



In situ electrochemical synthesis of graphene-poly(arginine) composite for *p*-nitrophenol monitoring

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ABSTRACT

Para-Nitrophenol (*p*-nitrophenol) is a common industrial pollutant occurring widely in water bodies, yet actual monitoring methods are limited. Herein we proposed a fully electrochemically *in situ* synthesized graphene-polyarginine composite functionalized screen printed electrode, as a novel *p*-nitrophenol sensing platform. The electrode was characterized by morphologic, spectrometric and electrochemical techniques. *p*-nitrophenol in both pure aqueous solution and real water samples was tested. Results show a detection limit as low as the nanomolar level, and display a linear response and high selectivity in the range of 0.5–1250 μ M. Molecular simulation reveals a detailed synergy between graphene and poly-arginine. The preferable orientation of nitrophenol molecules on the graphene interface in the presence of poly-arginine induces H- and ionic binding. This sensor is an ideal prototype for *p*-nitrophenol quantification in real waters.

1. Introduction

Nitrophenols, such as *p*-nitrophenol, is vastly used in pharmaceuticals, pesticides, dyes, explosives and petroleum industries (Wang et al., 2016; Kumar et al., 2017). In air, water and soil, *p*-nitrophenol is found with high chemical stability and reluctant biodegradability, which make its removal challengeable (Han et al., 2019). It is highly toxic and can cause irreversible damages to human liver and kidneys. Therefore, *p*-nitrophenol has been classified priority pollutant among top 114 organic pollutants by the US Environmental Protection Agency, with allowed limit of 0.43 μ M (60 ppb) in drinking water (Menazea and Mostafa, 2020; Chakraborty et al., 2021).

Common analysis of *p*-nitrophenol include spectrophotometry (Zhang and Wilson, 2008; Niazi and Yazdanipour, 2007), ultra-high-pressure liquid chromatography (Chung et al., 2012),

electrophoresis (Guo et al., 2004), molecular recognition assay (Liu et al., 2019; Dai et al., 2020) and fluorescent assays (Dai et al., 2020; Bogireddy et al., 2020). Although these methods present some advantages, they are limited by the need for man-sized instruments, expenses, experienced operators and slowness. Alternatively, electrochemical methods feature outstanding performances such as high-sensitivity, fast response, low-cost, easy operation and instrumental miniaturization (Karaman et al., 2021a, 2021b; Xiao et al., 2018, 2020; Karimi-Maleh et al., 2021a, 2021b; Balasubramanian et al., 2019; Li et al., 2012). Electrochemical detectors of nitrophenols have been recently designed using various active materials as well as electro-analytical techniques, such as semiconductive metal composites (Zhang et al., 2018; Wang et al., 2019a, 2019b; Su et al., 2021), metal nanoparticles (Madhu et al., 2014; Chao et al., 2017), polymers (Zhu et al., 2020) and carbon nanoparticles (Karaman et al., 2021a, 2021b; Karimi-Maleh et al.,

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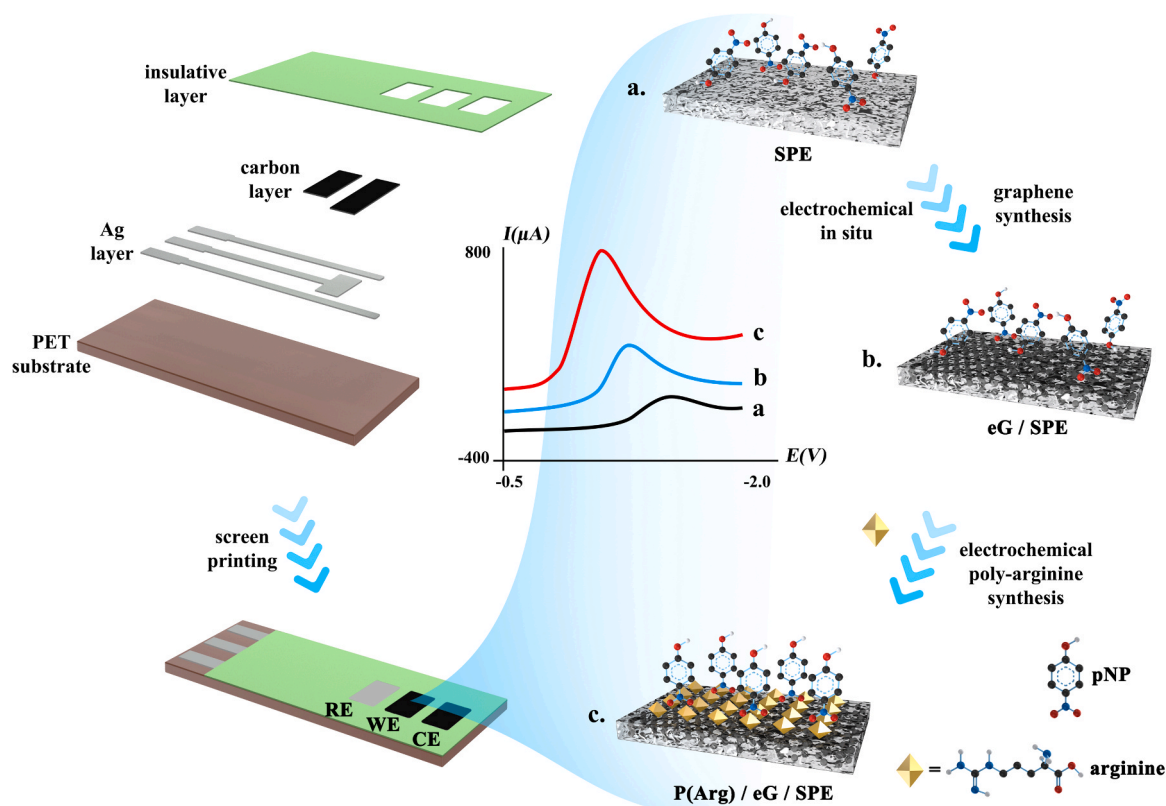
Table 1
Electrochemical biosensors for detection of *p*-nitrophenol.

Electrode architecture	Electrochemical technology	Linear range (μM)	Sensitivity ($\mu\text{A } \mu\text{M}^{-1} \text{cm}^{-2}$)	LOD (μM)	Ref.
AgNWs-PANI/GCE	DPV	0.6–32	–	0.052	Chao et al., 2017
Cu-curcumin/GCE	DPV	0.1–1030	1.57	68.2	Dinesh and Saraswathi, 2017
FeO ₂ /TiO ₂ @mc/GCE	Amperometry	5–310	3.89	0.183	Wang et al., 2019a, 2019b
AgPd@UiO-66- NH ₂ /GCE	DPV	100–370	1259.1	32	Hira et al., 2019
HA-NP/GCE	DPV	1–300	–	0.6	Yin et al., 2010
ZnCo ₂ O ₄ /GCE	DPV	1–4000	0.318	0.3	Zhang et al., 2018
PANI-array@CNF/GCE	DPV	0.01–0.1	1.09	0.0015	Zhu et al., 2020
		0.1–100	0.24		
PEDOT-PSS/ITO	LSV	0–80	0.129	4.51	Hryniewicz et al., 2018
GO/GCE	LSV	0.1–120	–	0.02	
GC/rGO-Ag	SWV	0.01–0.1	12.65	0.0012	Ikhsan et al., 2016
rGO- β -CD-CS	DPV	5–40	0.38	–	Li et al., 2017
SWCNT/GCE	DPV	0.01–5.00	–	2.5	Yang, 2004
MWCNT/GCE	SWV	1–30	–	0.12	Arvinte et al., 2011
Mn-Fe ₂ O ₄ /3D-G	DPV	5–100	0.646	0.019	Su et al., 2021
AC/GCE	DPV	1–500	–	0.16	Madhu et al., 2014
P(Arg)/eG/SPE	SWV	0.5–1250	0.661	0.012	This work
	Amperometry (Ox)	5–1250	1.295	0.0024	
	Amperometry (Red)	5–200	0.793	0.0039	

2021a, 2021b). In particular, carbon nanomaterials were successfully applied to the detection of both small compounds and bio-macromolecules (Luo et al., 2020; Shen et al., 2019; Sengupta et al., 2020). However, optimizing all specifications such as high sensitivity, low detection limit, wide linear response, easy handling and low cost is challenging, as depicted in a comparison of actual methods (Table 1).

