

doi:10.1093/aobpla/plaa047 Advance Access Publication September 11, 2020 Studies

# STUDIES

# Difference between emergent aquatic and terrestrial monocotyledonous herbs in relation to the coordination of leaf stomata with vein traits

Wanli Zhao<sup>1,2,3,4,#</sup>, Peili Fu<sup>5,6,#</sup>, Guolan Liu<sup>4</sup> and Ping Zhao<sup>\*1,2,3</sup>

<sup>1</sup>Key Laboratory of Vegetation Restoration and Management of Degraded Ecosystems, South China Botanical Garden, Chinese Academy of Sciences, Guangzhou, China, <sup>2</sup>Center of Plant Ecology, Core Botanical Gardens, Chinese Academy of Sciences, Guangzhou, China, <sup>3</sup>Guangdong Provincial Key Laboratory of Applied Botany, South China Botanical Garden, Chinese Academy of Sciences, Guangzhou, China, <sup>4</sup>Shandong Key Laboratory of Eco-Environmental Science for Yellow River Delta, Binzhou University, Binzhou, China, <sup>5</sup>CAS Key Laboratory of Tropical Forest Ecology, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Yunnan 666303, China, <sup>6</sup>Ailaoshan Station of Subtropical Forest Ecosystem Studies, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Sciences, Jingdong, Yunnan 676209, China

\*Corresponding author's e-mail address: zhaoping@scib.ac.cn

\*These authors contributed equally to this paper.

Associate Editor: Kristine Crous

Form & Function. Chief Editor: Kate McCulloh

## Abstract

Emergent aquatic plants mostly occur in shallow waters and root in bottom substrates, but their leaves emerge from the water surface and are thus exposed to air, similar to the leaves of terrestrial plants. Previous studies have found coordination between leaf water supply and demand in terrestrial plants; however, whether such a coordination exists in emergent aquatic plants remains unknown. In this study, we analysed leaf veins and stomatal characteristics of 14 emergent aquatic and 13 terrestrial monocotyledonous herb species (EMH and TMH), with 5 EMH and 8 TMH belonging to Poaceae. We found that EMH had significantly higher mean leaf area, leaf thickness, stomatal density, stomatal number per vein length and major vein diameter, but lower mean major vein length per area (VLA) and total VLA than TMH. There was no significant difference in stomatal length, minor VLA and minor vein diameter between the two groups. Stomatal density and total VLA were positively correlated among the EMH, TMH, as well as the 8 Poaceae TMH species, but this correlation became non-significant when data from both the groups were pooled. Our results showed that the differences in water supply between emergent aquatic and terrestrial plants modify the coordination of their leaf veins and stomatal traits.

Keywords: Leaf hydraulics; optimization; Poaceae; stomatal density; vein density.

## Introduction

In leaves, the xylem supplies water to the photosynthetic tissues to prevent their desiccation during photosynthetic  $CO_2$  exchange with the atmosphere (Brodribb *et al.* 2007; Feild and Brodribb 2013). Stomata control gas exchange between the leaves and atmosphere (Hetherington and Woodward 2003; Simonin and Roddy 2018). Therefore, both stomatal density and size play

vital roles in controlling maximum transpiration, i.e. leaf water demand (Franks and Beerling 2009). Leaf veins transport water from the petiole across the lamina to the mesophyll mainly for transpiration (Niklas 1999). Indeed, previous studies have shown that vein density (vein length per leaf area, VLA) is a key determinant of leaf water supply capacity in terrestrial plants

Received: 16 January 2020; Editorial decision: 25 August 2020; Accepted: 8 September 2020

© The Author(s) 2020. Published by Oxford University Press on behalf of the Annals of Botany Company. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited. (Sack and Scoffoni 2013; Scoffoni and Sack 2017). Coordination between stomatal density and VLA across species indicates different strategies for the maintenance of water balance (Brodribb et al. 2013; Schneider et al. 2017). A positive correlation between the minor VLA and stomatal density has been found in many species across different habitats (Zhang et al. 2012; Carins Murphy et al. 2016; Zhao et al. 2016). However, we still know little about this relationship in aquatic plants.

