



Ecosystem Health and Sustainability

ISSN: (Print) (Online) Journal homepage: https://www.tandfonline.com/loi/tehs20

Modeling methane dynamics in three wetlands in Northeastern China by using the CLM-Microbe model

Yunjiang Zuo, Yihui Wang, Liyuan He, Nannan Wang, Jianzhao Liu, Fenghui Yuan, Kexin Li, Ziyu Guo, Ying Sun, Xinhao Zhu, Lihua Zhang, Changchun Song, Li Sun & Xiaofeng Xu

To cite this article: Yunjiang Zuo, Yihui Wang, Liyuan He, Nannan Wang, Jianzhao Liu, Fenghui Yuan, Kexin Li, Ziyu Guo, Ying Sun, Xinhao Zhu, Lihua Zhang, Changchun Song, Li Sun & Xiaofeng Xu (2022) Modeling methane dynamics in three wetlands in Northeastern China by using the CLM-Microbe model, Ecosystem Health and Sustainability, 8:1, 2074895, DOI: 10.1080/20964129.2022.2074895

To link to this article: <u>https://doi.org/10.1080/20964129.2022.2074895</u>



Taylor & Francis Taylor & Francis Group

OPEN ACCESS Check for updates

Modeling methane dynamics in three wetlands in Northeastern China by using the CLM-Microbe model

Yunjiang Zuo^{a,b}, Yihui Wang^c, Liyuan He^c, Nannan Wang^a, Jianzhao Liu^{a,b}, Fenghui Yuan^{d,e}, Kexin Li^{a,b}, Ziyu Guo^a, Ying Sun^a, Xinhao Zhu^a, Lihua Zhang^f, Changchun Song^a, Li Sun^a and Xiaofeng Xu^c

^aKey Laboratory of Wetland Ecology and Environment, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun, Jilin, China; ^bUniversity of the Chinese Academy of Sciences, Beijing, China; ^cBiology Department, San Diego State University, San Diego, CA, USA; ^dKey Laboratory of Forest Ecology and Management, Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang, Liaoning, China; ^eDepartment of Soil, Water, and Climate, University of Minnesota, Saint Paul, MN, USA; ^fCollege of Life and Environmental Sciences, Minzu University of China, Beijing, China

ABSTRACT

Wetlands account for up to 70% of the natural source of methane (CH_4) in terrestrial ecosystems on a global scale. Soil microbes are the ultimate producers and biological consumers of CH₄ in wetlands. Therefore, simulating microbial mechanisms of CH₄ production and consumptionwould improve the predictability of CH₄ flux in wetland ecosystems. In this study, we applied a microbial-explicit model, the CLM-Microbe, to simulate CH₄ flux in three major natural wetlands in northeastern China. The CLM-Microbe model was able to capture the seasonal variation of gross primary productivity (GPP), dissolved organic carbon (DOC), and CH₄ flux. The CLM-Microbe model explained more than 40% of the variation in GPP and CH₄ flux across sites. Marsh wetlands had higher CH₄ flux than mountain peatlands. Ebullition dominated the CH₄ transport pathway in all three wetlands. The methanogenesis dominates while methanotroph makes a minor contribution to the CH4 flux, making all wetlands a CH4 source. Sensitivity analysis indicated that microbial growth and death rates are the key factors governing CH₄ emission and vegetation physiological properties (flnr) and maintenance respiration predominate GPP variation. Explicitly simulating microbial processes allows genomic information to be incorporated, laying a foundation for better predicting CH₄ dynamics under the changing environment.

Introduction

Natural wetlands store approximately 30% of soil carbon (C) on the land (Melton et al. 2013); those C will be released into the atmosphere either as methane (CH₄) or CO₂, depending on the soil redox potential (Le and Jean 2001). Wetlands roughly account for one-third of the increasing atmospheric CH₄ concentration (Bhullar et al. 2014; Bridgham et al. 2013; Saunois et al. 2020). CH₄, a potent greenhouse gas, has a global warming potential 28 times larger than carbon dioxide (CO₂) (Bridgham et al. 2013; Harmsen et al. 2020; Schaefer 2019). The rising CH₄ concentration contributed to 22% of climate warming caused by anthropogenic activities since the Industrial Revolution (IPCC 2017). However, the estimates of CH₄ emissions remain highly uncertain (Saunois et al., 2020), particularly in natural wetlands (Jackson et al., 2020). From a global perspective, natural wetlands are the largest and most uncertain source of atmospheric CH_4 (Bousquet et al. 2006a; Kirschke et al. 2013a; Zhang et al. 2017a).

The land surface CH₄ flux depends on the balance of microbial methanogenesis and methanotrophy (Fazli, Man, and Shah 2013). When there is a lack of electron acceptors (oxygen, nitrate, sulfate, iron, and manganese), anaerobic fermentation will occur, and methanogens will use fermentation products (hydrogen and CO₂, acetic acid, and methyl compounds) to produce CH₄ (Cordruwisch, Seitz, and Conrad 1988; McGlynn 2017; Sieber, McInerney, and Gunsalus 2012; Thauer et al. 2008; Timmers et al. 2017). Under aerobic conditions, CH₄ is oxidized to CO₂ by methanotrophs. Both processes occur and determine the direction of CH₄ flux. Although these processes have been wellunderstood, microbial models with explicit representation of methanogenesis and methanotrophy are still in their infancy (Xiaofeng et al. 2015; Xu et al., 2016). In addition, the ecosystem research on modeling wetland CH₄ processes in China is rare (Sun et al. 2018; Wang et al., 2019; Xiaofeng and Tian 2012).

CONTACT Li Sun 🐼 sunli@iga.ac.cn;xxu@sdsu.edu 🗈 Key Laboratory of Wetland Ecology and Environment, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Changchun 130102, Jilin, China

© 2022 The Author(s). Published by Taylor & Francis Group and Science Press on behalf of the Ecological Society of China.

ARTICLE HISTORY Received 31 December 2021 Revised 28 April 2022

Accepted 03 May 2022 **KEYWORDS** CH₄ flux; microbe; CLM-Microbe model; wetlands

This article has been republished with minor changes. These changes do not impact the academic content of the article.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Wetland ecosystem models can be categorized as empirical models and process-based models. The empirical models are primarily based on empirical equations obtained from a large amount of data to make predictions. The observed CO₂ and CH₄ fluxes are directly related to environmental factors such as groundwater level, soil temperature, and net primary productivity (NPP) (Zhang et al. 2017a). However, due to the lack of mechanistic representation of the CH₄ cycle in these empirical models, there are usually large discrepancies between the measured and simulated flux when the models are used in different times and regions (Huang, Sass, and Fisher 1998; Meng et al. 2012; Morin et al. 2014; Wania et al. 2013), which has restricted the popularization and application of these models (Xu et al., 2016). The process-based models consider the major biogeochemical processes of CH₄ cycling to achieve accurate predictability of CH₄ cycling within a specific ecosystem (Kirschke et al. 2013; Hangin et al. 2016). In recent years, some CH₄ models have been developed and applied to simulate CH₄ cycling in natural wetlands. For example, Lipson et al. (2012) used the DLEM model to explore the CH₄ exchange between the atmosphere and marshland over China. Tingting et al. (2020) verified the application of the CH4MOD model in the natural wetlands of the Sanjiang Plain. Zhang et al. (2020a) used the TRIPLEX-GHG model to simulate the spatial pattern of CH₄ emissions in the Qinghai-Tibet Plateau. Additionally, Lund-Potsdam-Jena Wetland Hydrology and Methane (LPJ-WHyMe), Terrestrial Ecosystem Model (TEM), Denitrification-Decomposition model (DNDC), and other models have been applied to simulate wetland CH₄ fluxes in the Arctic, Europe, and the world (Chadburn et al. 2020; Tingting et al. 2020; Melton et al. 2013; Nzotungicimpaye et al. 2020; Youmi et al. 2020; Wania, Ross, and Prentice 2010).

The CLM-Microbe is a microbial-explicit model that simulates substrates, production, oxidation, transport of CH₄ and their environmental controls, along with microbial mechanisms for soil C mineralization (Xiaofeng et al. 2014, 2015; Yihui et al. 2019). A key advantage is that it represents multiple microbial functional groups. In this study, we used the CLM-Microbe model to simulate GPP and CH₄ flux in three wetlands. There are three objectives of this study: (1) to evaluate the performance of the CLM-Microbe model in simulating CH₄ flux in wetlands of northeastern China; (2) to investigate the differences in CH₄ cycling among three major wetlands; (3) to identify the key factors controlling on GPP and CH₄ flux among the wetland types.

Methodology

Site description

This study applied the model to three wetland ecosystems in northeast China, including marshland in the Sanjiang Plain (47.58°N, 133.52°E), peatland in the Changbai Mountain (42.35°N, 126.38°E), and the swamp in the Lesser Khingan Mountain (48.16°N, 128.5°E). These three sites represent two major wetland types in northeastern China, freshwater marshes on the lowland<apos;>s plains (Sanjiang Plain) and mountain peatlands (Changbai Mountain and Lesser Khingan Mountain).

