.05 and $r^2 = 0.22$, P < 0.001, respectively; Fig. S1). However, for ECM species, we found negative relationships between BI and root diameter $(r^2 = 0.15, P < 0.05)$ and between CT and SRL $(r^2 = 0.12, P < 0.05)$ (Fig. S2). Additionally, RTD was negatively correlated with SRL $(r^2 = 0.95, P < 0.001;$ Fig. 4) for all species, and this correlation showed similar patterns for both AM ($r^2 = 0.93$, P < 0.001; Fig. S1) and ECM ($r^2 = 0.96$, P < 0.001; Fig. S2) species, separately. RN was negatively correlated with BI ($r^2 = 0.04$, P < 0.05) for all species, but had no significant relationships with BI when taking a close look at mycorrhizal type.

Multivariate ordination RES

In accordance with other research on the RES, the pattern of multivariate ordination was represented by two independent axes (dimensions) since the eigenvalues of the first two principal components were > 1 for all species (Table S5). The results of pPCA showed that the first two axes accounted for 55.5% of total variation for all species. The first axis was best described by SRL and RTD, while the second axis was represented by root diameter, BI and CT (Fig. 5a,b; Table S5). The results of the ANOVA analysis on the second axis scores of AM vs ECM species from the result of pPCA for all species indicated a significant influence of mycorrhizal type on RES axis too (P < 0.001; Table S6). We found a similar pattern of RES among AM species to that among all species with the first two axes explaining 31.3% and 27.3% of total variations (Fig. 5c,d; Table S5). For ECM species, the first

axis was also driven by SRL and RTD, explaining 32.5% of total variation, while the second axis was mostly driven by BI, RD and RN, explaining 19.6% of total variation (Fig. 5e,f; Table S5). A similar pattern was found for ECM angiosperms (i.e. when excluding gymnosperms from the ECM group) with a shift in the location of root diameter from the second axis to the first axis (Table S5; Fig. S3).

Linkages of root traits and RES with edaphic nutrients

We found divergent correlations between root traits and soil nutrient availability for all species and for species belonging to the different mycorrhizal groups (Table S7). Branching intensity was positively correlated with soil nitrate concentrations for AM species (r=0.38, P<0.001; Table S7), but was negatively correlated with soil available P and nitrate concentrations for ECM species (r=-0.48, P<0.01 and r=-0.43, P<0.01, respectively; Table S7). CT was negatively correlated with concentrations of available P and nitrate for all species (r=-0.23, P<0.05 and r=-0.24, P<0.05, respectively; Table S7) and AM species (r=-0.34, P<0.01 and r=-0.37, P<0.001, respectively; Table S7). Branching intensity and CT were unrelated to soil ammonium availability for all species (Table S7).

Consistent with these relationships between soil available nutrients and root traits, we observed several significant relationships between our pPCA axes and soil nutrient availability (Fig. 5; Table S8). For AM species, we found a significant tendency towards greater CT and root diameter on the second axis with decreasing soil availability of P ($r^2 = 0.05$, P < 0.05, Fig. 5d; Table S8) and nitrate $(r^2 = 0.08, P < 0.05)$. By contrast, ECM species showed significant tendency towards increased BI on the second axis with decreasing soil availability of P ($r^2 = 0.17$, P < 0.05, Fig. 5f; Table S8) and nitrate ($r^2 = 0.15$, P < 0.05). We found no significant relationships of the second axis with any soil nutrient factor for all species (Fig. 5b; Table S8). Additionally, we found a significant tendency towards higher RTD on the first axis with decreasing soil nitrate availability for all species and AM species ($r^2 = 0.06$, P < 0.01 and $r^2 = 0.06$, P < 0.05, respectively; Fig. 5a,c; Table S8), but decreasing soil P availability for ECM species ($r^2 = 0.13$, P < 0.05, Fig. 5e; Table S8). Soil ammonium availability was not correlated with either of the two axes for all species and both mycorrhizal types (Table S8).

Discussion

Building on a recent global representation of the RES, we demonstrate the context dependency of the RES as a conceptual framework. Specifically, we reveal two functionally independent ('root-lifespan' vs 'fungal-symbiosis') dimensions that differ from previous representations of the RES. First, a dimension reflecting aspects of resource 'conservation', hereafter called the 'root-lifespan' dimension to differentiate it from the 'conservation' axis described by Bergmann *et al.* (2020), which is characterized by a trade-off between SRL and RTD – two key traits that are associated with root construction strategy – rather than a trade-off between RTD and RN. Second, the 'fungal-symbiosis'

New Phytologist





Fig. 3 Plant phylogeny influences on key root economics traits. The inside diagram displays the phylogenetic tree of 112 tree species. The bottom colours for Latin names represent different clades, and circles at the tip of the phylogenetic branch denote mycorrhizal type coloured in blue for arbuscular mycorrhizal (AM) species or in orange for ectomycorrhizal (ECM) species. For the outside diagram, the standardized values of root economics traits involved in trait interplay are mapped onto each species with values ranging from low (green) to high (purple). Root diameter (RD) exhibits the highest phylogenetic conservatism among these root traits (Table 1). Specific information about clade and mycorrhizal type for each species is exhibited in Supporting Information Table S1. BI, branching intensity; CT, cortex thickness; RTD, root tissue density; SRL, specific root length.

dimension, which is best characterized by the direct (BI and CT) and indirect (root diameter) symbiotic traits, rather than a simple collaboration axis characterized by a trade-off between SRL and root diameter (Bergmann *et al.*, 2020). This refined perspective of the RES adds two traits which differentiate AM and ECM

mycorrhizal pathways (Fig. S4) on root functional morphologies. Additionally, two major changes distinguish our findings from these of Bergmann *et al.* (2020): the absence of RN along our 'root-lifespan' axis and the sharp decoupling of SRL with the 'fungal-symbiosis' axis.