Although many species show coordination between stomatal density and VLA, some species have unique strategies for maintaining water balance. For example, no significant positive correlation had been found between VLA and stomatal density among terrestrial and epiphytic Cymbidium species, which is mainly due to the high water storage capacity of these species (Zhang et al. 2015). The high capacitance buffers water potential declines in the transpiration stream, and then reduces the dependence of transpiration on water uptake from the soil (Meinzer and Grantz 1991; Ogburn and Edwards 2013; Roddy et al. 2018). Available water resources play important roles in leaf venation development (Uhl and Mosbrugger 1999; Roddy et al. 2019), and a negative correlation has been found between VLA and water availability in several herbs (Napp-Zinn 1988). Whereas a positive correlation between these factors has been found in plants growing under nearly saturated air humidity in tropical rainforests (Pyykkö 1979). Under arid conditions, some species have apparent over-investment in leaf venation to compensate for the adverse effect that the thicker leaves have on photosynthesis (de Boer et al. 2016). In contrast, plants growing in water may have lower drought stress given their submergence in water, whereas exposure of their leaves to air may render a high vapour pressure deficit that may influence the evolution of vein and stomatal traits (Fanourakis et al. 2011; Hovenden et al. 2012; Carins Murphy et al. 2014).

The coordination between VLA and stomatal density in maintaining homeostasis in leaf water content is crucial for continued physiological function (Brodribb et al. 2011; Roddy et al. 2020). Zhao et al. (2016) found that tree species in subtropical mountain forest had lower VLA, but similar stomatal density when compared with tree species in a tropical mountain forest, which caused significant differences in the coordination between stomatal density and VLA between these two types of forests. The stomatal number per vein length, which is calculated from dividing stomatal density by VLA (Zhao et al. 2017), could also be used to compare the difference in the coordination between leaf water supply and demand. Zhao et al. (2017) found that three leguminous species under certain environmental conditions had stable stomatal number per vein length, which showed the coordination between leaf water supply and demand. When the environmental conditions changed, the stomatal number per vein length would change accordingly. At present, studies comparing the differences between stomatal number per vein length and VLA and stomatal density are rare.

Emergent aquatic plants mostly occur in shallow waters and root in the bottom substrates, but their leaves emerge from the water surface and are thus exposed to air, which are similar to terrestrial plant leaves (Golub *et al.* 1991; Lacoul and Freedman 2006). Approximately 11 % of monocotyledonous plants are aquatic (Les and Schneider 1995; Lacoul and Freedman 2006; Conklin *et al.* 2019). Most monocots have a distinct hierarchy of gridded 'parallel' or 'striate' major veins with midribs, large and intermediate longitudinal veins that are analogous to major vein orders and small longitudinal veins and transverse veins that are analogous to minor veins (Ueno *et al.* 2006; Sack and Scoffoni 2013). The pan-tropical Ochnaceae species have dense major veins, whereas the coordination of total VLA and stomatal density across 55 species in this family is maintained (Schneider *et al.* 2017). The dense major veins also exist in emergent aquatic monocotyledonous herbs (EMH), while the relationship between the total VLA and stomatal density of those species is still unclear.

Noticeably, water supply is not a limiting factor for EMH, but it is for terrestrial monocotyledonous herbs (TMH), especially in the dry season. With sufficient water supply, plants typically have higher photosynthesis and transpiration rates (Passioura 2002). Under sufficient water supply, a low VLA has the potential benefits of reducing construction costs and displacing mesophyll (Baresch et al. 2019; Sporck and Sack 2010). In this study, we chose EMH as the subject and TMH as the baseline to clarify the relationship between VLA and stomatal density in these two groups. We hypothesized that compared with terrestrial species, the emergent aquatic species would have higher vein diameter but lower VLA, and higher stomatal number per vein length. The results of this study may deepen the understanding of the relationship between leaf vein and stomatal traits.

#### **Methods**

#### Site and sampling

This study was carried out in the South China Botanical Garden (SCBG; 23°10'N, 113°21'E, elevation 41 m), Chinese Academy of Sciences, Guangzhou City, Guangdong Province, China. The mean annual temperature in the garden is 21.7 °C and the mean annual precipitation is 1761 mm (with more than 80 % rain from May to September).

EMH grow in a shallow freshwater pool at SCBG (Fig. 1) and TMH are common and grow along the roads. We collected 4–6 mature leaves from 4 to 6 individual plants in full sunlight of each species and stored them in a refrigerator at 4 °C in July 2017. In total, 14 EMH, belonging to eight families and 13 TMH belonging to six families, were collected (Table 1). As 5 EMH and 8 TMH of the 27 species in this study belonged to Poaceae (EMH<sub>p</sub> and TMH<sub>p</sub>, respectively), we also checked our hypothesis at the family level, which would reduce the influence of phylogenetic factors.

The leaves were scanned using an HP CLJM277 scanner, and leaf area was measured using Image J software (http://rsbweb. nih.gov/ij/index.html).