The Sanjiang Plain features the largest freshwater wetland in China. It has a continental monsoon climate, with a mean annual temperature of 2.52°C and a mean annual precipitation of 558 mm. The wetland in the Changbai Mountain has a continental monsoon climate, with a mean annual temperature of 3.3°C and mean annual precipitation of 1054 mm. The precipitation is primarily occurring during July-August. The early spring snowfall and precipitation lead to a water table of 10 cm. The dominant plant community in the wetland is Carex sphagnum. There is huge Holocene peat stored in the wetland, with a thickness of 4-5 m. The upper part is mainly moss peat, and a frozen layer is formed in winter. The Lesser Khingan Mountain is located between Greater Khingan Mountain and Changbai Mountain and is one of the major areas with swamps in China. This site has a continental humid monsoon climate, with a mean annual temperature of 0.4°C and mean annual precipitation of 630 mm (primarily occurring during July-August). It primarily consists of forest swamps, shrub swamps, grass swamps, and moss swamps.

Data sources

The CH₄ flux was monitored using the close-path eddy covariance technique and static chamber approach. Flux measurement periods were from May to October in 2012 and 2013 at the Sanjiang Plain (Table 1) (Sun et al. 2018). The CH₄ flux for the Changbai Mountain and Lesser Khingan Mountain was obtained through periodic sampling by a static chamber approach (Huang 2016; Shi 2019). We extracted CH₄ flux data for these two sites from the published articles by software GetData version 2.26 (http://getdata-graphdigitizer.com/). The observational GPP data of Sanjiang Plain and Changbai Mountain are calculated with net ecosystem C exchange (NEE) and ecosystem respiration (ER) (Cao 2015.). The GPP data of Lesser Khingan Mountain is obtained by extracting the GPP data of MODIS (MOD17A) products (Yuan et al. 2020; Maosheng et al. 2005).

Modeling experiment

Model description and driving forces

The CLM-Microbe model branches from the framework of default CLM4.5 by developing a new microbial functional group-based module for CH₄ production and

Sites	Data	Year	Source				
Sanjiang Plain	Atmospheric forcing data	2005-2018	Sanjiang Plain Marsh Wetland Ecological Experimental Station				
	GPP	2012-2013	(http://sjm.cern.ac.cn/meta/metaData)				
	CH4	2012-2013					
	DOC	2013-2014	Xiaofeng et al. (2015)				
Changbai Mountains	Atmospheric forcing data	2003-2018	China Meteorological Forcing Dataset				
			(https://data.tpdc.ac.cn/en/data/8028b944-daaa-4511-8769-965612652c49/)				
	GPP	2011	Cao (2015.)				
	CH4	2016	Shi (2019)				
	DOC	2017	Han et al. (2018)				
Lesser Khingan Mountains	Atmospheric forcing data	2010-2018	China Meteorological Forcing Dataset				
			(https://data.tpdc.ac.cn/en/data/8028b944-daaa-4511-8769-965612652c49/)				
	GPP	2010-2011	MODIS				
			(https://lpdaac.usgs.gov/products/mod17a2hv006/)				
	CH₄	2013-2014	Huang (2016)				
	DOC	2014	Xu et al., (2016)				

Table 1. Atmospheric forcing data and flux data source.

consumption (Xiaofeng et al. 2015), in association with the decomposition subroutines in CLM4.5 (Koven et al. 2013; Thornton et al. 2007). It incorporates new mechanisms of DOC fermentation, hydrogenotrophic methanogenesis, acetoclastic methanogenesis, aerobic methanotrophy, anaerobic methanotrophs, and H₂ production (Xiaofeng et al. 2015; Yihui et al. 2019). Detailed mathematical expressions for CH₄ production and consumption processes were organized in Xiaofeng et al. (2015) and Yihui et al. (2019), Wang et al. (2022)). The processes of microbial assimilation of C can be referred to in Xiaofeng et al. (2014) and Liyuan et al. (2021a), Liyuan et al. (2021b)). The code for the CLM-Microbe model is archived at Github (https:// github.com/email-clm/clm-microbe). The model version used in this study was checked out from GitHub on 18 June 2018.

In our previous study, the CLM-Microbe model was validated for simulating the dynamics of CO₂ and CH₄ emissions from incubation experiments on Arctic soils with invariant soil temperature and soil water content (Xiaofeng et al. 2015). In addition, the CLM-Microbe model was applied to examine the microtopographic impacts on CO₂ and CH₄ flux in the Arctic tundra ecosystem (Yihui et al. 2019) and microbial seasonality on soil C cycling in terrestrial ecosystems (He et al., 2021b). In this study, we focused on the fully incorporated CLM-Microbe model, with model simulations for each wetland type. Eighteen key parameters were chosen for model parameterization, which represents the decomposition of organic C, methanogenesis, microbial growth, and plant photosynthesis and respiration and therefore controls the GPP and CH₄ flux (Table 2).

The model driving forces include meteorological, edaphic, and vegetation datasets. The meteorological data include air temperature, relative humidity, incoming solar radiation, longwave radiation, precipitation rate, surface pressure, and surface winds. Meteorological variables of Changbai Mountain (2003– 2018) and Lesser Khingan Mountain (2010–2018) were extracted from China<apos;>s Meteorological Forcing Dataset (https://data.tpdc.ac.cn/en/data/8028b944daaa-4511-8769-965612652c49/) by the longitude and latitude information. Since the standardized forcing data are in half-hourly time steps, the extracted 3-hourly data for each study site was interpolated to half-hourly step by using linear interpolation in R programming (R for Window version 4.0.2). The forcing data of Sanjiang Plain marsh wetland (2005–2018) was from the Sanjiang Plain Experimental station (Table 1). The observed soil and vegetation variables, such as plant functional type (PFT), were used for model parameterization and validation.

Model implementation

In this study, we set up model simulations with the CLM-Microbe model separately for each site. The model implementation was carried out in three stages. First, the accelerated model spin-up was set up for 2,000 years to allow the system to accumulate

Table 2. Key parameters for sensitivity analysis (see Xiaofeng et al. 2015).