8 Research



Fig. 4 Pairwise correlations among seven root economics traits for all species. The bivariate relationships among root traits using general linear and phylogenetically corrected regressions for all tree species (n = 112). Significant correlations are represented by regression lines coloured in blue for general linear regression and in red for phylogenetically corrected regression in scatter plots (upper triangle). Correlation coefficients are displayed in the same colours for the two regression analyses (lower triangle). BI, branching intensity; CT, cortex thickness; RC, root carbon concentration; RD, root diameter; RN, root nitrogen concentration; RTD, root tissue density; SRL, specific root length.

At the root-system level, root economics operate on plantderived C ('capital') and soil-derived nutrients ('commodities'), leading to a trade-off between C investment and nutrient return (Bloom *et al.*, 1985). Although both SRL and RTD are associated with the rate of return per unit of investment, they represent opposite directions based on the theoretical formula: SRL=4/ $(\pi \times \text{RTD} \times \text{root diameter}^2)$ (Ryser, 1996; Ostonen *et al.*, 2007) or empirical trade-off relationships (Ma *et al.*, 2018; Zhou *et al.*, 2018; Han & Zhu, 2021). These traits have traditionally been regarded as the belowground equivalent of aboveground specific leaf area and leaf tissue density that locate at the two ends of the one-dimensional 'fast-slow' leaf economics spectrum (Ryser,





Fig. 5 Fertility-driven root economics pattern varies with mycorrhizal type. Root trait biplots of phylogenetically informed principal component analyses (leaf diagram) for (a, b) all 112 tree species; (c, d) 77 arbuscular mycorrhizal (AM) species (blue circle); (e, f) 35 ectomycorrhizal (ECM) species (orange circle). The 'root-lifespan' dimension is characterized by specific root length (SRL) and root tissue density (RTD), while the 'fungal-symbiosis' dimension is predominated by cortex thickness (CT) and/or branching intensity (BI), depending on mycorrhizal type. Along with the two direct symbiotic traits, root diameter (RD) also loads on the 'fungal-symbiosis' dimension representing the outcome of the interplay between morphological and symbiotic root traits. Edaphic fertility is embedded into the biplots to indicate the influences of different soil nutrients on root economics with colour gradients from dark green (low NO₃⁻) to light green (high NO₃⁻), from dark grey (low AP) to light grey (high AP) and from dark brown (low AP + NO₃⁻) to light brown (low AP + NO₃⁻). The linear regressions are fitted (right diagram) between soil nutrient availability and axis loadings along the 'root-lifespan' dimension (PC1) for all species (a), AM species alone (c), ECM species alone (e) as well as along 'fungal-symbiosis' dimension and Table S8 for more details about the linear regression results. AP, available soil phosphorus concentration; NO₃⁻, soil nitrate concentration; PC, principal component; RC root carbon concentration; RN, root nitrogen concentration.

1996; Wright et al., 2004; Reich, 2014). Belowground, species with an 'acquisitive root strategy' tend to develop absorptive roots with higher SRL and shorter lifespan (McCormack et al., 2012), while species with a conservative root strategy tend to develop absorptive roots with higher RTD and longer lifespan (Ryser, 1996; Eissenstat et al., 2000). Therefore, the 'root-lifespan' dimension of the RES represents a meaningful shift from high SRL to high RTD and depicts a belowground acquisition-conservation trade-off for tree species. The strong relationship between SRL and RTD could be due to the low variability in root diameter, with SRL being mostly defined by RTD (Fig. 2b). Patterns of variation in absorptive root diameter from the current study (Table 1) differ markedly from previously published global patterns of variation in fine root (<2 mm) diameter for woody species (CV = 45.1%, Table S3; Fig. S5) in the GRooT database (Guerrero-Ramírez et al., 2021), which may cause the contrasting RES patterns. As such, the 'root-lifespan' dimension could be sensitive to root diameter variation, particularly when incorporating multiple root categorizations for woody species. Given that only a small share (c. 37%) of data in the GRooT database comes from absorptive roots, further studies with more standardized root categorization (e.g. absorptive roots) are needed to determine the general pattern of the RES.

The reasons why RN does not align with the axis of RTD variation, as observed across the global GRooT database (Guerrero-Ramírez et al., 2021) used by Bergmann et al. (2020) accounting for 7% shared variation between RN and RTD, remain unclear to us as this trend appeared consistent across their subset of woody species. Moreover, RN varied substantially in our data, although variation was smaller than in the GRooT data set (CV = 37.3%, Table S3; Fig. S5). One potential reason may relate to our focus on absorptive roots of the first and second orders, according to the most recent functional categorization of roots (McCormack et al., 2015), whereas the GRooT database includes a majority of root data representative of the historical focus on roots < 2 mm. Since roots of the most distal orders are known to be the most metabolically active and display higher RN concentrations than higher-order roots (Iversen et al., 2017; Freschet et al., 2021a), the RN gradient in Bergmann et al. (2020) could represent to some extent a gradient of more to less metabolically active sets of fine roots, which could explain some of the observed covariation with RTD. Indeed, inclusion of higher root orders should lead to both lower RN and higher RTD (Freschet & Roumet, 2017). Nonetheless, the negative correlation between RN and RTD was also observed on data from first-order roots only (6% shared variation; Ma et al., 2018).

Another potential reason might relate to the context dependency of the RN and root-lifespan relationship. RN scales with root metabolism (e.g. respiration, nutrient acquisition and assimilation rates), and the nutritional value of root tissue for primary consumers (Freschet *et al.*, 2021b). However, it is also strongly influenced by excess nutrient uptake and N storage, with variable outcomes depending on environmental conditions, and could therefore display a partly orthogonal orientation to root lifespan. Some of the N found in root tissues can be further associated with the capacity of roots to transport water and elements and thus partly positively covaries with stele fraction and RTD. Additionally, N is common in herbivore and pathogen defence compounds in roots and thus is partly associated positively with root lifespan (Guo *et al.*, 2008a). Finally, N is also found in higher concentration in mycorrhizal fungi tissues than in the root tissues (Langley & Hungate, 2003), suggesting that roots with high mycorrhizal colonization intensity are likely to display high RN. This is despite the fact that mycorrhiza formation can increase plant resistance to soil pathogens and herbivores (Jung *et al.*, 2012; Babikova *et al.*, 2014). Therefore, while we do see some evidence, both conceptually and in the literature, to describe root-lifespan axis along a continuum from high RTD to high RN, we also see multiple reasons why the alignment of the root-lifespan axis with root N concentration may vary across environments.