# Stomatal density and modelled maximum stomatal conductance

As all the species in this study were amphistomatous, stomatal density, stomatal length and width (refer to guard cell length and width, respectively) were determined from both adaxial and abaxial cuticles of EMH and TMH. The leaves were prepared and measured following the protocols of Zhao *et al.* (2016). Because of stomatal size–number trade-off, we also estimated the theoretical modelled maximum stomatal conductance ( $g_{max}$ ) as reported by Franks and Farquhar (2001):

$$g_{\max} = \frac{d}{v} \times D \times \frac{a}{l + (\pi/2)\sqrt{a/\pi}},$$
(1)

where *d* is the diffusivity of water in air (m<sup>2</sup> s<sup>-1</sup>); v is the molar volume of air (m<sup>3</sup> mol<sup>-1</sup>); *D* is the stomatal density; *a* is the maximum pore area approximated as  $\pi(p/2)^2$ , where *p* is 1/2 guard cell length; *l* is the pore depth that is represented by 1/2 guard cell width, assuming guard cells inflate to a circular cross-section (Franks and Beerling 2009).



Figure 1. Habitat of emergent aquatic (A) and terrestrial (B) herbs. Photos by WL Zhao and QG Mao.

#### Leaf vein measurements and categories

The leaves that had been used to measure stomatal traits were also used to measure the VLA. The leaves were placed in bottles containing 5 % NaOH aqueous solution and were heated in a water bath (Yiheng HWS24, Shanghai, China) until the veins were exposed. We used distilled water to soak the leaves for 30 min, and then the leaves were dyed with 1 % methylene blue solution, rinsed again, mounted on slides and photographed. For the species with reticulate pattern veins, we distinguished vein order hierarchy according to Sack et al. (2012), and for other species with parallel or striate venation, we distinguished vein order hierarchy according to Christin et al. (2013). Image J (http://rsbweb.nih.gov/ij/index.html) was employed to measure the VLA of different vein categories. Although the transverse veins of parallel or striate venation have an important role in leaves, the proportion of them was small in the whole leaf venation (McKown and Dengler 2010; Lundgren et al. 2019), and we excluded them in this study. We measured major VLA (1°-3°) separately, but for minor VLA, 4° and higher orders were grouped into one class. The major vein diameters were performed including the bundle sheath, and we measured the different orders from the middle of the leaves, and the mean minor vein diameter was calculated for orders 4° and higher. Stomata number per vein length (no. mm<sup>-1</sup>) was calculated from dividing stomatal density by total VLA.

We roughly estimated the xylem construction cost of leaf veins with a dimensionless index of cell wall volume per leaf area (CC, McKown et al. 2010). A modified, yet simplified, method of Schneider et al. (2017) for lumen diameter and conduit density per vein order determination was applied for total vein diameter determination based on the assumption that both variables correlate with vein diameter. Thus, we used the following equation to calculate the xylem construction cost of leaf veins:

$$CC = \sum_{i=1}^{\nu} \pi \times d_i \times D_i, \qquad (2)$$

where  $d_i$  is the diameter of vein order i and  $D_i$  is the density of the same order.

#### Leaf thickness

After measuring stomatal density and VLA, the same leaves were used to measure leaf thickness using freehand sections. The leaf sections were placed in water, and then mounted on slides and photographed. We measured the leaf thickness using Image J (http://rsbweb.nih.gov/ij/index.html) software.

#### Data analysis

Independent t-tests were used to assess differences in leaf functional traits between EMH and TMH. Correlations between leaf traits were analysed with Pearson's correlation coefficients. Principal component analysis (PCA) was used to analyse the correlations among the 18 plant functional traits and the distributions of the 27 species along the PCA axes by using 'FactoMineR' and 'factoextra' packages in R ver. 3.6.3 (R Core Team 2020). The phylogeny tree of the 27 studied species was generated from Phylomatic web site (http://phylodiversity.net/ phylomatic/) by using the stored tree 'zanne2014' (Zanne et al. 2014). Phylogenetic ANOVA were used to test the differences in