ParameterEcological meaningKACEHalf-saturation coefficient of available carbon mineralizationAceProdACmaxMaximum rate of acetate production from available carbonYAceMethanogensGrowth efficiency of acetoclastic methanogensYH2MethanogensGrowth efficiency of H2-CO2-dependent methanogensYMethanotrophyGrowth efficiency of acetoclastic methanogensYMethanotrophyGrowth efficiency of acetoclastic methanogensYMethanotrophyGrowth efficiency of acetoclastic methanogensDeadRAceMethanogensDeath rate of acetoclastic methanogensDeadRH2MethanogensDeath rate of acetoclastic methanogensDeadRH2MethanogensDeath rate of H2-CO2-dependent methanogensGrowRmethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophBulk denitrificationDeath rate of aerobic methanotrophBulk denitrificationBulk denitrificationbdrnBulk denitrificationFroot_leafNew fine root C per new leaf CK_domDecomposition rate constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter						
KACEHalf-saturation coefficient of available carbon mineralizationAceProdACmaxMaximum rate of acetate production from available carbonYAceMethanogensGrowth efficiency of acetoclastic methanogensYH2MethanogensGrowth efficiency of H2-CO2-dependent methanogensYMethanotrophyGrowth efficiency of acetoclastic methanogensGrowRAceMethanogensGrowth rate of acetoclastic methanogensDeadRAceMethanogensDeath rate of acetoclastic methanogensDeadRAceMethanogensDeath rate of H2-CO2-dependent methanogensDeadRH2MethanogensDeath rate of H2-CO2-dependent methanogensGrowRmethanotrophsGrowth rate of H2-CO2-dependent methanogensDeadRMethanotrophsGrowth rate of aerobic methanotrophGrowRmethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophBulk denitrificationBulk denitrificationbr_mrBase rate of maintenance respirationFroot_leafNew fine root C per new leaf C M2 for maintenance constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter	Parameter	Ecological meaning				
AceProdACmaxMaximum rate of acetate production from available carbonYAceMethanogensGrowth efficiency of acetoclastic methanogensYH2MethanogensGrowth efficiency of H2-CO2-dependent methanogensYMethanotrophyGrowth efficiency of aerobic methanotroph Growth efficiency of acetoclastic methanogensYMethanotrophyGrowth efficiency of aerobic methanotroph Growth rate of acetoclastic methanogensDeadRAceMethanogensDeath rate of acetoclastic methanogensDeadRH2MethanogensDeath rate of H2-CO2-dependent methanogensDeadRH2MethanogensDeath rate of aerobic methanotrophGrowRmethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth rate of aerobic methanotrophDeadRMethanotrophsGrowth respiration parameterbdnrBulk denitrificationbr_mrBase rate of maintenance respirationFroot_leafNew fine root C per new leaf CK_domDecomposition rate constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter	KACE	Half-saturation coefficient of available carbon mineralization				
YAceMethanogensGrowth efficiency of acetoclastic methanogensYH2MethanogensGrowth efficiency of H2-CO2-dependent methanogensYMethanotrophyGrowth efficiency of aerobic methanotroph Growth rate of acetoclastic methanogensGrowRAceMethanogensGrowth rate of acetoclastic methanogensDeadRAceMethanogensDeath rate of acetoclastic methanogensGrowRH2MethanogensDeath rate of H2-CO2-dependent methanogensDeadRH2MethanogensDeath rate of aerobic methanotroph methanogensDeadRH2MethanogensDeath rate of aerobic methanotroph Death rate of aerobic methanotrophDeadRMethanotrophs DeadRMethanotrophsGrowth rate of aerobic methanotroph Death rate of aerobic methanotrophBrrFraction of leaf N in the Rubisco enzyme grpercBulk denitrification br_mrBase rate of maintenance respiration Froot_leafNew fine root C per new leaf C K_domDecomposition rate constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter	AceProdACmax	Maximum rate of acetate production from available carbon				
YH2MethanogensGrowth efficiency of H2-CO2-dependent methanogensYMethanotrophy GrowRAceMethanogensGrowth efficiency of aerobic methanotroph Growth rate of acetoclastic methanogensDeadRAceMethanogens GrowRH2MethanogensGrowth rate of acetoclastic methanogens 	YAceMethanogens	Growth efficiency of acetoclastic methanogens				
YMethanotrophy GrowRAceMethanogensGrowth efficiency of aerobic methanotroph GrowRAceMethanogensDeadRAceMethanogens DeadRAceMethanogensGrowth rate of acetoclastic methanogens Death rate of acetoclastic methanogensDeadRAceMethanogens 	YH2Methanogens	Growth efficiency of H ₂ -CO ₂ -dependent methanogens				
GrowRAceMethanogensGrowth rate of acetoclastic methanogensDeadRAceMethanogensDeath rate of acetoclastic methanogensGrowRH2MethanogensDeath rate of H2-CO2-dependent methanogensDeadRH2MethanogensDeath rate of H2-CO2-dependent methanogensGrowRmethanotrophsGrowth rate of aerobic methanotroph Death rate of aerobic methanotrophGrowRmethanotrophsGrowth rate of aerobic methanotroph Peath rate of aerobic methanotrophBrarBulk denitrification Base rate of maintenance respiration Froot_leafK_domDecomposition rate constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter	YMethanotrophy	Growth efficiency of aerobic methanotroph				
DeadRAceMethanogens Death rate of acetoclastic methanogens GrowRH2Methanogens Growth rate of H2-CO2-dependent DeadRH2Methanogens Death rate of H2-CO2-dependent methanogens Death rate of H2-CO2-dependent BeadRMethanotrophs Growth rate of aerobic methanotroph DeadRMethanotrophs Growth rate of aerobic methanotroph DeadRMethanotrophs Death rate of aerobic methanotroph Beadr Fraction of leaf N in the Rubisco enzyme grperc Growth respiration parameter bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	GrowRAceMethanogens	Growth rate of acetoclastic methanogens				
GrowRH2Methanogens Growth rate of H2-CO2-dependent methanogens DeadRH2Methanogens Death rate of H2-CO2-dependent methanogens GrowRmethanotrophs Growth rate of aerobic methanotroph Death rate of aerobic methanotroph Brand Fraction of leaf N in the Rubisco enzyme grperc Growth respiration parameter Bulk denitrification bdnr Busk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	DeadRAceMethanogens	Death rate of acetoclastic methanogens				
DeadRH2Methanogens Death rate of H2-CO2-dependent methanogens GrowRmethanotrophs Growth rate of aerobic methanotroph DeadRMethanotrophs Death rate of aerobic methanotroph DeadRMethanotrophs Death rate of aerobic methanotroph flnr Fraction of leaf N in the Rubisco enzyme grperc Growth respiration parameter bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	GrowRH2Methanogens	Growth rate of H ₂ -CO ₂ -dependent methanogens				
GrowRmethanotrophs DeadRMethanotrophsGrowth rate of aerobic methanotroph Death rate of aerobic methanotrophflnrFraction of leaf N in the Rubisco enzyme grpercgrpercGrowth respiration parameterbdnrBulk denitrificationbr_mrBase rate of maintenance respiration Froot_leafK_domDecomposition rate constant dissolved organic matterDom_diffusDiffusion of dissolved organic matter	DeadRH2Methanogens	Death rate of H ₂ -CO ₂ -dependent methanogens				
DeadRMethanotrophs Death rate of aerobic methanotroph flnr Fraction of leaf N in the Rubisco enzyme grperc Growth respiration parameter bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	GrowRmethanotrophs	Growth rate of aerobic methanotroph				
flnr Fraction of leaf N in the Rubisco enzyme grperc Growth respiration parameter bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	DeadRMethanotrophs	Death rate of aerobic methanotroph				
grperc Growth respiration parameter bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	flnr	Fraction of leaf N in the Rubisco enzyme				
bdnr Bulk denitrification br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	grperc	Growth respiration parameter				
br_mr Base rate of maintenance respiration Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	bdnr	Bulk denitrification				
Froot_leaf New fine root C per new leaf C K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	br_mr	Base rate of maintenance respiration				
K_dom Decomposition rate constant dissolved organic matter Dom_diffus Diffusion of dissolved organic matter	Froot_leaf	New fine root C per new leaf C				
Dom_diffus Diffusion of dissolved organic matter	K_dom	Decomposition rate constant dissolved organic matter				
	Dom_diffus	Diffusion of dissolved organic matter				

C. Then, a final spin-up was set up for 50 years to allow the ecosystem to transient to a state with the realistic decomposition rates before the transient simulations that cover the period of 1850–2018 (Koven et al. 2013; Thornton et al., 2005).

The model parameterization was initialized with the default parameters in Xiaofeng et al. (2015) and Yihui et al. (2019); it was performed within their ranges to determine the optimal values of parameters in the microbial module for simulating the observational GPP and CH₄ flux for each site. For the marshland in the Sanjiang Plain, the observed data of GPP and CH₄ flux during 2012–2013 were used for model validation. For the peatland at the Changbai Mountain, we extracted the GPP data during 2011 (Cao 2015.) and CH₄ flux data during 2016 (Shi 2019) from the publications. For Lesser Khingan Mountain, we extracted the GPP data during 2010–2011 from MODIS and CH₄ flux data during 2013–2014 (Huang 2016).

Model evaluation

Simple linear regression was conducted to evaluate the model performance in terms of GPP and CH_4 flux. The error statistics were used to quantify the difference between the modeled results and observational data. In order to verify the accuracy of the model, we used three accuracy evaluation indicators, coefficient of determination (R^2), root mean square error (RMSE), and mean absolute error (MAE), to evaluate model efficacy.

$$R^{2} = \frac{\sum_{i=1}^{n} (y_{i} - \widehat{y}_{i})^{2}}{\sum_{i=1}^{n} (y_{i} - \overline{y})^{2}}$$
(1)

$$RMSE = \sqrt{\frac{\sum_{i=1}^{n} (y_i - \widehat{y_i})^2}{n}}$$
(2)

$$MAE = \frac{\sum_{i=1}^{n} |y_i - \hat{y}_i|}{n}$$
(3)

Where y_i is the observed value; $\hat{y_i}$ means the simulated value; n is the number of data points. The MAE indicates the mean error of the model simulation, and thus lower MAE values suggest better model performance. The RMSE quantifies the mean error of model simulation with low values indicating high model accuracy. Higher R² values indicate better performance of the model, while lower R² values mean worse model performance and a smaller proportion of variation is explained by the model. It is noteworthy that R² is not suitable for assessing the goodness-of-fit for the dataset with small sample size.

Sensitivity analysis

To identify the most important process and the most sensitive parameters for CH_4 and GPP dynamics, a global sensitivity analysis was conducted for each wetland type. It focused on the 18 parameters related to

plant and microbial processes that are critical for microbial biogeochemistry (Table 2). For each parameter, we set up model simulations with +20% and -20% changes and investigated the responses of the modeled GPP and CH_4 flux. The index *S*, comparing the change in the model output relative to the model response for a nominal set of parameters, was calculated based on the flowing equation (Xiaofeng et al. 2015):

$$S = \frac{(Ra - Rn)/Rn}{(Pa - Pn)/Pn}$$
(4)

where S is the ratio of the standardized change in model response to the standardized change in parameter values. Ra and Rn are model responses for altered and nominal parameters, respectively, and Pa and Pn are the altered and nominal parameters, respectively. S is negative if the direction of model response opposes the direction of parameter change (Xiaofeng et al. 2015; Yihui et al. 2019; Yuan et al., 2021a 2021; Yuan et al., 2021b).