Among carbon materials, graphene has unique properties including high electrical conductance, huge surface area and absorbability, excellent electro-catalytic activity and versatile surface chemistry, which makes graphene an ideal electrochemical sensing material for *p*-nitrophenol (Cinti and Arduini, 2017; Shari et al., 2019). Wiench et al. (2017) demonstrated that the hydrothermally reduced graphene oxide interface is an excellent *p*-nitrophenol electro-catalyst, and

extraordinary sensing stability within pH 4.0–5.0 was achieved. Nevertheless, interface of raw graphene is not optimal for detection, thus requiring architecture engineering and design of graphene composites. Mono-dispersions prevent graphene from the loss of electrochemical activity caused by graphene sheet restacking, which limits the contact of the analyte with graphene active sites (Shang et al., 2008; Chen et al., 2012; Cheng et al., 2017). As a consequence, scientists have designed composites by grafting functional materials such as metal nanoparticles, metal compounds, polymers, carbons and peptides (Yang et al., 2008; Cuenya, 2013; Su et al., 2021). Manganese cobalt oxide was introduced into graphene oxide (GO) as a dispersant to acquire better dispersed GO micro-architecture for *p*-nitrophenol analysis. Results showed that the metal oxide-GO composite exhibited much better



Scheme 1. Schematic presentation of the fabrication of the electrodes.

electro-catalytic activity than the oxide and GO alone (Manjula and Chen, 2021).

Moreover, modifying graphene morphology by preparing graphene sponges and graphene-carbon hybrids can improve electro-catalytic properties (Asadian et al., 2017; Mohamed et al., 2018; Zhu et al., 2018). Su et al. (2021) fabricated a 3D graphene framework by using urushiol as an inter-layer spacer, and the pores within the framework were adapted to accommodate metal oxide catalyst. Sensor prepared thereof presented a limit of detection (LOD) to *p*-nitrophenol as low as 19 nM. In one study (Jaiswal and Tiwari, 2021), graphene framework was prepared by zinc ion chelating with a GO suspension. The coordination product formed 3D multi-layered structure to fully expose electro-catalytically active sites. The framework was highly sensitive to *p*-nitrophenol with LOD about 96 nM. Moreover, it can be used for distinguishing *p*-nitrophenol from *o*-nitrophenol. But those systems are costly and poorly stable in real conditions. In fact, we discovered that graphene can be easily synthesized *in situ* from any carbonaceous precursor by simple electrochemical methods (Li et al., 2019).

In the present work, we present a novel electrochemical sensor based on electrochemically *in situ* synthesized graphene interface anchored with poly-arginine (Sch. 1). This is the first *p*-nitrophenol sensor based on fully electrochemically synthesized nano-interface. The sensor shows high sensitivity for *p*-nitrophenol, low detection limit as well as wide dynamic range. Ideal repeatability, stability, anti-interference ability and real-sample applications are also observed. This sensor is suitable for multiple in-the-field scenario uses with satisfactory performances, cost-effectiveness, scalability and user-friendliness. Finally, we hypothesized that poly-arginine grafting improved *p*-nitrophenol detection by optimizing graphene surface morphology and favoring H-bond and ionic interactions between arginine and nitrophenols.

2. Experimental section

2.1. Chemicals and reagents

Monopotassium phosphate (KH₂PO₄), potassium hydroxide (KOH), hydrochloric acid (HCl, 37%), potassium ferricyanide (K₃[Fe(CN)₆], 99.5%), potassium ferricyanide trihydrate (K₄[Fe(CN)₆]·H₂O, 99.0%), phosphoric acid (H₃PO₄, 99.0%), graphite powder (C, 800 mesh, 99.95%), sodium chloride (NaCl, 99.8%), potassium chloride (NaCl, 99.8%), calcium chloride (CaCl₂, 99.9%), potassium nitrate (KNO₃, 99.0%), sodium sulfite (Na₂SO₃, 99.0%), *p*-nitrophenol, *o*-nitrophenol (*o*NP, 98.0%), *m*-nitrophenol (*m*NP, 99.0%), *p*-aminophenol (*p*AP, 98.0%), catechol (99.0%), *p*-nitrobenzene (*p*NB, 99.0%), phenol (99.0%), hydroquinone (HQ, 99.5%), *p*-hydroxybenzaldehyde (*p*HBA, 98.0%), 2,3-dimethylphenol (DMP, 98.0%), bisphenol A (BPA, 99.0%) and sodium dodecylsulfonate (SDS, 98.0%) were purchased from Aladdin Biochemical Technology Co., Ltd., Shanghai, used at achievement without further purification. Other reagents are of AR grade. Ultrapure water (resistivity: $\geq 18 \text{ M}\Omega \text{ cm}^{-1}$) was used for all experiments.

2.2. Fabrication of screen printed electrode

The procedure to prepare the screen printed electrode (denoted as SPE hereafter) is presented in Scheme 1. The electrode comprises a working electrode, a counter electrode and a pseudo-reference electrode. The patterns distribute into 4 layers, *i.e.*, the polyethylene terephthalate (PET) substrate, the silver conductive/reference layer, the carbon working/counter layer and the insulative layer. All the patterns are transferred to nylon films (200 mesh) for preparing stencils by the local supplier. Silver ink (NT-ST60B, NanoTop Electronic Technology Ltd., PRC) was evenly spread onto the stencil and dragged the squeegee across the patterned area, leaving the ink transfer to the PET substrate. Curing the ink under 90 °C for 90 min to obtain the conductive areas as well as the pseudo-reference electrode. Carbon ink (C2030519P4, Gwent Electronic Materials Ltd., UK) and insulative coating (CR-18T-

CK, Asahi Chemical Research Laboratory CO., LTD., Japan) were sequentially applied with the corresponding stencils and identical procedure to that of the silver ink, to form the working electrode, counter electrode and the insulative areas, respectively. Cut the whole printed board to obtain single SPE electrodes with each has a 2.0 × 2.0 mm working electrode and counter electrode, and a 2.0 × 1.0 mm reference electrode.

2.3. Preparation of the electrodes cGO/SPE, eG/SPE and P(Arg)/eG/SPE

Preparation of chemically synthesized graphene oxide and the screen printed electrode modified thereof was modified from the literature (Marcano et al., 2010), and detailed in Supplementary Note 1, (denoted the electrode as cGO/SPE).

To prepare the electrochemically *in situ* synthesized graphene modified screen printed electrode (eG/SPE), bare screen printed electrode was immersed into 0.2 M H₃PO₄ solution contains 0.5 v/v% SDS, 1.8 V direct current potential was imposed to the working electrode vs. an external Ag/AgCl reference electrode for 200 s. Then the electrode was fully rinsed with water and processed by cyclic voltammetry scans (− 2.0 to 0 V, 100 mV/s for 10 consecutive cycles) in 0.1 M KNO₃ solution. After rinsing with water, the electrode was designated eG/SPE. The electrochemically *in situ* synthesized graphene-poly(arginine) modified screen printed electrode (P(Arg)/eG/SPE) was fabricated by immersing eG/SPE into 0.1 M phosphate buffer solution (pH 7.0) contains 20 mM L-arginine, perform a 10 cycles cyclic voltammetry scans under 50 mV s^{−1} between the potential range of − 2.0–2.0 V.