Importantly, root trait trade-offs are not necessarily bivariate and can be highly complex, involving multiple trait interactions at the root-system level (i.e. trait interplay, Fig. 2b). For example, root diameter and SRL have been shown to correlate positively and negatively with root lifespan, respectively (Eissenstat et al., 2000; McCormack et al., 2012), although their main contribution may appear limited along the single collaboration axis in simplified representations of the RES. Here, we included BI and CT, traits that are more directly related to mycorrhizal symbioses compared to root diameter, finding additional trade-offs among these three traits (Fig. 4). Consistent with our hypothesis, CT and BI that are most directly associated with mycorrhizal symbiosis pathways loaded evidently on the 'fungal-symbiosis' dimension, which varied with mycorrhizal type (Table S6). Root cortex tissue provides the main habitat for AM fungi to grow hyphae for nutrient foraging (Comas et al., 2014), explaining the emergence of CT on the 'fungalsymbiosis' dimension for AM species. We noticed that large CT fractions and high BI represented opposite sides on the 'fungalsymbiosis' dimension for AM species (Fig. 5c,d; Table S5), which was further evidenced by the negative relationship between these two traits (Fig. S1; also see Kong et al., 2014). This pattern points to a trade-off between nutrient acquisition strategies given the intracellular symbiosis pathway of AM species. Larger CT on the 'fungal-symbiosis' dimension indicates a fungal-symbiosis dependency in root strategy, where plants invest more C to expand cortical space for increased habitat availability for AM fungi at the cost of reduced conductivity of water flow into the roots (Wen et al., 2019). On the contrary, for AM tree species, larger BI indicates a self-dependent strategy in allocation towards construction of longer and more ramified roots, which are better able to exploit soil volume by themselves (Freschet & Roumet, 2017; Wen et al., 2019). Interestingly, this refined 'fungal-symbiosis' dimension is relatively well captured by root diameter alone, which covaries negatively with BI and positively with CT, highlighting the key role of root diameter in defining root economics strategy.

Compared to AM species, ECM species (except gymnosperms) have reduced the dependence on the root cortex but have relied more on the branching structure for symbiosis (Smith & Read, 2008; Martin *et al.*, 2016). Consistent with this evolutionary shift, BI rather than CT loaded on the 'fungal-symbiosis' dimension for ECM species (Fig. 5e,f; Table S5). This is not surprising when taking a closer look at co-evolution of the ECM symbiosis pathway. In

ECM angiosperms, hyphae grow attached to epidermal cells of emerging lateral roots to form the intercellular Hartig net and sheathing mantle (Kottke et al., 1987; Smith & Read, 2008; Martin et al., 2016). Compared with AM symbiosis, the maintenance of these unique structures is believed to consume more C, thus demanding more C supply transporting through sieve tubes (Smith & Read, 2008; Kong et al., 2021). The greater branching may enable larger surface area contacting with sieve tubes to gain allocated C for mycorrhizal symbiosis, leading to the compression of root cortex space. Although ECM gymnosperms have similar symbiosis pathway to AM species, CT was still absent on the 'fungal-symbiosis' dimension, possibly resulting from the limited sample size of Pinaceae (n = 7, 20% of the total). Root diameter shifted from the 'fungal-symbiosis' dimension to the 'root-lifespan' dimension when excluding ECM gymnosperms, further confirming the independence of ECM angiosperms on traits associated with root thickness (Table S5; Fig. S3). Unlike the trade-off between CT and BI in AM species, the branching traits may represent a higher probability of root tip colonization by ECM fungi as well as a potential feedback mechanism of ECM fungal colonization on root architecture (Comas et al., 2014; van der Heijden et al., 2015; Martin et al., 2016; Tedersoo & Brundrett, 2017).

Interestingly, root diameter in parallel to the two direct symbiotic traits defined the 'fungal-symbiosis' dimension, although it is inherently associated with SRL and RTD (Fig. 5; Table S3). This pattern likely arises due to two reasons. First, relationships among diameter-related traits are nonlinear (Ma et al., 2018; Kong et al., 2019). RTD is independent of (Kramer-Walter et al., 2016; Bergmann et al., 2020), or coaxial with SRL in the RES (Kong et al., 2014), and can have variable relationships with root diameter (Withington et al., 2006; Comas & Eissenstat, 2009; Kramer-Walter et al., 2016). According to the formula illustrated above, the bivariate relations among these traits are most likely negative or neutral, depending on the magnitude of trait covariation (Bergmann et al., 2020). Based on their globally high RTD and low SRL values (Freschet & Roumet, 2017), tree species are likely to be relying relatively strongly on mycorrhizal partners, therefore contributing less than other plant functional types to a trade-off between high SRL 'do-it-yourself' strategy and high root diameter 'outsourcing' strategy. Second, in addition to hydraulic- and mechanical-related functions, root diameter represents a multifunctional trait associating with both root-lifespan and fungal-symbiosis (Fig. 2; Mao et al., 2018; Jiang et al., 2020). The role of root diameter in root economics may thus result from the interplay between morphological and symbiotic root traits (Fig. 2). It has been reported that more than 80% of the variation in diameter of absorptive roots can be explained by CT in woody species (Gu et al., 2014; Kong et al., 2019). This pattern was also confirmed by the positive relationship between root diameter and CT (Fig. 4), indicating that within the data analysed here, root diameter tends to be more directly linked to symbiotic traits than to other morphological traits (i.e. SRL and RTD).