Results

Comparison of simulated GPP and CH₄ flux against observational data

The CLM-Microbe model was able to reconstruct the GPP and CH₄ flux during the study period for all three sites (Table 3; Figure 2). For example, the simulated temporal variations in GPP and CH₄ flux were consistent with observational data ($R^2 \ge 0.41$ for all sites). There were slight differences in simulated and observed variables among sites; specifically, modeled GPP was more consistent with the observed data for the Lesser Khingan Mountain ($R^2 = 0.93$, P < 0.001; Figure 2c) than for the Sanjiang Plain ($R^2 = 0.49$, P < 0.001; Figure 2a) and Changbai Mountain ($R^2 = 0.45$, P < 0.001; Figure 2b). For the dynamics of CH₄ flux, modeled CH₄ flux was more consistent with the observational fluxes for the Changbai Mountain ($R^2 = 0.91$, P = 0.012; Figure 2e) than for the Sanjiang Plain ($R^2 = 0.55$, P < 0.001; Figure 2d) and Lesser Khingan Mountain ($R^2 = 0.41$, P = 0.01; Figure 2f). Furthermore, the molded DOC, the immediate substrate of CH₄ production, was consistent with the observational data (Table 3).

The MAE and RMSE values suggested large variations in model performance when simulating GPP and CH_4 among sites. The MAE and RMSE varied within a range of 28- and 27- fold, respectively, among sites. Both MAE and RMSE values for CH_4 showed 30-fold variations among sites.

*Variability of CH*₄ *flux and GPP dynamic among three sites*

Modeled GPP exhibited large variabilities among sites (Figure 3). The marshland wetland in the Sanjiang Plain has an annual modeled GPP of 589.1 g $C/m^2/$

Table 3. Site level evaluation of the goodness-of-fit criteria computed for the simulated GPP ($gC/m^2/d$), CH_4 flux ($gC/m^2/d$) and DOC.

	GPP			CH ₄ flux			DOC (mg/cm ³)	
Site	R ²	RMSE	MAE	R ²	RMSE	MAE	Modeled	Observed
Sanjiang Plain	0.49	1.13	0.86	0.55	41.1	33.67	0.28	0.24
Changbai Mountain	0.45	0.04	0.03	0.91	0.94	0.76	1.11	2.27
Lesser Khingan Mountain	0.93	0.14	0.08	0.41	1.38	1.17	0.26	1.44

MAE, mean absolute error; RMSE, root mean square error; R^2 , R square. MAE and RMSE values indicate the mean error of the model, smaller values represent higher model performance. R^2 values mean the proportion of variation is explained by the mode; higher R^2 values indicate better model performance. R^2 is not suitable for assessing the goodness-of-fit for a small amount of data due to the large bias in small samples.

year; the annual GPP was simulated to be 1319.0 g C/m²/year for the wetlands in the Changbai Mountain, and modeled 171.3 g C/m²/year for the swamp in the Lesser Khingan Mountain. For all three sites, the GPP reached its highest value in the summers of the year. The daily modeled maximum values of GPP in the Sanjiang Plain, Changbai Mountain, and Lesser Khingan Mountain were 5.8 g C/m²/d, 12.5 g C/m²/d, and 1.9 g C/m²/d, respectively. Compared with the Sanjiang Plain, the simulated GPP of Changbai Mountain and Lesser Khingan Mountain and Lesser Khingan Mountain has a much higher fluctuation.

Meanwhile, there were large variations in modeled CH_4 flux among sites, with a range of 0.34 to 26.9 g C/m²/year. The annual modeled CH_4 emissions in the Sanjiang Plain were the largest at 26.9 g C/m²/year, followed by the Lesser Khingan Mountain at 0.66 g C/m²/year, and the Changbai Mountain has the smallest modeled CH_4 emissions at only 0.34 g C/m²/year. The Sanjiang Plain freshwater marsh wetlands, as the largest source of CH_4 emissions, have released 79 times more annual CH_4 flux than the Mountain peatland (Lesser Khingan Mountain). The peak modeled CH_4 flux occurred in the summer period, and the



Figure 1. Site location map of the study area; purples diamonds are the sites for model simulations with the background as wetland distribution in northeastern China.



Figure 2. Scatter plots of the observed and simulated GPP (a-c) and CH_4 (d-f) for the Sanjiang Plain, Changbai Mountain, and Lesser Khingan Mountain. The Orange dots represent GPP, and the green dots represent CH_4 ; solid lines are the regression, dash lines are the 1:1 line.

maximum modeled daily emissions of Sanjiang Plain, Changbai Mountain, and Lesser Khingan Mountain were 247.9 mg C/m²/year, 4.2 mg C/m²/year, and 5.9 mg C/m²/year, respectively.

Modeled biomass of methanogens

The modeled microbial biomass for methanogenesis showed obvious seasonality at different depths at three sites (Figure 4). There was almost no fluctuation of microbial biomass in spring and winter. The microbial biomass of Lesser Khingan Mountain began to increase suddenly from mid-July and then dropped to stabilize in early October. But there was almost no seasonal change in acetoclastic methanogens biomass below 40 cm (Figure 4c). Different from Lesser Khingan Mountain, the seasonal changes of methanogenesis in the Sanjiang Plain and the Changbai Mountain were relatively similar, strongly fluctuating between April and October (Figure 4 a-4b, Figure 4 d-4e).

There are significant differences in biomass of methanogens at the three sites (Figure 4). The simulation results showed that Changbai Mountain has the largest acetoclastic methanogens biomass, with



Figure 3. Observed and the modeled (a-c) GPP and (d-f) CH_4 . (a), Sanjiang Plain (b), Changbai Mountain (c), Lesser Khingan Mountain. DOY: day of the year.

a mean of $1.37e^{-4}$ mol/m³, followed by Lesser Khingan Mountain and Sanjiang Plain, with a mean of $2.32e^{-6}$ and $1.57e^{-9}$ mol/m³, respectively. The Sanjiang Plain had the largest hydrogenotrophic methanogens biomass, followed by Changbai Mountain and Lesser Khingan Mountain, with a mean value of $2.55e^{-6}$, $1.85e^{-6}$, and $7.47e^{-7}$ mol/m³, respectively.

Modeled CH₄ transport pathway among three sites

Modeled CH_4 flux showed that wetland types had a profound impact on CH_4 transport pathways (Figure 5). The modeled CH_4 flux showed obvious seasonality in plant-mediated transport, diffusion, and ebullition at Sanjiang Plain and Changbai Mountain, while the plant-mediated transport did not show a clear seasonal pattern in Lesser

Khingan Mountain. The model estimated that the main CH₄ transport pathway in the wetland was ebullition at an annual scale. The contribution of ebullition transmission in the freshwater wetland (Sanjiang Plain) to CH₄ emissions is lower than that of mountain peatlands (Changbai Mountain and Lesser Khingan Mountain), which are 58.96% in Sanjiang Plain, 82.41% in Changbai Mountain, and 71.06% in Lesser Khingan Mountain. Plantmediated transport in freshwater wetland, as the second-largest transport pathway after ebullition, is significantly different from mountain peatlands. Among them, the contribution rate of plantmediated transport in Lesser Khingan Mountain was only 0.003%, which was almost negligible (Figure 5c). Diffusion, as the second-largest transport pathway of alpine peatlands, has the smallest



Figure 4. Temporal trend of microbial functional group (acetoclastic methanogens (a, c, e), and hydrogenotrophic methanogens (b, d, f)) across three sites. (a, b) Sanjiang Plain; (c, d) Changbai Mountain; (e, f) Lesser Khingan Mountain.



Figure 5. The CLM-Microbe model simulated CH₄ transport pathways and their contributions to the annual CH₄ flux at three sites. (a) Sanjiang Plain; (b) Changbai Mountain; (c) Lesser Khingan Mountain; DOY = day of the year.

contribution to CH_4 emissions in freshwater wetlands indeed. In addition, the simulation results can clearly foresee the early spring CH_4 pulse when the frozen soil melts in the spring (Figure 5).

Sensitivity analysis

The dynamics of CH_4 and GPP are sensitive to some parameters of photosynthesis, plant growth respiration, maintenance respiration, decomposition, CH_4

production, growth and death of methanogens, and growth and death of methanotrophs (Figure 6). The simulated CH₄ fluxes were sensitive to most of the 18 key parameters, while GPP was sensitive to the parameters controlling respiration and photosynthesis. Specifically, the CH₄ flux was strongly sensitive to the parameters of GrowRmethanotrophs, AceProdAcemax, KAce, DeadACEMethanogens, GrowACEMethanogens for Sanjiang Plain and Changbai Mountain, which means that acetate production and methanotrophs were the key controls on CH₄ flux. The changes of GrowRmethanotrophs are significantly different in the CH₄ flux response between the two wetland types. In freshwater wetland (S = 0.02 for CH₄ flux with -20% change and S = 0.02 for CH₄ flux with +20% change in Sanjiang Plain), whether the GrowRmethanotrophs increase or decreases, the CH₄ flux shows an upward trend, but the opposite is true in mountain peatlands (S = -1.14 for CH₄ flux with -20% change and S = -2.34 for CH₄ flux with +20% change in Changbai Mountain). The magnitude of the AceProdAcemax on CH₄ flux were the same between Sanjiang Plain (S = 0.18 for CH₄ flux with -20% change and S = 0.08 for CH₄ flux with +20% change) and Changbai Mountain (S = 0.63 for CH_4 flux with -20% change and S = 0.50 for CH₄ flux with +20% change). But it showed a positive correlation in Lesser Khingan Mountain (S = -0.03 for CH₄ flux with -20% change and S = 0.04 for CH₄ flux with +20% change).