2.4. Characterizations

Scanning electronic microscope (SEM, Hitachi, X-650), Raman spectroscopy (JY Labram HR 800, 632.8 nm laser), X-ray photoelectron spectroscopy (XPS, ESCALAB 250 using Al K α radiation source), and Fourier transform infrared spectroscopy (FTIR, Nicolet 6700 with a DTGS detector and Omnic 8.0 software, Thermo Fisher Scientific, USA) were used to characterize surface state of the electrodes include morphology, chemical composition, and their changes. Unless stated otherwise, voltammetric, amperometric, and electrochemical impedance spectroscopy (EIS) experiments were performed with the build-in three-electrode system of the SPE. All electrochemical experiments were carried out with the electrochemical workstation (CHI 760E, CH Instruments Inc., USA) equipped with a 5-mL homemade reactor under ambient condition.

2.5. Molecular simulation

The calculation is based on a grid-based molecular docking method that employs CHARMM. Random ligand conformations are generated from the initial ligand structure through high temperature molecular dynamics, followed by random rotations. The Calculate Energy (DFT) protocol was used to calculate the energy or optimize the geometry of a set of small molecules using the density functional quantum mechanics method. Finally, the theoretical computational modeling were presented.

3. Results and discussion

3.1. Characterization of SPE, eG/SPE and P(Arg)/eG/SPE

We analyzed electrodes including the bare screen printed electrode (SPE), the electrochemically *in situ* synthesized graphene modified screen printed electrode (eG/SPE) and the electrochemically *in situ* synthesized graphene-poly(arginine) modified screen printed electrode (P(Arg)/eG/SPE). Cyclic voltammetry was applied to perform the *in situ* electrochemical polymerization of arginine in 0.1 M phosphate buffer solution (pH = 7.0) in presence of 2.0 mM L-arginine. (− 2.0 to 2.0 V,

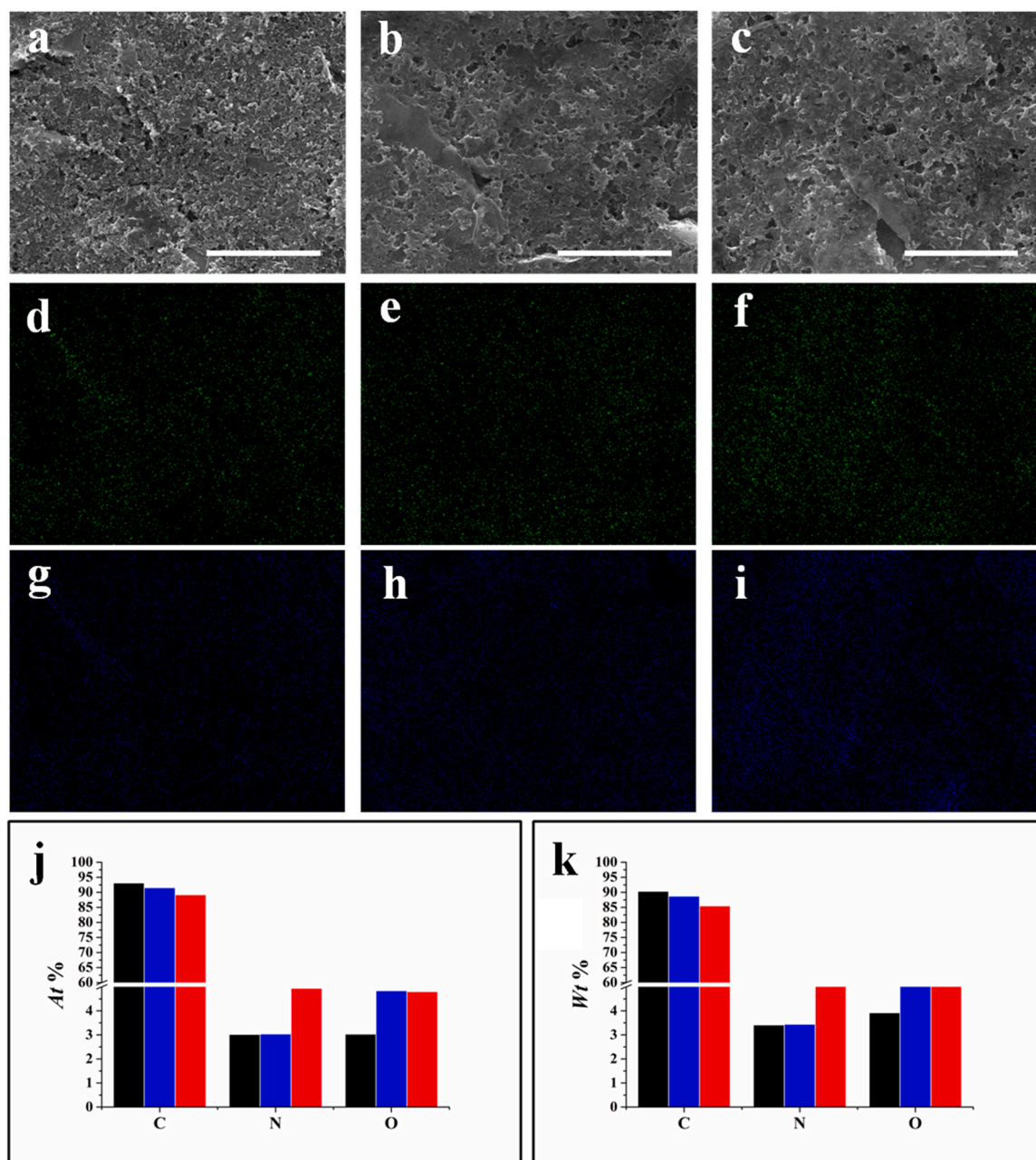


Fig. 1. Characterization of SPE, eG/SPE and P(Arg)/eG/SPE. Scanning electron microscopy of SPE (a), eG/SPE (b) and P(Arg)/eG/SPE (c). Energy dispersive spectroscopy of N (d, e, f) and O (g, h, i), and the corresponding column graph showing the atomic percentages (j) and weight percentages (k) on SPE (black), eG/SPE (blue) and P(Arg)/eG/SPE (red). (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

100 mV s⁻¹). Ten consecutive cycles of the electro-deposition process of P(Arg) onto newly prepared eG/SPE was shown in Fig. S1. Results show that the electrolytic current became weak up cycle-wise with slower rates, approximating to a finally stable state. This indicates the accumulation of poly-arginine onto the eG/SPE surface with weak or none conductivity of acquired poly-arginine, according to previous studies (Behbahani et al., 2012; Ali et al., 2020).

The morphology of the three electrodes was analyzed by SEM (Fig. 1). The scattered graphite particles are clearly visible and show the surface state of the electrode (Fig. 1a). After the electrochemical processing, graphite particles are partially dissociated with some flat, edged plates (Fig. 1b). This morphology, generally observed in graphite contained electrodes under acoustic anodization conditions (Palanisamy et al., 2014; Niu et al., 2017; González-Sánchez et al., 2018), facilitates the *in situ* synthesis of a graphene oxide layer (Li et al., 2019).

EDS mapping results show the differences of N and O contents in the raw electrode, graphene oxide electrode and polyarginine-grafted graphene oxide electrode (Fig. 1j). The oxygen increase supports the formation of graphene oxides during anodization. Then, the increase of nitrogen agrees with the formation and attachment of polyarginine on the surface (Fig. 2).