Although CT diverged from root diameter on the 'fungalsymbiosis' dimension for ECM species, a trade-off appeared between root diameter and BI (Figs 5e,f, S2), implying that

New Phytologist (2022)

www.newphytologist.com

thinner-rooted ECM species had more branching and vice versa. Ectomycorrhizal fungi have co-evolved divergent soil exploration and nutrient use strategies in complementarity with root morphologies (Agerer, 2001; Tedersoo & Smith, 2013). Ectomycorrhizal fungal groups with a long-distance exploration (i.e. long hyphae) are commonly associated with thicker root species to compensate for the deficiency in nutrient exploitation of these species (Liu et al., 2015). By contrast, ECM species with thinner roots and more branching tend to associate with short-distance soil exploring fungal taxa (i.e. those with short fungal hyphae), which demand less construction and maintenance costs (Chen et al., 2018). For instance, trees in the Pinaceae family have relatively thick roots and can associate with rhizomorphic hyphae that explore longer distance into the surrounding soil (Agerer, 2001). Consistently, ECM gymnosperms (i.e. Pinaceae in this study) had larger root diameters and lower BIs than ECM angiosperms (Table S9). The trade-off between root diameter and BI persisted when excluding gymnosperms from the ECM group (Table S5; Fig. S3), implying the robustness of this trade-off among ECM species. Overall, these findings jointly highlight that the type of mycorrhizal symbiosis acts as a critical driver to the complexity of root economics owing to the contrasting evolutionary history and symbiosis pathways between AM and ECM species.

The symbiosis intensity lies in the resource needs of the symbiotic partners, which is fundamentally mediated by the soil matrix. Plants can establish symbioses with mycorrhizal fungi to forage immobile soil P (van der Heijden et al., 2015; Liu et al., 2018). The tight linkages between the 'fungal-symbiosis' dimension and soil P availability for both AM and ECM species (Fig. 5d,f; Table S8) support that edaphic fertility drives, at least to some extent, the position of plant species within the multidimensional RES (Ding et al., 2020). These relationships were clearly dominated by symbiotic root traits, as revealed by the negative relationships of soil available P concentration with CT for AM species and with BI for ECM species (Table S7). Thus, soil P availability can shape the 'fungal-symbiosis' dimension of the RES for both mycorrhizal types. Soil nitrate availability presented similar correlations with this dimension for both AM and ECM species (Fig. 5d,f; Table S7). This synergy could relate to the close linkage between soil nitrate and P concentrations (Fig. 5) or highlight the importance of mycorrhizal fungi in plant acquisition of N (Hodge, 2004; Miller & Cramer, 2005; Treseder, 2013). We included species from subtropical and temperate forests where soil ammonium and nitrate availability varied greatly (Table S10). The young soils that originate from volcanic parent material in temperate forests typically have high levels of nitrate, while the older highly weathered soils in subtropical forests show higher levels of ammonium than nitrate (Table S10). Despite this, it seems that the dominant form of N has limited effect on the RES.

The 'root-lifespan' dimension of the RES was driven by soil P availability in ECM species but by soil nitrate availability in AM species (Fig. 5c; Table S8). The observed negative relationships between the 'root-lifespan' dimension and soil fertility suggest that plants tend to employ a conservative strategy by increasing cell wall fraction and thus RTD for extending root lifespan under

infertile habitats (Eissenstat & Yanai, 1997; Ding et al., 2020). The distinct controls of soil nitrate and P availability over the 'root-lifespan' dimension of AM and ECM species could be related to the contrasting nutrient economies between mycorrhizal types (Phillips et al., 2013). Different from the organicnutrient ECM species, AM species are assumed to specialize in inorganic nutrient uptake, given the limited saprotrophic ability of most AM fungi (Smith & Read, 2008). In addition, our results suggest that, independent of plant reliance on mycorrhizal symbiosis intensity, AM species may increasingly rely on thin nonmycorrhizal roots to acquire nitrate in conditions of high nitrate availability, whereas ECM species may rely more largely on mycorrhizal symbiosis. Although AM and ECM trees exhibit similar abilities to acquire mineral P, ECM fungi may have an advantage of immobile P acquisition over longer distances and from organic sources (Tedersoo & Bahram, 2019), which may offset the costly maintenance of the ECM structures (Phillips et al., 2013). One may thus expect that ECM species tend to construct roots with higher RTD to extend root lifespan and symbiotic partnership with soil fungi for maximizing P acquisition in P-deficient soils (Kramer-Walter et al., 2016; Kou et al., 2017). However, the relationships of edaphic fertility with morphological traits and the RES were overall weak (Fig. 5; Tables S7, S8), which may point towards the influence of other belowground interactions on the RES. For example, interspecific root interactions, root-pathogen-herbivore interactions and root phosphatase (Valverde-Barrantes et al., 2013; Bardgett et al., 2014; Han et al., 2022) may likewise shape the structure of the RES.

In conclusion, by elaborating on root trait-function linkage and fertility-driven root trait interplay, we demonstrate a bidimensional RES based on root functions: root-lifespan vs fungalsymbiosis, and provide a refined framework for theoretical and mechanistic understanding of the complexity of root economics within tree species. Irrespective of mycorrhizal type, the 'rootlifespan' dimension rendered by SRL and RTD depicts a resource acquisition-conservation trade-off representing the maintenance of root lifespan along the edaphic fertility gradient. Interestingly, the 'fungal-symbiosis' dimension characterized by the direct (BI and CT) and indirect (root diameter) symbiotic traits differs between AM and ECM species or even possibly between angiosperms and gymnosperms within ECM groups. These fundamental divergences have likely resulted from the contrasting evolutionary history and symbiosis pathways between mycorrhizal groups. Arbuscular fungi co-evolved with a large habitat dependence of a thicker root cortex, but increased root branching was prioritized for ECM angiosperms. Importantly, the availability of soil P and nitrate are key determinants for shaping the RES, particularly the 'fungal-symbiosis' dimension via potentially differentiating root-fungal symbiosis intensity among species. Formal testing of the hypothesis that increased root colonization by ECM fungi is related to higher root branching should therefore be a key priority of future research in this area. Overall, our findings suggest that root-fungal symbiosis pathways and soil fertility have jointly influenced root trait variation, which underpins the role of complex belowground interactions in framing root economics.