In addition, photosynthesis and DOC also affect CH₄ production. In the freshwater wetland, CH₄ flux was sensitive to the fraction of leaf nitrogen in the Rubisco enzyme functioning in photosynthesis (flnr). The high response was in freshwater marshes (S = 1.44 for CH_4 flux with -20% change and S = -1.11 for CH₄ flux with +20% change in Sanjiang Plain), followed by peatlands (S = 0.14 for CH₄ flux with -20% change and S = 0.07 for CH₄ flux with +20% change in Changbai Mountain). CH₄ flux also responded to changes in autotrophic respiration (grperc and br_mr; Figure 6a). In the mountain peatlands (Changbai Mountain and Lesser Khingan Mountain), the parameters of the acetic acid methanogenesis pathway are still the main factors affecting CH₄ production. But its sensitivity is weaker than that of freshwater marshes (Figures 6 b and c).

The most important processes of GPP dynamics are related to photosynthesis and respiration, which control C fixation and release from wetlands. The flnr and grperc were identified as the primary factor for GPP, NPP, ER, and NEE (Figure 6). As increases in the flnr lead to decrease in GPP, NEE, ER and NPP in the Sanjiang Plain (S = -1.71 for GPP, S = -2.49 for NEE, S = -1.61 for ER, and S = -1.47 for NPP with +20% change) but GPP, NEE, ER and NPP raise in the Changbai Mountain (S = 0.32 for GPP, S = 0.37 for NEE, S = 0.32 for ER, and S = 0.35 for NPP with +20%change) and Lesser Khingan Mountain (S = 1.00 for GPP, S = 0.58 for NEE, S = 0.93 for ER and, S = 0.47 for NPP with +20% change). There was a significant increase in GPP, NEE, ER and NPP for all sites with decreased flnr. As decrease in the grperc lead to decrease in GPP, ER and NPP in the Sanjiang Plain and Lesser Khingan Mountain but a NEE rise in the Changbai Mountain. In addition, GPP, NEE, ER and

NPP were sensitive to maintenance respiration (br_mr) and *froot_leaf* in the Sanjiang Plain (Figure 6a).

Discussions

Simulation of CH₄ emissions from wetlands in Northeast China

The CLM-Microbe model produced consistent results with the observed CH₄ flux and GPP in wetland ecosystems (Figure 1–2). However, we also observed poor CH₄ flux simulation results at some sites, such as CH₄ flux at Changbai Mountain and Lesser Khingan Mountain (Figure 4, Table 3). There may be several reasons for the relatively poor performance of the CLM-Microbe model at those sites. First, the available observational data were not too large, resulting in small R² values at those sites. The R² is not suitable for assessing the goodness-of-fit for a small amount of data due to the large bias in small samples (He et al., 2021a). Compared with the Sanjiang Plain, we observed smaller MAE and RMSE values in Changbai Mountain and Lesser Khingan Mountain. Second, the difference in measurements methods for CH₄ flux is also an important factor. In the method section, we mentioned that the eddy covariance technique is used to measure CH₄ flux in Sanjiang Plain, while the static chamber approach is used in Changbai Mountain and Lesser Khingan Mountain. As we know, the static chamber approach has its inherent shortcomings. It changes the temperature and pressure in the box, destroys the soil environment, and affects CH₄ emissions. In addition, this method has large spatial variability and large single-point observation errors, requiring a lot of repetition, and it is difficult to achieve long-term continuous observation (Wang et al. 2013). The biggest advantage of the eddy covariance technique is to achieve long-term continuous observation of large-scale ecosystems without damaging the environment (Wang et al. 2013). The inter-annual differences in CH₄ flux measured by the static chamber approach are obvious. Third, although the microbial processes of CH₄ production and oxidation are included in our model, the lack of microbial data makes it difficult to verify the microbial biomass changes of methanogens. This might be one of the reasons for relatively low model performances. Yihui et al. (2019) also pointed out the impact of topography on simulations should be considered when using the CLM-Microbe model.

Wetland types impacts on CH₄ flux

Our results show that there are significant differences in GPP and CH_4 flux between freshwater wetlands and peatlands, which is mainly due to differences in vegetation composition (Figure 3). The dominant species in the



Figure 6. Sensitivity analysis of the CLM-Microbe model in terms of gross primary productivity (GPP), net ecosystem carbon exchange (NEE), ecosystem respiration (ER), net primary productivity (NPP), and methane (CH₄) flux to 18 parameters (*KAce, AceProdACmax, GrowAceMethanogens, GrowH2Methanogens, GrowRMethanotrophs, DeadAceMethanogens, DeadH2Methanogens, YH2Methanogens, YMethanotrophs, k_dom, dom_diffus, froot_leaf, flnr, grperc, bdnr, and br_mr) for (a) Sanjiang Plain, (b)Changbai Mountain, and (c) Lesser Khingan Mountain. The symbols "+"and "-" indicate a 20% increase or 20% decrease of parameter values. Darker red and darker blue indicate a stronger positive or negative model response to a parameter change. S is negative if the direction of model response opposes the direction of a parameter change, and vice versa.*

Sanjiang Plain are aerenchyma sedge plants, while the Changbai Mountain is dominated by Carex sphagnum and shrubs, and the Lesser Khingan Mountain is dominated by sphagnum moss. There are obvious differences in plant species and productivity among wetland types, and this difference leads to differences in CH₄ flux (Ding et al. 2002). Through the simulation results of the model, we found that there is a large difference in plantmediated transport between the three sites. The plantmediated transport in the Sanjiang Plain accounted for 23.96%, while in Lesser Khingan Mountain only accounts for 0.003% of CH₄ emissions (Figure 5). Plant species are important factors influencing the ability of plants to transport CH₄ (Sun et al. 2012; Bhullar et al., 2014; Han et al. 2017; Lawrence et al. 2017). Previous studies have pointed out that the plant-mediated transport pathway of the C. lasiocarpa wetland only accounts for about 30% of CH₄ emissions, which is similar to our results (Sun et al. 2012; Ding, Cai, and Wang 2004). In addition, not only due to the high capacity for freshwater marsh plants to transport CH₄ into the

atmosphere but due to more plant litter inundated in the standing water, which could provide substrate for CH_4 production (Ding et al. 2002).

Through sensitivity analysis, we found that the rate of microbial growth and death is a key constraint on CH₄ dynamics in wetlands (Xiaofeng et al. 2015; Hernández et al. 2017). In the summer months, higher microbial biomass was associated with a higher CH₄ flux in the wetland (Figure 3 and Figure 4). The results implied that a positive correlation existed between the microbial biomass and CH₄ flux. Additionally, the microbial composition also has a significant impact on the methanogenic pathway in wetlands (Xiaofeng et al. 2015; Xueyang et al. 2020). The acetotrophic and hydrogenotrophic pathways have been reported to be the main methanogenic pathway in most environments (Angle et al. 2017; Thauer 1998; Zhang et al. 2019). Freshwater marshes are dominated by hydrogenotrophic methanogens, while alpine peatlands are dominated by acetoclastic methanogens (Figure 4), which can indicate that the Changbai Mountain and Lesser Khingan Mountain are mainly produced by aceticlastically (by

disproportionation of acetate to CO_2 and CH_4), and the Sanjiang Plain is dominated by hydrogenotrophic methanogenesis (CO_2 + H_2) (Hernández et al. 2017).

Model implications

This study has dual implications for model development and understanding CH₄ mechanisms in natural wetlands of northeastern China. The successful application of the CLM-Microbe model in simulating CH₄ flux in regions beyond of its original domain indicates the feasibility of the model structure. Meanwhile, the simulated dynamics of methanogenesis are consistent with the CH₄ production, suggesting the microbial dominance on CH₄ production. Many previous studies reported the predominance of substrates or water table on CH₄ flux (Whiting and Chanton 1993; Lipson et al. 2012; Zona et al. 2009); the causes might be the microbial responses to water table and ignorance of microbial mechanisms in those studies. Further genomic analysis in association with CH₄ processes should provide critical information for model development and validation (Xiaofeng et al. 2015).

The implication of CH₄ budget is primarily for the wetlands in Northeast China. Northeast China features the largest freshwater wetlands in China, and the natural wetlands can be found in mountains and plains. The large C storage in these ecosystems will likely be decomposed by microorganisms and released in the form of CH₄ when the climate continues warming (Song et al. 2012; Wang et al. 2017). In this study, the CLM-Microbe model performed well in capturing the variabilities in GPP and CH₄ flux for primary wetland types in northeastern China, which emphasizes the importance of spatial heterogeneity in simulating CH₄ flux in wetland ecosystem CH₄ models. The simulated differences in GPP and CH₄ emissions between freshwater wetlands and peatlands are consistent with previous studies (Sun et al. 2018; Xueyang et al. 2020). Furthermore, the sensitivity analysis showed that the photosynthetic (flnr) and respiratory (br_mr) parameters of plants are highly sensitive at all sites. Therefore, plant composition is one of the main reasons for the difference in CH₄ emissions. In addition, the peak CH₄ emission varies among different wetland types, especially in Sanjiang Plain and Lesser Khingan Mountain. The reason may be that Lesser Khingan Mountain is located in the permafrost region, and the methanogens reach their maximum activity at the end of the growing season, thereby increasing the CH₄ flux (Sun et al. 2018). Given the changing environment, the differences in CH₄ flux among wetlands call for accurate estimations of regional CH₄ flux at the regional scale (Song et al. 2019).