FTIR results show that arginine has distinctive peaks at 1329 cm⁻¹ and 1447 cm⁻¹, corresponding to symmetric stretching of free carboxyls, whereas these peaks decrease in poly-arginine (Fig. 2a). Similarly, the peak of C—N and N—H stretching at 3348 cm⁻¹ corresponding to free amino groups decreases in poly-arginine. The formation of amide bonds in polyarginine is supported by new peaks of C—N and N—H stretching at 1077 cm⁻¹, C—N stretching and N—H bending at 1524 cm⁻¹, and carbonyl stretching at 1644 cm⁻¹. Overall, these data confirm arginine polymerization on the electrode surface (Zhang et al.,

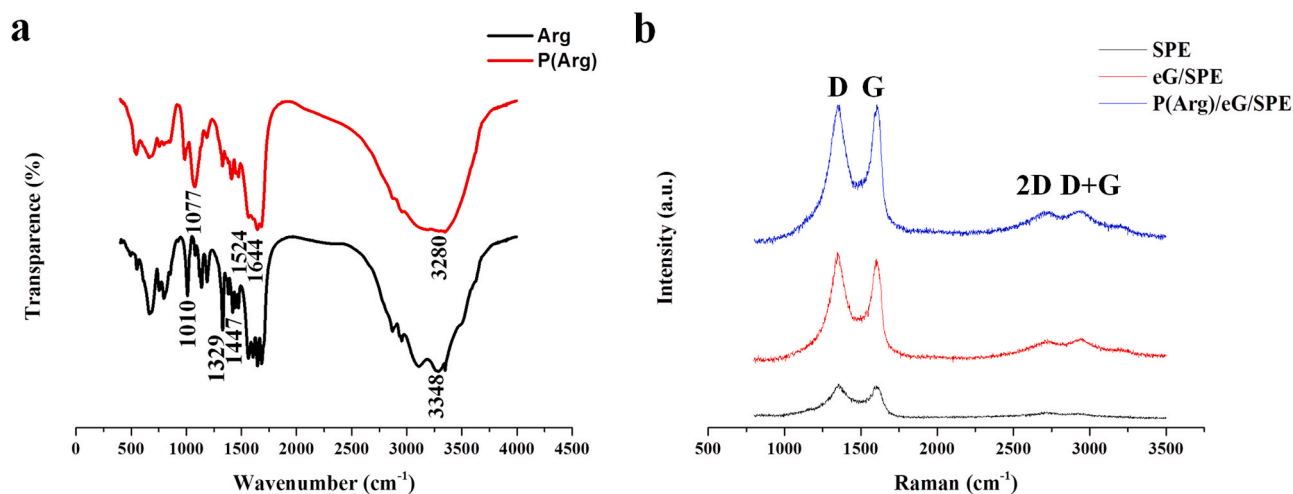


Fig. 2. FTIR (a) of Arg and P(Arg), and Raman spectra (b) of SPE, eG/SPE and P(Arg)/eG/SPE.

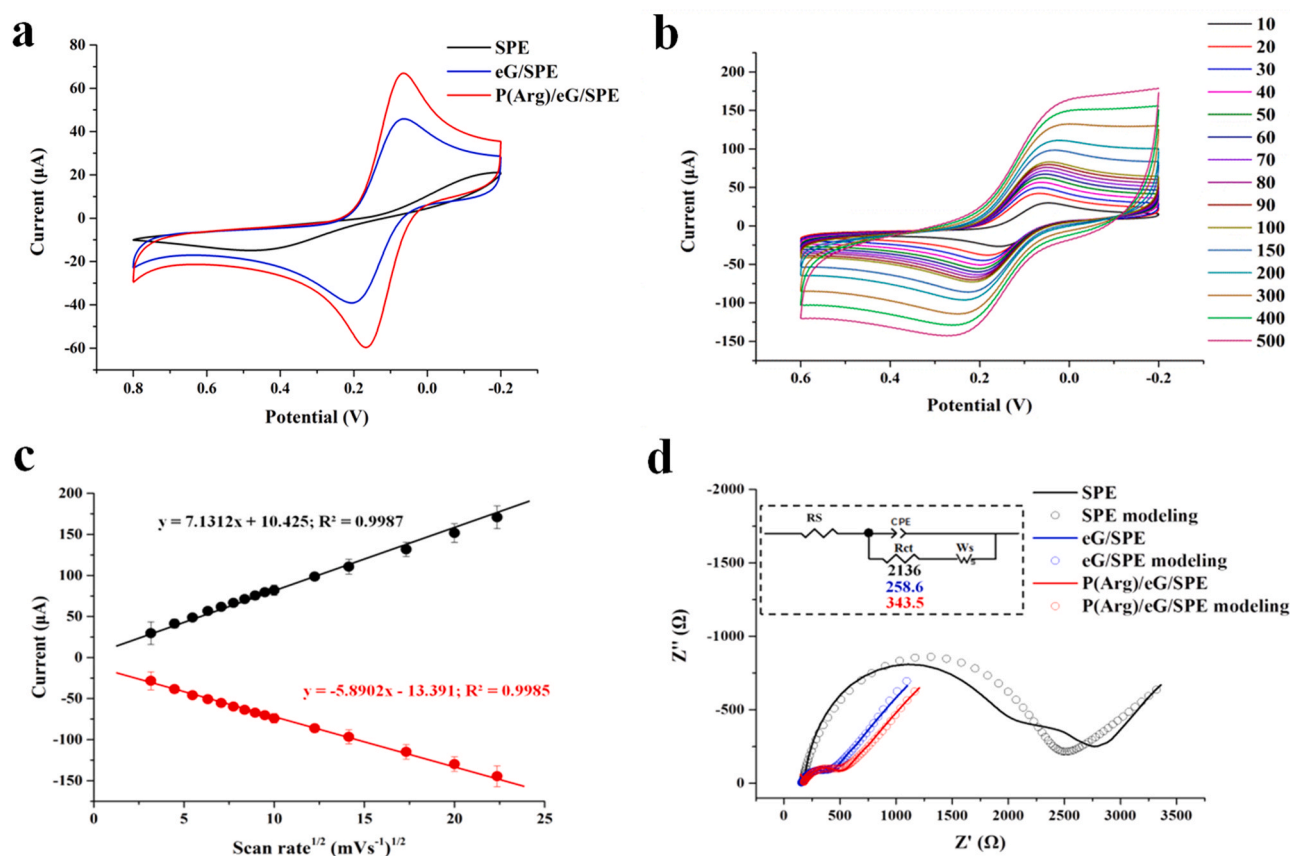


Fig. 3. Electrochemistry characterization of SPE, eG/SPE and P(Arg)/eG/SPE. Cyclic voltammograms of SPE, eG/SPE and P(Arg)/eG/SPE in 0.1 M phosphate buffer solution (pH = 7.0) containing 1.0 mM of $[\text{Fe}(\text{CN})_6]^{3-}$ and $[\text{Fe}(\text{CN})_6]^{4-}$ at scan rate of 100 mV s^{-1} (a). Cyclic voltammetry curves of P(Arg)/eG/SPE under different scan rates (b). Corresponding current-square root of scan rate relation from b (c). Nyquist plots of SPE, eG/SPE and P(Arg)/eG/SPE (d); inset showed the equivalent circuit from electrochemical impedance spectroscopy modeling (dotted curves) with the method and solution in [Supplementary Note 2](#).

2012; Li et al., 2021).

Raman spectra show two main peaks (Fig. 2b), D peak at 1323 cm^{-1} and the G peak at 1580 cm^{-1} representing in-plane vibrations of sp^2 carbons and the intrinsic structural defects on graphene lattice, respectively (Ambrosi et al., 2014). The increase of the D/G ratio from 0.955 in the raw electrode to 1.062 in the anodized electrode is explained by the rising number of carbon defects. The 2D band at $\sim 2700 \text{ cm}^{-1}$ and D + G band at $\sim 2910 \text{ cm}^{-1}$ are suggesting the

formation of a mono-layered graphenic structure, as observed previously (Li et al., 2019). The decrease of the D/G ratio from 1.062 in the anodized electrode to 0.998 in the poly-arginine grafted anodized electrode is explained by the growth of the poly-arginine layer (McCull et al., 2003).