Acknowledgements

This research was financially supported by grants from the National Natural Science Foundation of China (nos. 31730014, 32071557, 41830646, 31700377). We are grateful to the editor Richard J. Norby and three anonymous reviewers for their help-ful comments and suggestions.

Author contributions

LK and HY conceived the ideas and designed the study. HY performed the experiments, collected the data, performed data analyses and drew the graphics. LK, HY and GTF led the writing of the manuscript. HW, JAH, SL, OJV-B, XF, RW, XD, LJ, SM, FY and MZ substantially contributed to writing the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

ORCID

Grégoire T. Freschet https://orcid.org/0000-0002-8830-3860 Xiaoli Fu https://orcid.org/0000-0001-9116-4745 James Aaron Hogan https://orcid.org/0000-0001-9806-3074 Liang Kou https://orcid.org/0000-0002-2187-0721 Oscar J. Valverde-Barrantes https://orcid.org/0000-0002-7327-7647 Ruili Wang https://orcid.org/0000-0002-0384-7739 Han Yan https://orcid.org/0000-0002-4578-8407

Data availability

All data supporting the findings of this study can be accessed in Dryad, doi: 10.5061/dryad.j0zpc869s (Yan *et al.*, 2022).

References

- Agerer R. 2001. Exploration types of ectomycorrhizae. *Mycorrhiza* 11: 107–114. Babikova Z, Gilbert L, Bruce TJA, Dewhirst SY, Pickett JA, Johnson D. 2014.
- Arbuscular mycorrhizal fungi and aphids interact by changing host plant quality and volatile emission. *Functional Ecology* **28**: 375–385.
- Bailey T, Robinson N, Farrell M, Macdonald B, Weaver T, Antille DL, Chin A, Brackin R. 2022. Storage of soil samples leads to overrepresentation of the contribution of nitrate to plant-available nitrogen. *Soil Research* 60: 22–32.
- Bardgett RD, Mommer L, De Vries FT. 2014. Going underground: root traits as drivers of ecosystem processes. Trends in Ecology & Evolution 29: 692–699.
- Bergmann J, Weigelt A, van der Plas F, Laughlin DC, Kuyper TW, Guerrero-Ramirez N, Valverde-Barrantes OJ, Bruelheide H, Freschet GT, Iversen CM *et al.* 2020. The fungal collaboration gradient dominates the root economics space in plants. *Science Advances* 6: eaba3756.
- Blomberg SP, Garland JRT, Ives AR. 2003. Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* 57: 717–745.
- Bloom AJ, Chapin FS III, Mooney HA. 1985. Resource limitation in plants an economic analogy. *Annual Review of Ecology and Systematics* 16: 363–392.
- Bonfante P, Genre A. 2010. Mechanisms underlying beneficial plant-fungus interactions in mycorrhizal symbiosis. *Nature Communications* 1: 1–11.
- Brundrett MC. 2002. Coevolution of roots and mycorrhizas of land plants. *New Phytologist* 154: 275–304.

New Phytologist

Brundrett MC. 2009. Mycorrhizal associations and other means of nutrition of vascular plants: understanding the global diversity of host plants by resolving conflicting information and developing reliable means of diagnosis. *Plant and Soil* **320:** 37–77.

Chen WL, Eissenstat DM, Koide RT. 2018. Root diameter predicts the extramatrical hyphal exploration distance of the ectomycorrhizal fungal community. *Ecosphere* 9: e02202.

Comas LH, Callahan HS, Midford PE. 2014. Patterns in root traits of woody species hosting arbuscular and ectomycorrhizas: implications for the evolution of belowground strategies. *Ecology and Evolution* 4: 2979–2990.

Comas LH, Eissenstat DM. 2009. Patterns in root trait variation among 25 coexisting North American forest species. *New Phytologist* 182: 919–928.

Ding JX, Kong DL, Zhang ZL, Cai Q, Xiao J, Liu Q, Yin HJ. 2020. Climate and soil nutrients differentially drive multidimensional fine root traits in ectomycorrhizal-dominated alpine coniferous forests. *Journal of Ecology* 108: 2544–2556.

Eissenstat DM, Wells CE, Yanai RD, Whitbeck JL. 2000. Building roots in a changing environment: implications for root longevity. *New Phytologist* 147: 33–42.

Eissenstat DM, Yanai RD. 1997. The ecology of root lifespan. Advances in Ecological Research 27: 1–60.

Freckleton RP, Harvey PH, Pagel M. 2002. Phylogenetic dependence and ecological data: a test and review of evidence. *The American Naturalist* 160: 716–726.

Freschet GT, Pagès L, Iversen CM, Comas LH, Rewald B, Roumet C, Klimešová J, Zadworny M, Poorter H, Postma JA. 2021a. A starting guide to root ecology: strengthening ecological concepts and standardising root classification, sampling, processing and trait measurements. *New Phytologist* 232: 973–1122.

Freschet GT, Roumet C. 2017. Sampling roots to capture plant and soil functions. *Functional Ecology* 31: 1506–1518.

Freschet GT, Roumet C, Comas LH, Weemstra M, Bengough AG, Rewald B, Bardgett RD, De Deyn GB, Johnson D, Klimešová J et al. 2021b. Root traits as drivers of plant and ecosystem functioning: current understanding, pitfalls and future research needs. *New Phytologist* 232: 1123–1158.

Freschet GT, Violle C, Bourget MY, Scherer-Lorenzen M, Fort F. 2018. Allocation, morphology, physiology, architecture: the multiple facets of plant above-and below-ground responses to resource stress. *New Phytologist* 219: 1338–1352.

Genre A, Lanfranco L, Perotto S, Bonfante P. 2020. Unique and common traits in mycorrhizal symbioses. *Nature Reviews Microbiology* 18: 649–660.