The prospect

With a modeling study, we found that environmental factors affect CH₄ emission by influencing microbial processes in the natural wetlands. The important role of microorganisms in the wetland ecosystem was demonstrated (Given and Dickinson 1975; He et al., 2020a; Villa 2020; Wallenius et al. 2021; Xiaofeng et al. 2015; Xu et al., 2016; Zhang et al. 2020a). Although the results of this paper prove that the CLM-Microbe model can simulate the CH₄ flux in northern wetlands, follow-up works are still needed. First, we have recognized the importance of microorganisms in the production and oxidation of CH₄ in wetlands; genomic data in association with the microbial function should be thus used for model parameterization. This allows the model to more accurately capture the dynamics in CH₄ production and oxidation. Second, with a warming climate, the area of permafrost has shrunk, and the active layer has deepened, which has caused a substantial impact on the C budget in wetlands. However, the scarcity of CH₄ flux data caused difficulties in model validation and application. We call for the monitoring of specific CH₄ processes and the establishment of an open data platform, such as ChinaFLUX and FLUXNET, to further develop the model.

Conclusions

We parameterized the CLM-Microbe model against the field observational data of CO₂ and CH₄ flux in three major wetland types in northeastern China and then applied the model to understand processes and environmental factors affecting GPP and CH₄ flux at different wetland types in northeastern China. The results showed that the CLM-Microbe model was able to reconstruct the observed GPP dynamics and CH₄ flux. The modeled results showed that freshwater marsh wetlands (Sanjiang Plain) have higher CH₄ emissions than mountain peatlands (Changbai Mountain and Lesser Khingan Mountain). However, Changbai Mountain has the largest GPP, followed by Sanjiang Plain, and Lesser Khingan Mountain has the smallest. The model estimated that the main CH₄ flux transport pathway in the wetland was ebullition at an annual time scale. The modeled results showed that the freshwater marsh wetlands mainly use the hydrogenotrophic CH₄ production, and the mountain peatlands mainly use the process of acetolactic CH₄ production. Model sensitivity analysis determined that the growth and death rate of microorganisms and the availability of substrates are the most important factors in controlling CH₄ emissions from wetland ecosystems. Photosynthesis and respiration have a more significant impact on GPP, NEE, ER, and NPP. Our finding highlights the importance of explicit representation of microbial mechanisms on C cycling in northeastern

China, which will improve the simulation performance of CH_4 cycling under climate change. Our results also provide an important scientific basis for the quantification and prediction of the CH_4 budget in wetland areas in northeastern China.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This study was partially supported by the Strategic Priority Research Program of the Chinese Academy of Sciences (Grant No. XDA28020502), the National Natural Science Foundation (No. 41771102, 41730643, 32171873, 41701198) of China, and Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences. F.H., Y.W., and X.X. are grateful for the financial and facility support from the U.S. Department of Energy and National Science Foundation (2145130) that has partially funded the development of the CLM-Microbe model.

References

- Angle, J. C., T. H. Morin, L. M. Solden, A. B. Narrowe, G. J. Smith, M. A. Borton, C. Rey-Sanchez, et al. 2017. "Methanogenesis in Oxygenated Soils Is a Substantial Fraction of Wetland Methane Emissions." *Nature Communications* 8 (1): 1567. doi:10.1038/s41467-017-01753-4.
- Bhullar, G. S., P. J. Edwards, H. Olde Venterink, and F.-H. Yu. 2014. "Influence of Different Plant Species on Methane Emissions from Soil in a Restored Swiss Wetland." *Plos One* 9 (2): e89588. doi:10.1371/journal.pone.0089588.
- Bousquet, P., P. Ciais, J. B. Miller, E. J. Dlugokencky, D. A. Hauglustaine, C. Prigent, G. R. Van der Werf, et al. 2006. "Contribution of Anthropogenic and Natural Sources to Atmospheric Methane Variability." *Nature* 443 (7110): 439–443. doi:10.1038/nature05132.
- Bridgham, S. D., H. Cadillo-Quiroz, J. K. Keller, and Q. Zhuang.
 2013. "Methane Emissions from Wetlands: Biogeochemical, Microbial, and Modeling Perspectives from Local to Global Scales." *Global Change Biology* 19 (5): 1325–1346. doi:10.1111/gcb.12131.
- Cao, N. 2015. Study on the carbon dioxide flux in Peatland of volcanic cluster in the longgang region at changbai mountain —Take the Jinchuan peatland as an example . Master. Northeast Normal University.
- Chadburn, S. E., T. Aalto, M. Aurela, D. Baldocchi, C. Biasi, J. Boike, E. J. Burke, et al. 2020. "Modeled Microbial Dynamics Explain the Apparent Temperature Sensitivity of Wetland Methane Emissions." *Global Biogeochemical Cycles* 34 (11): e2020GB006678. doi:10.1029/2020GB006678.
- Cordruwisch, R., H. J. Seitz, and R. Conrad. 1988. "The Capacity of Hydrogenotrophic Anaerobic-Bacteria to Compete for Traces of Hydrogen Depends on the Redox Potential of the Terminal Electron-Acceptor." Archives of Microbiology 149 (4): 350–357. doi:10.1007/Bf00411655.
- Ding, W., Z. Cai, H. Tsuruta, and L. Xiaoping. 2002. "Effect of Standing Water Depth on Methane Emissions from Freshwater Marshes in Northeast China." Atmospheric Environment 36 (33): 5149–5157. doi:10.1016/S1352-2310(02)00647-7.

- Ding, W., Z. Cai, and D. Wang. 2004. "Preliminary Budget of Methane Emissions from Natural Wetlands in China." *Atmospheric Environment* 38 (5): 751–759. doi:10.1016/j. atmosenv.2003.10.016.
- Fazli, H. C., U. K. M. Man, and A., I. Shah. 2013. "Characteristics of Methanogens and Methanotrophs in Rice Fields: A Review." *Asia Pac J Mol Biol Biotechnol* 21 (1): 3–17.
- Given, P. H., and C. H. Dickinson. 1975. "Biochemistry and Microbiology of Peats." *Soil Biochemistry*, 3:123–212.
- Han, W., M. Shi, J. Chang, Y. Ren, R. Xu, C. Zhang, and Y. Ge. 2017. "Plant Species Diversity Reduces N₂O but Not CH₄ Emissions from Constructed Wetlands under High Nitrogen Levels." *Environmental Science and Pollution Research* 24 (6): 5938–5948. doi:10.1007/s11356-016-8288-3.
- Han, Y., M. Wang, S. Wang, Y. Dong, S. Liu, and X. Zhiwei. 2018. "Characteristics of Soil Enzyme Activity of Peat Bog in Jinchuan, Changbai Mountain." *Wetland Science* 16 (5): 671–678. doi:10.13248/j.cnki.wetlandsci.2018.05.014.
- Hanqin, T., L. Chaoqun, C. Philippe, A. M. Michalak, J. G. Canadell, S. Eri, D. N. Huntzinger, et al. 2016. "The Terrestrial Biosphere as a Net Source of Greenhouse Gases to the Atmosphere." *Nature* 531 (7593): 225-+. doi:10.1038/nature16946.
- Harmsen, M., D. P. van Vuuren, B. L. Bodirsky, J. Chateau,
 O. Durand-Lasserve, L. Drouet, O. Fricko, S. Fujimori,
 D. E. H. J. Gernaat, and T. Hanaoka. 2020. "The Role of Methane in Future Climate Strategies: Mitigation Potentials and Climate Impacts." *Climatic Change* 163 (3): 1409–1425. doi:10.1007/s10584-019-02437-2.
- He, L., Chun-Ta. L, Melanie A. M, Shohei. M, Xiaofeng. X. 2021a. "Microbial seasonality promotes soil respiratory carbon emission innatural ecosystems: A modeling study". *Global Change Biology*, 27(13): 3035-3051. https:// doi.org/10.1111/gcb.15627.
- He, L., David, A. L, Jorge L. M. R, Melanie. M, Robert G. B, Bruno. G, Peter. T, Xiaofeng. X. 2021b. "Dynamics of Fungal and Bacterial Biomass Carbon in Natural Ecosystems: Site-Level Applications of the CLM-Microbe Model". *Journal of Advances in Modeling Earth Systems* 13: (2) e2020MS002283. https://doi. org/10.1029/2020MS002283.
- Hernández, M., R. Conrad, M. Klose, K. Ma, and L. Yahai. 2017. "Structure and Function of Methanogenic Microbial Communities in Soils from Flooded Rice and Upland Soybean Fields from Sanjiang Plain, NE China." *Soil Biology* and *Biochemistry* 105: 81–91. doi:10.1016/j. soilbio.2016.11.010.
- Huang, Y., R. L. Sass, and F. M. Fisher Jr. 1998. "A semi-empirical Model of Methane Emission from Flooded Rice Paddy Soils." *Global Change Biology* 4 (3): 247–268. doi:10.1046/j.1365-2486.1998.00129.x.
- Huang, Shizhu 2016 Response of CH_4 and N_2O emission from peatlands to changed environments in Little Xing'an Mountains, northeat China . . Doctor. Northeast Forestry University.
- IPCC, 2017. "IPCC Fifth Assessment Report (AR5) Observed Climate Change Impacts Database, Version 2.01." NASA Socioeconomic Data and Applications Center (SEDAC), Palisades, NY.
- Jackson, R. B., M. Saunois, P. Bousquet, J. G. Canadell, B. Poulter, A. R. Stavert, P. Bergamaschi, Y. Niwa, A. Segers, and A. Tsuruta. 2020. "Increasing Anthropogenic Methane Emissions Arise Equally from Agricultural and Fossil Fuel Sources." *Environmental Research Letters* 15 (7): 071002. doi:10.1088/ 1748-9326/ab9ed2.
- Kirschke, S., P. Bousquet, P. Ciais, M. Saunois, J. G. Canade II, E. J. Dlugokencky, P. Bergamaschi, et al. 2013. "Three Decades of Global Methane Sources and Sinks." *Nature Geoscience* 6 (10): 813–823. doi:10.1038/ngeo1955.