XPS was applied to further identify the atomic changes of the electrodes. As is shown in Fig S2, C1s deconvolution spectrum of SPE shows peaks at 284.67 eV, 285.2 eV, 286.5 eV and 288.8 eV, attributed to sp^2

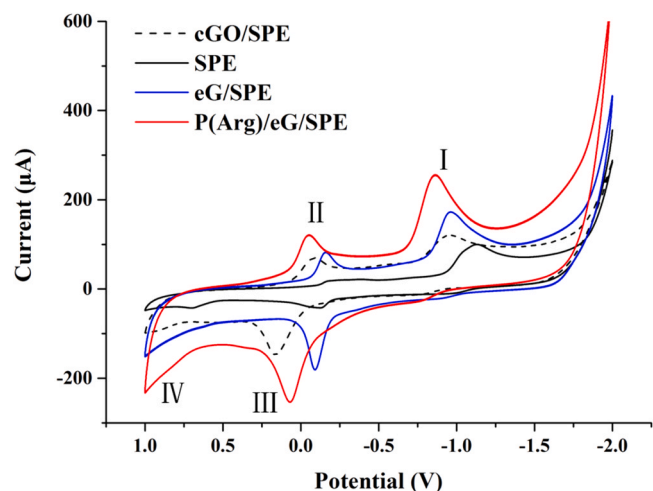


Fig. 4. Cyclic voltammograms of cGO/SPE, SPE, eG/SPE and P(Arg)/eG/SPE at scan rate of 100 mV s^{-1} in presence of 1.0 mM p-nitrophenol.

carbons, sp³ carbons, C-O and C=O/COO group, respectively. The data show the composition of graphite and the polymer binder contained in the carbon ink (Fig S2a). For eG/SPE, the ratio of sp² to sp³ increased, indicating the *in situ* production of graphene at the electrode surface (Fig S2b). For P(Arg)/eG/SPE, a new peak at 285.9 eV attributed to C-N linkage is observed, indicating the formation of P(Arg) (Fig S2c) (Ayiania et al., 2020). This peak was also found in the monomeric

arginine. By comparison, the C-NH₂ peak at 288.0 eV largely depleted in polyarginine, as previous report (Zhang et al., 2014). Overall, these data confirm electrochemical polymerization of arginine on the electrode.

To further understand the properties of these electrodes, electrochemical behaviors were first studied by cyclic voltammetry method. As shown in Fig. 3a, a characteristic reversible peak pair can be observed, showing their electro-catalytic activity towards ferro/ferricyanide ions. But manifestly, SPE (black) exhibited the worst electrochemical performance with lowest peak currents ($I_{pa} = 43.0 \mu\text{A}$), and a large peak potential difference (ΔE_p) value, indicating a slow electrochemical kinetics. In comparison, eG/SPE markedly enhanced the performance. The I_{pa} enlarged to $329 \mu\text{A}$ and the ΔE_p value reduced to $\sim 135 \text{ mV}$. For P(Arg)/eG/SPE, I_{pa} further increased to $592.8 \mu\text{A}$ and the ΔE_p value decreased to $\sim 85 \text{ mV}$. These data directly displayed the highly electro-catalytic activity of P(Arg)/eG/SPE to the surface dependent electro-activity species. Further, Fig. 3b and c provided the results with P(Arg)/eG/SPE to detect ferro/ferricyanide ions at different scan rates. Within the range of $10\text{--}500 \text{ mV s}^{-1}$, good linear correlation was obtained between the peak currents and their corresponding square root of scan rates ($I_{pc} = 7.1312c + 10.425$, $R^2 = 0.99$; $I_{pa} = -5.8902c - 13.391$, $R^2 = 0.99$). This indicated the electrochemical reaction of the surface dependent species followed a diffusion controlled kinetics (Li et al., 2018), which is advantageous to sensor applications for the target of the same category.

For all three electrodes in the high frequency range analyzed by electrochemical impedance spectroscopy (Fig. 3d), a semi-circular region always appeared and became the predominate variable, indicating that the electron transfer resistance, R_{ct} , is the main factor changed on

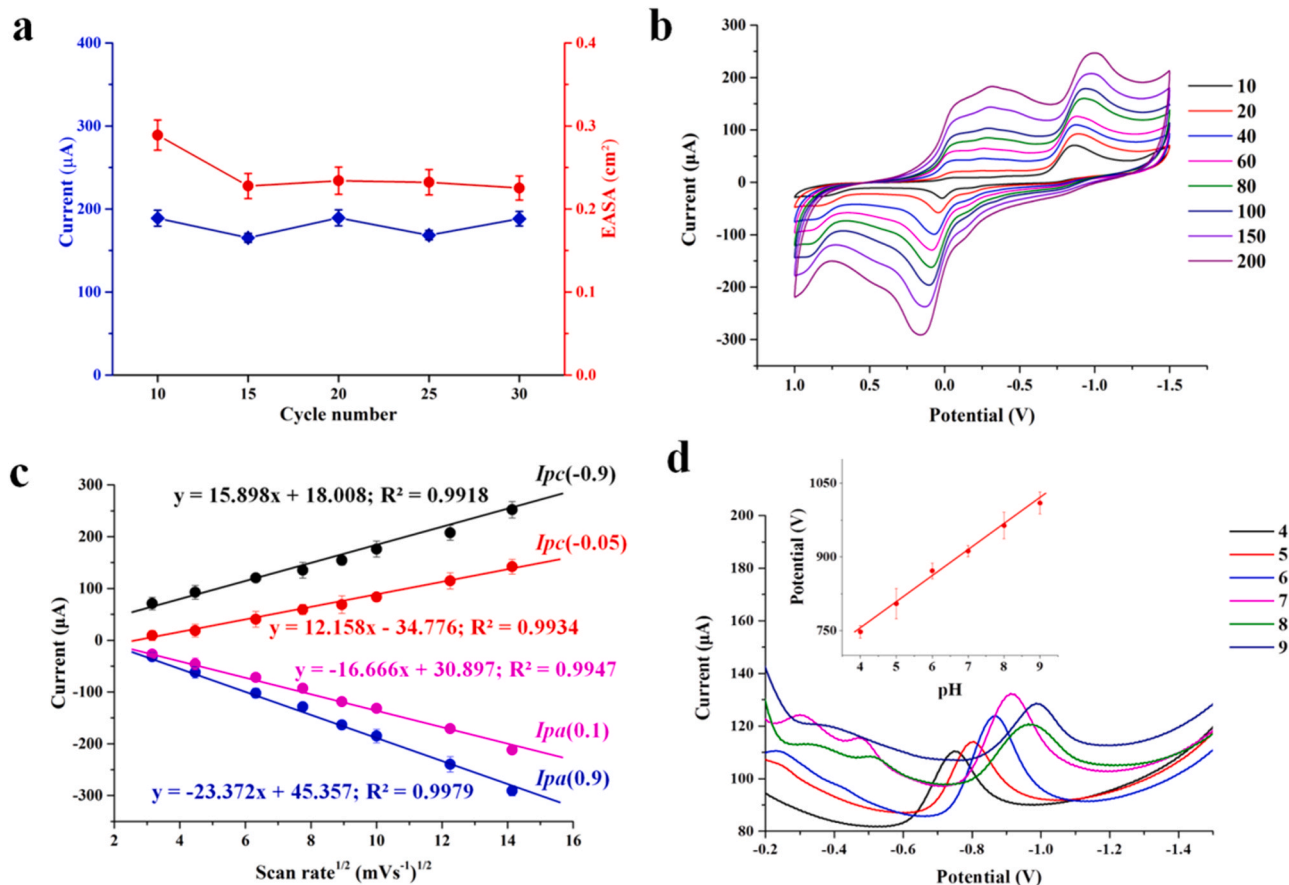


Fig. 5. p-Nitrophenol electro-catalytic current and electrochemically active surface areas (EASA) changes on the P(Arg)/eG/SPEs prepared by different cycle numbers of poly-arginine electro-deposition (a). Cyclic voltammograms of P(Arg)/eG/SPE in 0.1 M phosphate buffer solution ($\text{pH} = 7$) in presence of 1 mM p-nitrophenol at different scan rates (b). The corresponding current-square root of scan rate relation from a (c). Square wave voltammetry responses for 0.5 mM p-nitrophenol in 0.1 M phosphate buffer solution of different pH; the inset showing the relation between the peak potential and pH (d).