Gu JC, Xu Y, Dong XY, Wang HF, Wang ZQ. 2014. Root diameter variations explained by anatomy and phylogeny of 50 tropical and temperate tree species. *Tree Physiology* **34**: 415–425.

Guerrero-Ramírez NR, Mommer L, Freschet GT, Iversen CM, McCormack ML, Kattge J, Poorter H, van der Plas F, Bergmann J, Kuyper TW. 2021. Global root traits (GRooT) database. *Global Ecology and Biogeography* 30: 25– 37.

Guo DL, Mitchell RJ, Withington JM, Fan PP, Hendricks JJ. 2008a. Endogenous and exogenous controls of root life span, mortality and nitrogen flux in a longleaf pine forest: root branch order predominates. *Journal of Ecology* 96: 737–745.

Guo DL, Xia MX, Wei X, Chang WJ, Liu Y, Wang ZQ. 2008b. Anatomical traits associated with absorption and mycorrhizal colonization are linked to root branch order in twenty-three Chinese temperate tree species. *New Phytologist* **180**: 673–683.

Han MG, Chen Y, Li R, Yu M, Fu LC, Li SF, Su JR, Zhu B. 2022. Root phosphatase activity aligns with the collaboration gradient of the root economics space. *New Phytologist* 234: 873–849.

Han MG, Zhu B. 2021. Linking root respiration to chemistry and morphology across species. *Global Change Biology* 27: 190–201.

van der Heijden MGA, Martin FM, Selosse MA, Sanders IR. 2015. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* 205: 1406–1423.

Hodge A. 2004. The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytologist* 162: 9–24.

Iversen CM, McCormack ML, Powell AS, Blackwood CB, Freschet GT, Kattge J, Roumet C, Stover DB, Soudzilovskaia NA, Valverde-Barrantes OJ et al. 2017. A global Fine-Root Ecology Database to address below-ground challenges in plant ecology. *New Phytologist* 215: 15–26.

Jiang L, Wang HM, Li SG, Fu XL, Dai XQ, Yan H, Kou L. 2021. Mycorrhizal and environmental controls over root trait-decomposition linkage of woody trees. *New Phytologist* 229: 284–295.

Jiang PP, Wang HM, Meinzer FC, Kou L, Dai XQ, Fu XL. 2020. Linking reliance on deep soil water to resource economy strategies and abundance among coexisting understorey shrub species in subtropical pine plantations. *New Phytologist* 225: 222–233.

Jin Y, Qian H. 2019. V.PHYLOMAKER: an R package that can generate very large phylogenies for vascular plants. *Ecography* 42: 1353–1359.

Jung SC, Martinez-Medina A, Lopez-Raez JA, Pozo MJ. 2012. Mycorrhizainduced resistance and priming of plant defenses. *Journal of Chemical Ecology* 38: 651–664.

Kong DL, Ma CE, Zhang Q, Li L, Chen XY, Zeng H, Guo DL. 2014. Leading dimensions in absorptive root trait variation across 96 subtropical forest species. *New Phytologist* 203: 863–872.

Kong DL, Wang JJ, Valverde-Barrantes OJ, Kardol P. 2021. A framework to assess the carbon supply-consumption balance in plant roots. *New Phytologist* 229: 659–664.

Kong DL, Wang JJ, Wu HF, Valverde-Barrantes OJ, Wang RL, Zeng HY, Kardol P, Zhang H, Feng YL. 2019. Nonlinearity of root trait relationships and the root economics spectrum. *Nature Communications* 10: 2203.

Kottke I, Guttenberger M, Hampp R, Oberwinkler F. 1987. An *in vitro* method for establishing mycorrhizae on coniferous tree seedlings. *Trees* 1: 191–194.

Kou L, Jiang L, Fu XL, Dai XQ, Wang HM, Li SG. 2018. Nitrogen deposition increases root production and turnover but slows root decomposition in *Pinus elliottii* plantations. *New Phytologist* 218: 1450–1461.

Kou L, Li SG, Wang HM, Fu XL, Dai XQ. 2019. Unaltered phenology but increased production of ectomycorrhizal roots of *Pinus elliottii* under 4 years of nitrogen addition. *New Phytologist* 221: 2228–2238.

Kou L, McCormack ML, Chen WW, Guo DL, Wang HM, Gao WL, Yang H, Li SG. 2017. Nitrogen ion form and spatio-temporal variation in root distribution mediate nitrogen effects on lifespan of ectomycorrhizal roots. *Plant* and Soil 411: 261–273.

Kramer-Walter KR, Bellingham PJ, Millar TR, Smissen RD, Richardson SJ, Laughlin DC. 2016. Root traits are multidimensional: specific root length is independent from root tissue density and the plant economic spectrum. *Journal* of Ecology 104: 1299–1310.

Langley JA, Hungate BA. 2003. Mycorrhizal controls on belowground litter quality. *Ecology* 84: 2302–2312.

Laughlin DC, Mommer L, Sabatini FM, Bruelheide H, Kuyper TW, McCormack ML, Bergmann J, Freschet GT, Guerrero-Ramírez NR, Iversen CM et al. 2021. Root traits explain plant species distributions along climatic gradients yet challenge the nature of ecological trade-offs. Nature Ecology & Evolution 5: 1123–1134.

Liese R, Alings K, Meier IC. 2017. Root branching is a leading root trait of the plant economics spectrum in temperate trees. *Frontiers in Plant Science* 8: 315.

Liu BT, Li HB, Zhu B, Koide RT, Eissenstat DM, Guo DL. 2015. Complementarity in nutrient foraging strategies of absorptive fine roots and arbuscular mycorrhizal fungi across 14 coexisting subtropical tree species. *New Phytologist* 208: 125–136.

Liu XB, Burslem DFRP, Taylor JD, Taylor AFS, Khoo E, Majalap-Lee N, Helgason T, Johnson D. 2018. Partitioning of soil phosphorus among arbuscular and ectomycorrhizal trees in tropical and subtropical forests. *Ecology Letters* 21: 713–723.