- Koven, C. D., W. J. Riley, Z. M. Subin, J. Y. Tang, M. S. Torn,
 W. D. Collins, G. B. Bonan, D. M. Lawrence, and
 S. C. Swenson. 2013. "The Effect of Vertically Resolved Soil Biogeochemistry and Alternate Soil C and N Models on C Dynamics of CLM4." *Biogeosciences* 10 (11): 7109–7131. doi:10.5194/bg-10-7109-2013.
- Lawrence, B. A., S. C. Lishawa, N. Hurst, B. T. Castillo, and N. C. Tuchman. 2017. "Wetland Invasion by Typha× Glauca Increases Soil Methane Emissions." *Aquatic Botany* 137: 80–87. doi:10.1016/j.aquabot.2016.11.012.
- Le, M., and R. P. Jean. 2001. "Production, Oxidation, Emission and Consumption of Methane by Soils: A Review." *European Journal of Soil Biology* 37 (1): 25–50. doi:10.1016/S1164-5563(01)01067-6.
- Lipson, D. A., D. Zona, T. K. Raab, F. Bozzolo, M. Mauritz, and W. C. Oechel. 2012. "Water-table Height and Microtopography Control Biogeochemical Cycling in an Arctic Coastal Tundra Ecosystem." *Biogeosciences* 9 (1): 577–591. doi:10.5194/bg-9-577-2012.
- Liyuan, H., C.-T. Lai, M. A. Mayes, S. Murayama, and X. Xu. 2021a. "Microbial Seasonality Promotes Soil Respiratory Carbon Emission in Natural Ecosystems: A Modeling Study." *Global Change Biology* 27 (13): 3035–3051. doi:10.1111/gcb.15627.
- Liyuan, H., D. A. Lipson, M. Rodrigues, L. Jorge, M. Melanie, R. G. Björk, G. Bruno, T. Peter, and X. Xiaofeng. 2021b.
 "Dynamics of Fungal and Bacterial Biomass Carbon in Natural Ecosystems: Site-Level Applications of the CLM-Microbe Model." *Journal of Advances in Modeling Earth Systems* 13 (2): e2020MS002283. doi:10.1029/2020MS002283.
- Maosheng, Z., F. A. Heinsch, R. R. Nemani, and S. W. Running. 2005. "Improvements of the MODIS Terrestrial Gross and Net Primary Production Global Data Set." *Remote Sensing of Environment* 95 (2): 164–176. doi:10.1016/j.rse.2004.12.011.
- McGlynn, S. E. 2017. "Energy Metabolism during Anaerobic Methane Oxidation in ANME Archaea." *Microbes and Environments* 32 (1): 5–13. doi:10.1264/jsme2.ME16166.
- Melton, J. R., R. Wania, E. L. Hodson, B. Poulter, B. Ringeval, R. Spahni, T. Bohn, et al. 2013. "Present State of Global Wetland Extent and Wetland Methane Modelling: Conclusions from a Model inter-comparison Project (WETCHIMP)." *Biogeosciences* 10 (2): 753–788. doi:10.5194/ bg-10-753-2013.
- Meng, L., P. G. M. Hess, N. M. Mahowald, J. B. Yavitt,
 W. J. Riley, Z. M. Subin, D. M. Lawrence,
 S. C. Swenson, J. Jauhiainen, and D. R. Fuka. 2012.
 "Sensitivity of Wetland Methane Emissions to Model Assumptions: Application and Model Testing against Site Observations." *Biogeosciences* 9 (7): 2793–2819. doi:10.5194/bg-9-2793-2012.
- Morin, T. H., G. Bohrer, R. P. D. M. Frasson, L. Naor-Azreli, S. Mesi, K. C. Stefanik, and K. V. R. Schäfer. 2014. "Environmental Drivers of Methane Fluxes from an Urban Temperate Wetland Park." *Journal of Geophysical Research: Biogeosciences* 119 (11): 2188–2208. doi:10.1002/ 2014JG002750.
- Nzotungicimpaye, C.-M., A. H. MacDougall, J. R. Melton, C. C. Treat, E. Michael, L. F. W. Lesack, and K. Zickfeld. 2020. "WETMETH 1.0: A New Wetland Methane Model for Implementation in Earth System Models." *Geoscientific Model Development Discussions* 1–40. doi:10.5194/gmd-2020-176.
- Saunois, M., A. R. Stavert, B. Poulter, P. Bousquet, J. G. Canadell, R. B. Jackson, P. A. Raymond, et al. 2020. "The Global Methane Budget 2000-2017"." *Earth System Science Data* 12 (3): 1561–1623. doi:10.5194/essd-12-1561-2020.

- Schaefer, H. 2019. "On the Causes and Consequences of Recent Trends in Atmospheric Methane." *Current Climate Change Reports* 5 (4): 259–274. doi:10.1007/s40641-019-00140-z.
- Shi, Yao 2019 Effects of nitrogen input on carbon and nitrogen transformations in peatlands . Doctor, Northeast Normal University.
- Sieber, J. R., M. J. McInerney, and R. P. Gunsalus. 2012. "Genomic Insights into Syntrophy: The Paradigm for Anaerobic Metabolic Cooperation." *Annual Review of Microbiology* 66 (1): 66(429–452. doi:10.1146/annurevmicro-090110-102844.
- Song, C., X. Xiaofeng, X. Sun, H. Tian, L. Sun, Y. Miao, X. Wang, and Y. Guo. 2012. "Large Methane Emission upon Spring Thaw from Natural Wetlands in the Northern Permafrost Region." *Environmental Research Letters* 7 (3). doi:10.1088/1748-9326/ 7/3/034009.
- Song, Y., C. Song, J. Ren, M. Xiuyan, W. Tan, X. Wang, J. Gao, and A. Hou. 2019. "Short-Term Response of the Soil Microbial Abundances and Enzyme Activities to Experimental Warming in a Boreal Peatland in Northeast China." Sustainability 11 (3): 590. doi:10.3390/su11030590.
- Sun, X., C. Song, Y. Guo, X. Wang, G. Yang, L. Yingchen, R. Mao, and L. Yongzheng. 2012. "Effect of Plants on Methane Emissions from a Temperate Marsh in Different Seasons." *Atmospheric Environment* 60 (277): 277–282. doi:10.1016/j.atmosenv.2012.06.051.
- Sun, L., S. Changchun, P. M. Lafleur, M. Yuqing, W. Xianwei, G. Chao, Q. Tianhua, Y. Xueyang, and W. Tan. 2018. "Wetland-atmosphere Methane Exchange in Northeast China: A Comparison of Permafrost Peatland and Freshwater Wetlands." Agricultural and Forest Meteorology 249 (239): 239–249. doi:10.1016/j.agrformet.2017.11.009.
- Thauer, R. K. 1998. "Biochemistry of Methanogenesis: A Tribute to Marjory Stephenson: 1998 Marjory Stephenson Prize Lecture." *Microbiology* 144 (9): 2377–2406. doi:10.1099/00221287-144-9-2377.
- Thauer, R. K., A. K. Kaster, H. Seedorf, W. Buckel, and R. Hedderich. 2008. "Methanogenic Archaea: Ecologically Relevant Differences in Energy Conservation." *Nature Reviews Microbiology* 6 (8): 579–591. doi:10.1038/nrmicro1931.
- Thornton P E and Rosenbloom N A. (2005). Ecosystem model spin-up: Estimating steady state conditions in a coupled terrestrial carbon and nitrogen cycle model. Ecological Modelling, 189(1–2), 25–48. 10.1016/j.ecolmodel.2005.04.008
- Thornton, P. E., J.-F. Lamarque, N. A. Rosenbloom, and N. M. Mahowald. 2007. "Influence of carbon-nitrogen Cycle Coupling on Land Model Response to CO2 Fertilization and Climate Variability." *Global Biogeochemical Cycles* 21 (4). doi:10.1029/2006GB002868.
- Timmers, P. H. A., C. U. Welte, J. J. Koehorst, C. M. Plugge, M. S. M. Jetten, and A. J. M. Stams. 2017. "Reverse Methanogenesis and Respiration in Methanotrophic Archaea." Archaea-an International Microbiological Journal 2017. doi:10.1155/2017/1654237.
- Tingting, L., L. Yanyu, Y. Lingfei, W. Sun, Q. Zhang, W. Zhang, G. Wang, et al. 2020. "Evaluation of CH4MOD(wetland) and Terrestrial Ecosystem Model (TEM) Used to Estimate Global CH4 Emissions from Natural Wetlands." *Geoscientific Model Development* 13 (8): 3769–3788. doi:10.5194/gmd-13-3769-2020.
- Villa, J. A. 2020. "Functional Representation of Biological Components in Methane-Cycling Processes in Wetlands Improves Modeling Predictions." *Journal of Geophysical Research: Biogeosciences* 125 (10): e2020JG005794. doi:10.1029/2020JG005794.