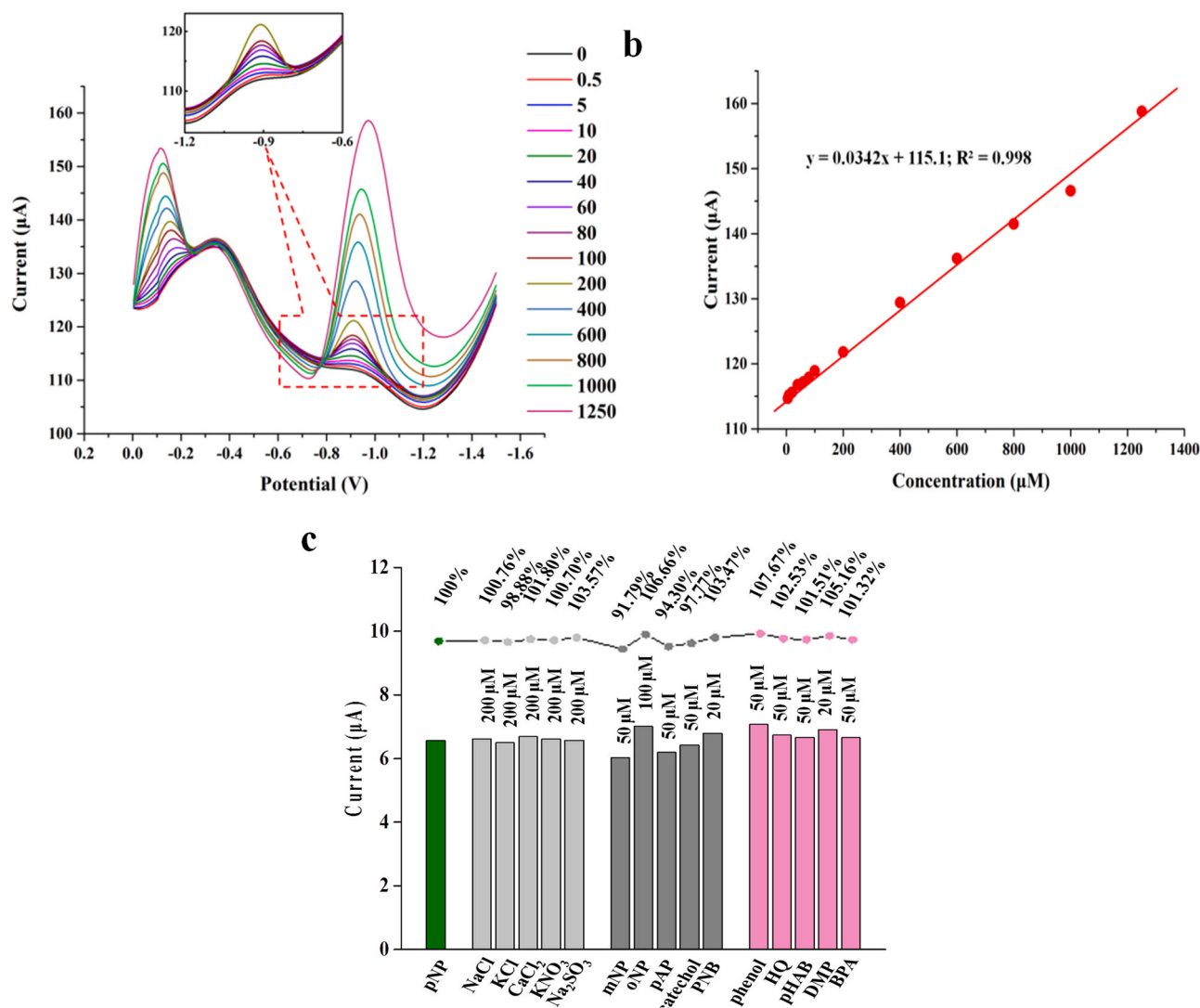


Fig. 6. Square wave voltammetry responses of P(Arg)/eG/SPE to diverse concentrations of *p*-nitrophenol (a). Corresponding relation between peak current and concentration (b). Selectivity of P(Arg)/eG/SPE over the foremost possible interfering compounds (c).

the electrodes. By further modeling and calculation, the equivalent circuits were acquired (Fig. 3d, inset). R_{ct} drastically declined from 2136 Ω on SPE to 258.6 Ω on eG/SPE, showing the high conductance of the *in situ* synthesized graphene layer. The R_{ct} value of P(Arg)/eG/SPE, however, increased weakly back to 343.5 Ω , revealing the weak conductivity of poly-arginine. This is consistent with the cyclic voltammetry result aforementioned. Actually, the *in situ* synthesized graphene layer predominantly contributed to the electro-catalytic activity and the surface conductivity. P(Arg) mainly enhanced the electrode interface to exert higher electro-catalytic activity (Fig. 4).

3.2. Electrochemical performances for *p*-nitrophenol detection

We compared the electro-catalytic performance of bare screen printed electrode (SPE), the electrochemically *in situ* synthesized graphene modified screen printed electrode (eG/SPE) and the electrochemically *in situ* synthesized graphene-poly(arginine) modified screen printed electrode (P(Arg)/eG/SPE), along with the counterpart, chemically synthesized graphene oxide modified screen printed electrode (cGO/SPE) in the presence of nitrophenol (Fig. 4). Peak I at -1200 mV is the irreversible reduction of nitrophenol. The reversible peak II at -200 mV and peak III at -170 mV represent the reduction of *p*-hydroxylaminophenol and *p*-nitrosophenol (Renu et al., 2021; Huang

et al., 2021). Peak IV at 750 mV is attributed to the oxidation of the phenolic moiety of nitrophenol. Results show that the polyarginine-grafted electrode displays the best performance with the highest currents, versus the oxidized electrode and the classical graphene oxide electrode, cGO/SPE. The raw electrode showed the weakest performance. The best potential for nitrophenol monitoring is at peak I.

3.3. Effect of electro-deposition of poly-arginine, pH and scan rate on the electro-catalytic performance

We studied the effect of the electrochemically active surface area, pH and scan rate on the performance of the polyarginine-grafted graphene electrode. Areas were changed by varying cyclic voltammetric deposition cycles. We observed little variation of current intensity from 10 to 30 cycles, whereas electrochemically active surface areas decreased above 10 cycles (Fig. 5a). Similar phenomena were observed in the previous reports (Zhang et al., 2012; Li et al., 2021). So 10 cycles were adopted for further experiments. The electrochemical behaviour of the electrode in response to nitrophenol at different scan rates is shown in Fig. 5b and linear correlations are observed at peak I, II, III and IV (Fig. 5c). Findings show that the nitrophenol transformation is controlled by diffusion in the 10–200 mV s^{-1} range. The study of the effect of pH shows that the polyarginine electrode reached optimum current at pH 6 and pH is well

Table 2
Recoveries of *p*-nitrophenol in real-world samples (n = 3).

Sample	Standard addition (μM)	Fund (μM)	Recovery %	SD %
Industrial wastewater	5	5.02	100.4	1.7
	25	24.59	98.4	1.2
	50	49.63	99.3	0.6
Lab. wastewater	5	4.88	97.6	1.4
	25	25.20	100.8	1.0
	50	50.14	100.3	0.7
River water	5	4.77	95.4	2.3
	25	24.81	99.2	1.4
	50	50.30	100.6	1.1

correlated with potential according to $E_p = 52.229 \text{ pH} + 545.65$ with r^2 of 0.99 (Fig. 5d). This finding indicates that the electro-reduction of nitrophenol is a proton-coupled process with equimolar transfer number of H^+ and e^- in a wide acidity range (Zhu et al., 2020).