Ma ZQ, Guo DL, Xu XL, Lu MZ, Bardgett RD, Eissenstat DM, McCormack ML, Hedin LO. 2018. Evolutionary history resolves global organization of root functional traits. *Nature* 555: 94–97.

- Mao Z, Wang Y, McCormack ML, Rowe N, Deng X, Yang X, Xia S, Nespoulous J, Sidle RC, Guo D *et al.* 2018. Mechanical traits of fine roots as a function of topology and anatomy. *Annals of Botany* 122: 1103–1116.
- Martin F, Kohler A, Murat C, Veneault-Fourrey C, Hibbett DS. 2016. Unearthing the roots of ectomycorrhizal symbioses. *Nature Reviews Microbiology* 14: 760–773.
- McCormack ML, Adams TS, Smithwick EAH, Eissenstat DM. 2012. Predicting fine root lifespan from plant functional traits in temperate trees. *New Phytologist* **195**: 823–831.
- McCormack ML, Dickie IA, Eissenstat DM, Fahey TJ, Fernandez CW, Guo DL, Helmisaari HS, Hobbie EA, Iversen CM, Jackson RB. 2015. Redefining fine roots improves understanding of below-ground contributions to terrestrial biosphere processes. *New Phytologist* 207: 505–518.
- McCormack ML, Guo DL. 2014. Impacts of environmental factors on fine root lifespan. *Frontiers in Plant Science* 5: 205.
- Miller AJ, Cramer MD. 2005. Root nitrogen acquisition and assimilation. *Plant and Soil* 274: 1–36.
- Münkemüller T, Lavergne S, Bzeznik B, Dray S, Jombart T, Schiffers K, Thuiller W. 2012. How to measure and test phylogenetic signal. *Methods in Ecology and Evolution* 3: 743–756.
- Orme D, Freckleton R, Thomas G, Petzoldt T, Fritz S, Isaac N, Pearse W. 2013. The CAPER package: comparative analysis of phylogenetics and evolution in R. R package v.5:1-36. [WWW document] URL https://CRAN.R-project.org/ package=caper [accessed 16 April 2018].
- Ostonen I, Lõhmus K, Helmisaari HS, Truu J, Meel S. 2007. Fine root morphological adaptations in Scots pine, Norway spruce and silver birch along a latitudinal gradient in boreal forests. *Tree Physiology* 27: 1627–1634.
- **Paradis E, Schliep K. 2019.** APE 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* **35**: 526–528.
- Phillips RP, Brzostek E, Midgley MG. 2013. The mycorrhizal-associated nutrient economy: a new framework for predicting carbon-nutrient couplings in temperate forests. *New Phytologist* 199: 41–51.
- Pregitzer KS, Deforest JL, Burton AJ, Allen MF, Ruess RW, Hendrick RL. 2002. Fine root architecture of nine North American trees. *Ecological Monographs* 72: 293–309.
- R Core Team. 2020. R: a language and environment for statistical computing. Vienna, Austria: Foundation for Statistical Computing. [WWW document] URL http://www.R-project.org/ [accessed 29 February 2020].
- Raudaskoski M, Kothe E. 2015. Novel findings on the role of signal exchange in arbuscular and ectomycorrhizal symbioses. *Mycorrhiza* 25: 243–252.
- Reich PB. 2014. The world-wide 'fast-slow' plant economics spectrum: a traits manifesto. *Journal of Ecology* 102: 275–301.
- Revell LJ, Collar DC. 2009. Phylogenetic analysis of the evolutionary correlation using likelihood. *Evolution* 63: 1090–1100.
- Ryser P. 1996. The importance of tissue density for growth and life span of leaves and roots: a comparison of five ecologically contrasting grasses. *Functional Ecology* 10: 717–723.
- Smith SA, Brown JW. 2018. Constructing a broadly inclusive seed plant phylogeny. American Journal of Botany 105: 302–314.
- Smith SE, Read DJ. 2008. *Mycorrhizal symbiosis*, 3rd edn. London, UK: Academic Press.
- Steidinger BS, Crowther TW, Liang J, Van Nuland ME, Werner GDA, Reich PB, Nabuurs GJ, de-Miguel S, Zhou M, Picard N et al. 2019. Climatic controls of decomposition drive the global biogeography of forest-tree symbioses. *Nature* 569: 404–408.
- Strullu-Derrien C, Selosse MA, Kenrick P, Martin FM. 2018. The origin and evolution of mycorrhizal symbioses: from palaeomycology to phylogenomics. *New Phytologist* 220: 1012–1030.
- Tedersoo L, Bahram M. 2019. Mycorrhizal types differ in ecophysiology and alter plant nutrition and soil processes. *Biological Reviews* 94: 1857–1880.
- Tedersoo L, Bahram M, Zobel M. 2020. How mycorrhizal associations drive plant population and community biology. *Science* 367: eaba1223.
- Tedersoo L, Brundrett MC. 2017. Evolution of ectomycorrhizal symbiosis in plants. In: Tedersoo L, ed. *Biogeography of mycorrhizal symbiosis*. Cham, Switzerland: Springer International, 407–467.