- Wallenius, A. J., D. Martins, S. Paula, P. Caroline, and M. S. M. Jetten. 2021. "Anthropogenic and Environmental Constraints on the Microbial Methane Cycle in Coastal Sediments." *Frontiers in Microbiology* 12 (293). doi:10.3389/fmicb.2021.631621.
- Wang, J. M., J. G. Murphy, J. A. Geddes, C. L. Winsborough, N. Basiliko, and S. C. Thomas. 2013. "Methane Fluxes Measured by Eddy Covariance and Static Chamber Techniques at a Temperate Forest in Central Ontario, Canada." *Biogeosciences* 10 (6): 4371–4382. doi:10.5194/bg-10-4371-2013.
- Wang, J., C. Song, A. Hou, and X. Fengming. 2017. "Methane Emission Potential from Freshwater Marsh Soils of Northeast China: Response to Simulated Freezing-Thawing Cycles." Wetlands 37 (3): 437–445. doi:10.1007/s13157-017-0879-3.
- Wang, Y., F. Yuan, K. A. Arndt, J. Liu, L. He, Y. Zuo, D. Zona, et al. 2022. "Upscaling Methane Flux from plot-level to Eddy Covariance Tower Domains by Combining the CLM-Microbe Model with Three Footprint Models." *Atmospheric Environment* Under review
- Wania, R., I. Ross, and I. C. Prentice. 2010. "Implementation and Evaluation of a New Methane Model within a Dynamic Global Vegetation Model: LPJ-WHyMe v1.3.1." *Geoscientific Model Development* 3 (2): 565–584. doi:10.5194/gmd-3-565-2010.
- Wania, R., J. R. Melton, E. L. Hodson, B. Poulter, B. Ringeval, R. Spahni, T. Bohn, et al. 2013. "Present State of Global Wetland Extent and Wetland Methane Modelling Methodology of a Model inter-comparison Project (WETCHIMP)." *Geosci. Model Dev* 6 (3): 617–641. doi:10.5194/gmd-6-617-2013.
- Whiting, G. J., and J. P. Chanton. 1993. "Primary Production Control of Methane Emission from Wetlands." *Nature* 364 (6440): 794–795. doi:10.1038/364794a0.
- Xiaofeng, X., and H. Tian. 2012. "Methane Exchange between Marshland and the Atmosphere over China during 1949-2008." *Global Biogeochemical Cycles* 26 (2). doi:10.1029/2010gb003946.
- Xiaofeng, X., J. P. Schimel, P. E. Thornton, S. Xia, Y. Fengming, and S. Goswami. 2014. "Substrate and Environmental Controls on Microbial Assimilation of Soil Organic Carbon: A Framework for Earth System Models." *Ecology Letters* 17 (5): 547–555. doi:10.1111/ele.12254.
- Xiaofeng, X., D. A. Elias, D. E. Graham, T. J. Phelps, S. L. Carroll, S. D. Wullschleger, and P. E. Thornton. 2015. "A Microbial Functional group-based Module for Simulating Methane Production and Consumption: Application to an Incubated Permafrost Soil." *Journal of Geophysical Research Biogeosciences* 120 (7): 1315–1333. doi:10.1002/ 2015JG002935.
- Xiaofeng, X., Y. Fengming, J. H. Paul, D. W. Stan, E. T. Peter, J. R. William, S. Xia, E. G. David, S. Changchun, and T. Hanqin. 2016. "Reviews and Syntheses: Four Decades of Modeling Methane Cycling in Terrestrial Ecosystems." *Biogeosciences*, 1–56. doi:10.5194/bg-2016-37.
- Xueyang, Y., C. Song, L. Sun, X. Wang, and W. Tan. 2020. "Towards an Improved Utilization of Eddy Covariance Data: Growing Season CO₂ Exchange from a Permafrost Peatland in the Great Hing'an Mountains, Northeast China." *Ecological Indicators* 115: 106427. doi:10.1016/j. ecolind.2020.106427.

- Yihui, W., Fengming. Y., F. Yuan., Baohua. G., Melanie S. H., MS. Torn., Daniel M. R., Jitendra. K., Liyuan. H., Donatella. Z., David A. L., Robert. W. Walter C. O., Stan D. W., Peter E. T., Xiaofeng. X. 2019. "Mechanistic Modeling of Microtopographic Impacts on CO2 and CH4 Fluxes in an Alaskan Tundra Ecosystem Using the CLM-Microbe Mode." Journal of Advances in Modeling Earth Systems 11 (12): 4288–4304 https://doi.org/ 10.1029/2019MS001771.
- Yihui, W., F. Yuan, F. Yuan, B. Gu, M. S. Hahn, M. S. Torn, D. M. Ricciuto, et al. 2019. "Mechanistic Modeling of Microtopographic Impacts on CO2 and CH4 Fluxes in an Alaskan Tundra Ecosystem Using the CLM-Microbe Model." Journal of Advances in Modeling Earth Systems 11 (12): 4288–4304. doi:10.1029/2019MS001771.
- Youmi, O., Z. Qianlai, L. Licheng, L. R. Welp, M. C. Y. Lau, T. C. Onstott, M. David, et al. 2020. "Reduced Net Methane Emissions Due to Microbial Methane Oxidation in a Warmer Arctic." *Nature Climate*

Change 10 (4): 317-321. doi:10.1038/s41558-020-0734-z.

- Yuan, F., J. Liu, Y. Zuo, Z. Guo, N. Wang, C. Song, Z. Wang, et al. 2020. "Rising Vegetation Activity Dominates Growing Water Use Efficiency in the Asian Permafrost Region from 1900 to 2100." Science of the Total Environment 736 (139587): 139587. doi:10.1016/j. scitotenv.2020.139587.
- Yuan, F., Y. Wang, D. Ricciuto, X. Shi, F. Yuan, T. Brehme,
 S. Bridgham, et al. 2021a. "Hydrological Feedbacks on Peatland CH4 Emission under Warming and Elevated CO₂ : A Modeling Study." *Journal of Hydrology* 603: 127137. doi:10.1016/j.jhydrol.2021.127137.
- Yuan, F., Y. Wang, D. Ricciuto, X. Shi, F. Yuan, P. Hanson, S. Bridgham, J. Keller, P. Thornton, and X. Xiaofeng. 2021b. "An Integrative Model for Soil Biogeochemistry and Methane Processes: II. Warming and Elevated CO₂ Effects on Peatland CH4 Emissions." Journal of Geophysical Research: Biogeosciences 126. doi:10.1029/ 2020JG005963.
- Zhang, B., H. Tian, L. Chaoqun, G. Chen, S. Pan, C. Anderson, and B. Poulter. 2017. "Methane Emissions from Global Wetlands: An Assessment of the Uncertainty Associated with Various Wetland Extent Data Sets." Atmospheric Environment 165: 310–321. doi:10.1016/j.atmosenv.2017.07.001.
- Zhang, Y., M. Anzhou, G. Zhuang, and X. Zhuang. 2019. "The Acetotrophic Pathway Dominates Methane Production in Zoige Alpine Wetland Coexisting with Hydrogenotrophic Pathway." *Scientific Reports* 9 (1): 9141. doi:10.1038/s41598-019-45590-5.
- Zhang, H., D. Goll, Y. Wang, P. Ciais, W. Wieder, R. Abramoff, Y. Huang, et al. 2020a. "Microbial Dynamics and Soil Physicochemical Properties Explain Large Scale Variations in Soil Organic Carbon." *Global Change Biology* 26 (4): 2668–2685. doi:10.1111/ gcb.14994.
- Zona, D., W. C. Oechel, J. Kochendorfer, U. Paw, K. T. Salyuk, P. C. A. N., Olivas, S. F. Oberbauer, and D. A. Lipson. 2009. "Methane Fluxes during the Initiation of a large-scale Water Table Manipulation Experiment in the Alaskan Arctic Tundra." *Global Biogeochemical Cycles* 23 (2). doi:10.1029/2009GB003487.