3.4. Voltammetric performances for sensitive *p*-nitrophenol detection

Square wave voltammetry was applied to examine P(Arg)/eG/SPE. Testing parameters are shown in Supplementary Note 3. Fig. 6a showed that P(Arg)/eG/SPE presented clear responses to *p*-nitrophenol concentration within range of 0–1250 μM. A good linear relationship was obtained between the I_{pc} values and the concentration of *p*-nitrophenol with the range of 0.5–1250 μM (Fig. 6b). The regressive equation was $I_{pc} (\mu\text{A}) = 0.0342 C (\mu\text{M}) + 115.1$ ($R^2 = 0.99$). The sensitivity of the sensor was calculated to be $0.66 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$ by the Eq. (1) (Chakraborty et al., 2021),

$$\text{Sensitivity} = k/A \quad (1)$$

where, k designates the slop of the above regressive equation, A is the electrochemically active surface area calculated in Supplementary Note 4, with the calculated $A = 0.0514 \text{ cm}^2$.

The limit of detection (LOD) was determined as 12 nM, calculated following the criterion of $S/N = 3$ (Su et al., 2021), using Eq. (2),

$$\text{LOD} = 3N/k \quad (2)$$

N = standard deviation value of the response; k = calibration curve slop.

Comparing with the previous results in Table 1, it can be seen the P(Arg)/eG/SPE fabricated in this research has excellent quantification performances as a *p*-nitrophenol electrochemical sensor.

Reproducibility of this sensor was evaluated by testing the square wave voltammetry responses with 5 P(Arg)/eG/SPE sheets prepared independently in 0.1 M phosphate buffer solution (pH = 7) containing 1.0 μM *p*-nitrophenol. The relative standard deviation (RSD) was found to be 5.2%. RDS was 1.2% for the repeatability analysis, which was tested by performing 5 independent measures in the same solution with each in triplet (Supplementary Fig S3a). For stability investigation, 3 newly prepared P(Arg)/eG/SPE sheets were stored under ambient condition and tested in the same solution every 24 h for 7 days. Over 90% signal retention rate was gotten within the 7-day experiment (Fig S3b and c).

In the real world, some possible interferences may cause detection distortion to the sensor (Su et al., 2021; Renu et al., 2021). We therefore chose some most likely interferent to investigate the sensor's selectivity. To investigate the selectivity of the sensor, we used 3 groups of possible

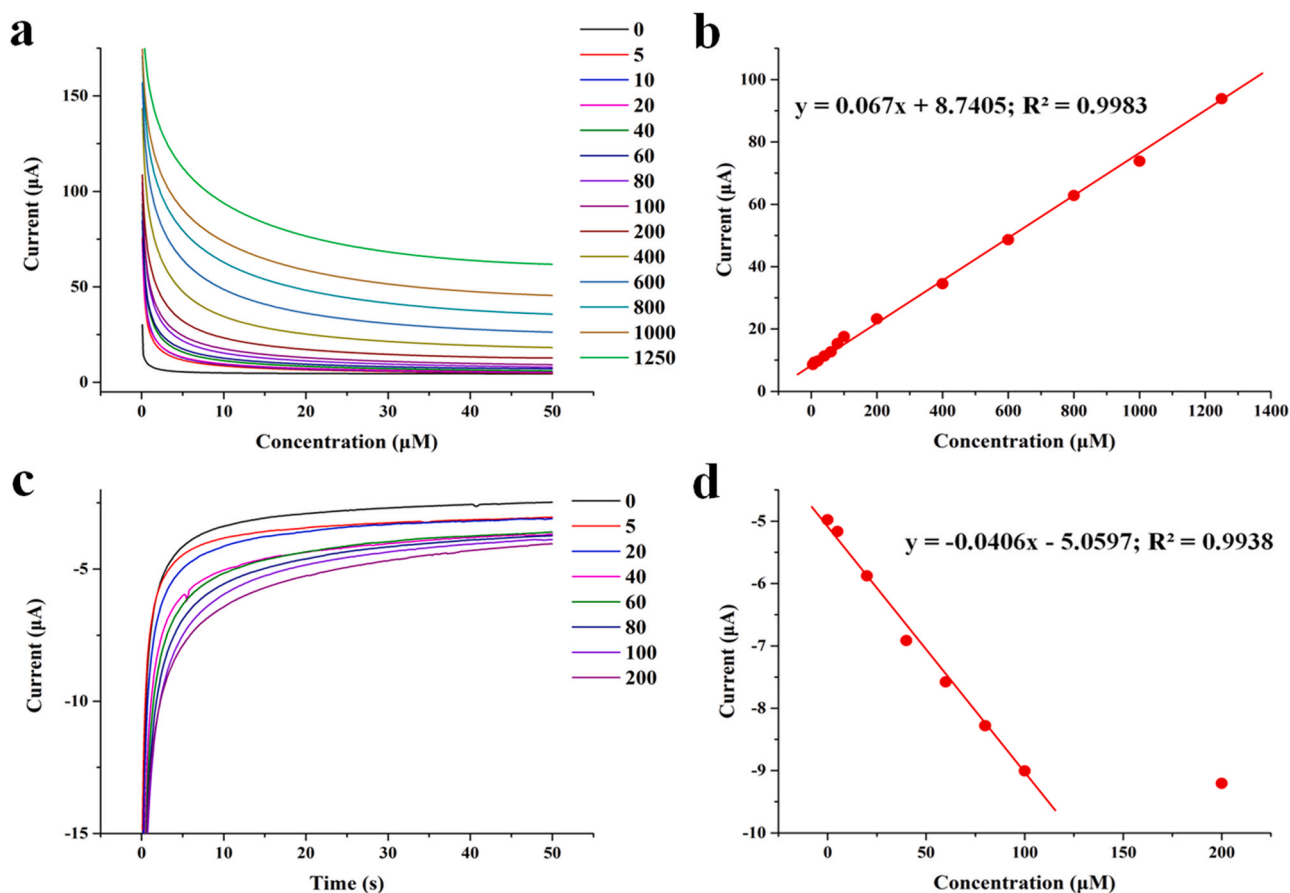


Fig. 7. Amperometric responses of P(Arg)/eG/SPE at -800 mV (a) and 800 mV (c) in presence of *p*-nitrophenol with different concentrations. The corresponding regressive relation between current and *p*-nitrophenol concentration sampled at 3rd second.

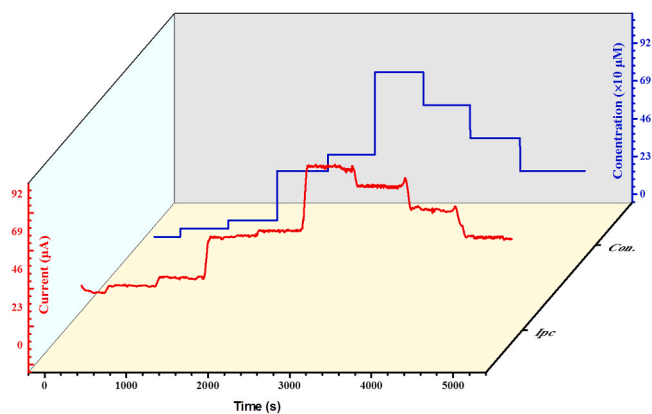


Fig. 8. Amperometric responses (red) of P(Arg)/eG/SPE to the pulsing concentrations of *p*-nitrophenol (blue). ($\times 10 \mu\text{M}$ means to multiply ten to the corresponding number on the Y-axis, in the unit of μM)

interferent to perform the anti-interference test (Fig. 6c). In the first group, common co-existing ions include Na^+ , K^+ , Ca^{2+} , Cl^- , NO_3^- , SO_3^{2-} with almost no influence to the sensing signal of *p*-nitrophenol. In the 2nd group, homologs and isomers of *p*-nitrophenol are studied with signal recovery ratios ranging from 91.79% to 106.66%. The 3rd was group composed of phenols which are also homologous to *p*-nitrophenol. Slight signal deviation from 101.32% to 107.67% is detected. For all experiments, the signal recovery ratios are within range of 91.79–107.67%, with the most cases less than 5.0%. Among these, isomer *o*-nitrophenol and phenol caused the most prominent interference due to high homology of the structure to *p*-nitrophenol, yet their influence is acceptable to real applications.