- Tedersoo L, Smith ME. 2013. Lineages of ectomycorrhizal fungi revisited: foraging strategies and novel lineages revealed by sequences from belowground. *Fungal Biology Reviews* 27: 83–99.
- The Angiosperm Phylogeny Group, Chase MW, Christenhusz MJM, Fay MF, Byng JW, Judd WS, Soltis DE, Mabberley DJ, Sennikov AN, Soltis PS et al. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. Botanical Journal of the Linnean Society 181: 1–20.
- **Treseder KK. 2013.** The extent of mycorrhizal colonization of roots and its influence on plant growth and phosphorus content. *Plant and Soil* 371: 1–13.
- Uyeda JC, Caetano DS, Pennell MW. 2015. Comparative analysis of principal components can be misleading. *Systematic Biology* 64: 677–689.
- Valverde-Barrantes OJ, Freschet GT, Roumet C, Blackwood CB. 2017. A worldview of root traits: the influence of ancestry, growth form, climate and mycorrhizal association on the functional trait variation of fine-root tissues in seed plants. *New Phytologist* 215: 1562–1573.
- Valverde-Barrantes OJ, Smemo KA, Blackwood CB. 2015. Fine root morphology is phylogenetically structured, but nitrogen is related to the plant economics spectrum in temperate trees. *Functional Ecology* 29: 796–807.
- Valverde-Barrantes OJ, Smemo KA, Feinstein LM, Kershner MW, Blackwood CB. 2013. The distribution of below-ground traits is explained by intrinsic species differences and intraspecific plasticity in response to root neighbours. *Journal of Ecology* 101: 933–942.
- Wambsganss J, Freschet GT, Beyer F, Goldmann K, Prada-Salcedo LD, Scherer-Lorenzen M, Bauhus J. 2021. Tree species mixing causes a shift in fine-root soil exploitation strategies across European forests. *Functional Ecology* 35: 1886–1902.
- Weemstra M, Kiorapostolou N, van Ruijven J, Mommer L, de Vries J, Sterck F. 2020. The role of fine-root mass, specific root length and life span in tree performance: a whole-tree exploration. *Functional Ecology* 34: 575–585.
- Weemstra M, Mommer L, Visser EJ, van Ruijven J, Kuyper TW, Mohren GM, Sterck FJ. 2016. Towards a multidimensional root trait framework: a tree root review. *New Phytologist* 211: 1159–1169.
- Weigelt A, Mommer L, Andraczek K, Iversen CM, Bergmann J, Bruelheide H, Fan Y, Freschet GT, Guerrero-Ramírez NR, Kattge J et al. 2021. An integrated framework of plant form and function: the belowground perspective. *New Phytologist* 232: 42–59.
- Wen ZH, Li HB, Shen Q, Tang XM, Xiong CY, Li HG, Pang JY, Ryan MH, Lambers H, Shen JB. 2019. Tradeoffs among root morphology, exudation and mycorrhizal symbioses for phosphorus-acquisition strategies of 16 crop species. *New Phytologist* 223: 882–895.
- Withington JM, Reich PB, Oleksyn J, Eissenstat DM. 2006. Comparisons of structure and life span in roots and leaves among temperate trees. *Ecological Monographs* 76: 381–397.
- Wright IJ, Reich PB, Westoby M, Ackerly DD, Baruch Z, Bongers F, Cavender-Bares J, Chapin T, Cornelissen JHC, Diemer M et al. 2004. The worldwide leaf economics spectrum. *Nature* 428: 821–827.
- Yan H, Freschet GT, Wang HM, Hogan JA, Li SG, Valverde-Barrantes OJ, Fu XL, Wang RL, Dai XQ, Jiang L *et al.* 2022. Data from: Mycorrhizal symbiosis pathway and edaphic fertility frame root economics space among tree species. *Dryad Dataset.* doi: 10.5061/dryad.j0zpc869s.
- Yan H, Kou L, Wang HM, Fu XL, Dai XQ, Li SG. 2019. Contrasting root foraging strategies of two subtropical coniferous forests under an increased diversity of understory species. *Plant and Soil* 436: 427–438.
- Yu GC, Smith DK, Zhu HC, Guan Y, Lam TT-Y. 2017. GGTREE: an R package for visualization and annotation of phylogenetic trees with their covariates and other associated data. *Methods in Ecology and Evolution* 8: 28–36.
- Zanne AE, Tank DC, Cornwell WK, Eastman JM, Smith SA, FitzJohn RG, McGlinn DJ, O'Meara BC, Moles AT, Reich PB *et al.* 2014. Three keys to the radiation of angiosperms into freezing environments. *Nature* 506: 89–92.
- Zhou M, Bai WM, Zhang YS, Zhang WH. 2018. Multi-dimensional patterns of variation in root traits among coexisting herbaceous species in temperate steppes. *Journal of Ecology* 106: 2320–2331.

Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

Fig. S1 Pairwise correlations among seven root economics traits for arbuscular mycorrhizal species.

Fig. S2 Pairwise correlations among seven root economics traits for ectomycorrhizal species.

Fig. S3 Root economics pattern for ectomycorrhizal angiosperms.

Fig. S4 The test of the root economics space using the four traits RD, RTD, SRL and RN in Bergmann *et al.* (2020).

Fig. S5 Density histograms of woody species root trait distribution of our own data on absorptive roots as compared to these of fine root (diameter $\leq 2 \text{ mm}$) traits from the GRooT database.

Table S1 Species information in the study.

Table S2 Loading scores of phylogenetically informed principal component analysis of root traits for all species, arbuscular mycorrhizal species and ectomycorrhizal species based on the corrected root tissue density data (carbon basis).

Table S3 Summary of the seven root traits used in this studybased on the GRooT database.

Table S4 Blomberg's *K* value of root traits for arbuscular mycorrhizal (n=77) and ectomycorrhizal (n=35) species.

Table S5 Results of phylogenetically informed principal component analysis of root traits for all species, arbuscular mycorrhizal species, ectomycorrhizal (ECM) species and ECM angiosperms.

Table S6 One-way ANOVA analysis on the effect of mycorrhizal type on scores of 'fungal-symbiosis' dimension of 112 tree species.

Table S7 Coefficients of Pearson's correlation for linkages of root traits with edaphic nutrients for all species, arbuscular mycorrhizal species and ectomycorrhizal species.

Table S8 Results of the general linear regression between princi-pal component scores and edaphic nutrients.

Table S9 Student's *t*-test evaluating the differences in root traits between angiosperms (n=28) and gymnosperms (n=7) of ectomycorrhizal species.

Table S10 Student's *t*-test evaluating differences in soil nutrient availability between species from subtropical (n=64) and temperate (n=48) forests.

Please note: Wiley Blackwell is not responsible for the content or functionality of any Supporting Information supplied by the authors. Any queries (other than missing material) should be directed to the *New Phytologist* Central Office.