Overall, results show good selectivity and anti-interference of the sensor. Moreover, real-world samples were used to perform the standard addition tests. As the results shown in Table 2, in all cases the error produced in the complex samples were less than 3%, which further demonstrated a worthy applicability. Collectively, P(Arg)/eG/SPE is an ideal sensor for portable *p*-nitrophenol detection with high sensitivity, wide quantification range and good reliability.

3.5. Amperometry for long-term *p*-nitrophenol monitoring

Many cases require long-term, consecutive quantifications of *p*-nitrophenol level other than the need for one-off detection. Considering this, the ability of the sensor to continual *p*-nitrophenol monitoring was

investigated. The *i-t* curves recorded in 0.1 M phosphate buffer solution ($\text{pH} = 7.0$) in presence of *p*-nitrophenol with various concentrations (Fig. 7). Data in Fig. 7a and c were obtained using peak I and peak IV with the working potential of -800 mV and 800 mV , respectively. Regressive curves were obtained by sampling the current values at the 3rd second of each curve (Fig. 7b and d). The regressive equations were gotten as follow: $I_{pc} (\mu\text{A}) = 0.067 C (\mu\text{M}) + 8.7405$ ($R^2 = 0.99$) and $I_{pa} (\mu\text{A}) = -0.041 C (\mu\text{M}) - 5.0597$ ($R^2 = 0.99$). The corresponding linear concentration ranges were $5\text{--}1250 \mu\text{M}$ and $5\text{--}200 \mu\text{M}$. Limit of detections were 2.4 and 3.9 nM , and sensitivities were 1.30 and $0.79 \mu\text{A} \mu\text{M}^{-1} \text{cm}^{-2}$. It is noticeable that for measuring higher concentration gradient *p*-nitrophenol samples under 800 mV , the catalytic current I_{pa} no longer increased or even weakened (data not shown). Electro-oxidation of *p*-nitrophenol may lead to *in situ* self-polymerization, along with the irreversible adsorption of the oxidation product (Chen et al., 2008; Karabiberoglu et al., 2019). The great surface fouling could largely passivate the sensor. Therefore, the reduction mode seemed to be preferable for the long-term monitoring.

A long-term consecutive monitoring of varied concentration of *p*-nitrophenol in phosphate buffer solution base is shown in Fig. 8. As the concentration of *p*-nitrophenol changed, the reductive current intensity presented a swift response. Even when *p*-nitrophenol concentration dropped to lower level, the sensor responded quickly with a in-step signal, indicating that this electrode was fouling-free and adaptive for consecutive monitoring.

3.6. Molecular modeling

Theoretical computational modeling was performed to study the interactions of *p*-nitrophenol with the P(Arg)/eG/SPE. According to the potential binding energy (ΔE_{AB}) predictions, two models were considered. The first one illustrated the arrangement of a *p*-nitrophenol molecule freely onto a graphene sheet. It was modeled with an in-plane defect which is common in electro-catalytic analyses (Liu et al., 2018; Wang et al., 2019a, 2019b). As shown in Fig. 9a, *p*-nitrophenol nearly parallelly attached to the graphene plane with the closest inter-molecular distance of 3.51 \AA (at the phenol moiety of *p*-nitrophenol), which is mainly induced by strongly non-specific adsorption. This is the reason why only limited performance gain harvested and the electrode fouling is severe on bare graphene interfaces.

In comparison, the existence of poly-arginine makes *p*-nitrophenol molecule re-orientate on graphene interface through comprehensive effect from H-bonding, ionic attraction and hydrophobic force (Fig. 9b). In this second model, *p*-nitrophenol is tightly and vertically located at the defect of graphene plane with their distance reduced to $\sim 2.25 \text{ \AA}$,

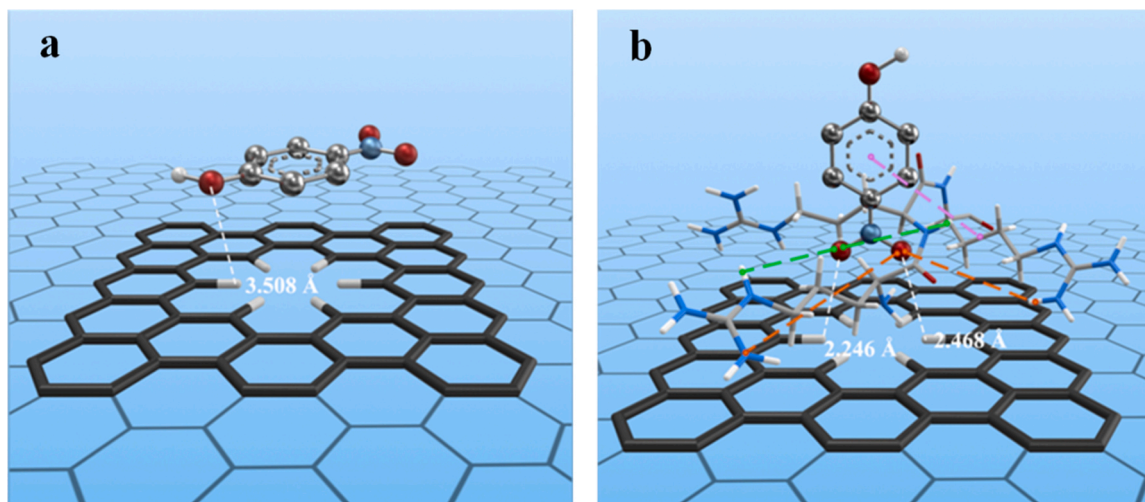


Fig. 9. Possible optimized configuration of *p*-nitrophenol-P(Arg)-graphene interactions (b) and that without P(Arg) (a) visualized by Rosetta package.

revealed by density function theory calculation result. More importantly, poly-arginine benefit to drawing $-NO_2$ moiety of *p*-nitrophenol towards the defect - the electrocatalytically active site on graphene plane. Consequently, P(Arg) assist preferable orientation of *p*-nitrophenol onto the sensing interface and present a synergy with *in situ* synthesized graphene to endow P(Arg)/eG/SPE with fast electrochemical kinetics. This verifies the vital importance of delicate molecular design of sensing interface to create optimized chemical micro-environment.

4. Conclusion

A fully electrochemically synthesized graphene screen printed electrode interface with poly-arginine anchored was successfully designed and applied for *p*-nitrophenol sensing. Using voltammetric technology, P(Arg)/eG/SPE fulfilled high sensitive *p*-nitrophenol detection with limit of detection as low as 12 nM. Moreover, a wide range quantification of 0.5–1250 μ M was fitted. By adapting amperometric method, highly sensitive and long-term consecutive *p*-nitrophenol level monitoring can be realized, with the limit of detection of 2.4 nM and linear concentration range of 5–1250 μ M. Specially, it is revealed by density function theory calculation that poly-arginine facilitates *p*-nitrophenol molecules to preferably orientate onto the graphene interface with fast electrochemical kinetics. Considering the very low cost of screen printed electrode fabrication, this sensor is an ideal prototype for the next generation versatile *p*-nitrophenol electrochemical sensing devices.

CRedit authorship contribution statement

Y.L. and L.X. designed the research. Y.L. J.Z. and S.W. conceptualized and performed the experiments. Y. L. and Y. M. contributed reagents, materials and analysis tools. Y. L., Y. M., J.S., L.X., and E.L. analyzed the data. R. G. performed the molecular simulation calculation. Y. L., J.S., L. X. and E.L. wrote and revised the manuscript. All the authors discussed the results and commented on the manuscript.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at [doi:10.1016/j.jhazmat.2021.126718](https://doi.org/10.1016/j.jhazmat.2021.126718).

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