code	Species	Family	C3/C4	Leaf vein type
	EMH			
1	Alisma canaliculatum	Alismataceae	C3	RV
2	Arundo donax	Poaceae	C3	PV
3	Canna glauca	Cannaceae	C3	RV
4	Canna indica	Cannaceae	C3	RV
5	Cortaderia selloana	Poaceae	C3	PV
6	Cyperus alternifolius	Cyperaceae	C4	PV
7	Limnocharis flava	Alismataceae	C3	RV
8	Oryza rufipogon	Poaceae	C3	PV
9	Oryza sativa	Poaceae	C3	PV
10	Phragmites australis	Poaceae	C4	PV
11	Pontederia cordata	Pontederiaceae	C3	RV
12	Thalia dealbata	Marantaceae	C3	PV
13	Typha angustifolia	Typhaceae	C3	PV
14	Typha orientalis	Typhaceae	C3	PV
	TMH			
15	Alpinia japonica	Zingiberaceae	C3	PV
16	Axonopus compressus	Poaceae	C4	PV
17	Coix lacrymajobi	Poaceae	C4	PV
18	Commelina communis	Commelinaceae	C3	RV
19	Cordyline fruticosa	Asparagaceae	C3	PV
20	Cyperus rotundus	Cyperaceae	C4	PV
21	Digitaria sanguinalis	Poaceae	C4	PV
22	Fargesia spathacea	Poaceae	C3	PV
23	Imperata cylindrica	Poaceae	C4	PV
24	Maranta arundinacea	Marantaceae	C3	PV
25	Oplismenus undulatifolius	Poaceae	C3	PV
26	Panicum bisulcatum	Poaceae	C4	PV
27	Setaria viridis	Poaceae	C4	PV

Table 1. The code, Latin names, Family, photosynthetic pathway as well as leaf vein type of the 14 emergent aquatic and 13 terrestrial monocotyledonous herbs (EMH and TMH) species. RV = reticulate venation, PV = parallel venation.

**Table 2.** Leaf traits (mean  $\pm$  SE) and the results of independent samples t-test between the 14 emergent aquatic and the 13 terrestrial monocotyledonous herb species (EMH and TMH), as well as 5 EMH<sub>p</sub> and 8 TMH<sub>p</sub>, which belong to Poaceae. LA = leaf area, LT = leaf thickness, SD = stomatal density, SL = stomatal length,  $g_{max}$  = maximum modelled stomatal conductance, VLA = vein length per area, VD = vein diameter, CC= the xylem construction cost of leaf veins, SV = stomatal number per vein length.

	Unit	For all		Poaceae	
Trait		ЕМН	TMH	ЕМНр	ТМНр
LA	cm <sup>2</sup>	77.1 ± 10.1	41.4 ± 9.5*	58.1 ± 11.7	25.6 ± 3.9 <sup>ns</sup>
LT	μm	498.7 ± 107.8	$216.1 \pm 33.1^*$	397.3 ± 48.9	147.7 ± 6.2**
SD	no. mm <sup>-2</sup>	514.7 ± 102.0	254.8 ± 43.2*	724.8 ± 84.4	316.3 ± 43.1**
SL	μm	$24.8 \pm 2.0$	$25.4 \pm 2.2^{ns}$	21.7 ± 1.5	$24.2 \pm 2.5^{ns}$
g <sub>max</sub>	molm <sup>-2</sup> s <sup>-1</sup>	$0.47 \pm 0.06$	0.26 ± 0.03**	$0.63 \pm 0.06$	$0.30 \pm 0.02^{**}$
1°VLA	mm mm <sup>-2</sup>	$0.15 \pm 0.06$	$0.07 \pm 0.01^{ns}$	$0.11 \pm 0.01$	$0.09 \pm 0.01^{ns}$
2°VLA	mm mm <sup>-2</sup>	$0.76 \pm 0.16$	$0.69 \pm 0.11^{ns}$	$1.09 \pm 0.1$	$0.83 \pm 0.12^{ns}$
3°VLA	mm mm <sup>-2</sup>	2.57 ± 0.55	6.29 ± 1.18**	$3.49 \pm 0.51$	$8.44 \pm 1.07^*$
major VLA	mm mm <sup>-2</sup>	$3.48 \pm 0.72$	7.05 ± 1.26*	$4.69 \pm 0.6$	9.36 ± 1.15*
minor VLA	mm mm <sup>-2</sup>	$3.44 \pm 0.34$	$1.88 \pm 0.52^{ns}$		
total VLA	mm mm <sup>-2</sup>	$4.7 \pm 0.54$	7.49 ± 1.13*	$4.69 \pm 0.6$	9.47 ± 1.09*
1°VD	μm	800.0 ± 160.7	484.5 ± 69.9	439.2 ± 93.6	$388.7 \pm 56.6^{ns}$
2°VD	μm	143.8 ± 21.8	73.9 ± 8.7**	94.1 ± 7.1	$73.3 \pm 4.7^{ns}$
3°VD	μm	46.1 ± 3.2	31.6 ± 4.3*	$50.2 \pm 2.4$	28.7 ± 3.8**
minor VD	μm	21.7 ± 1.9	16.3 ± 2ns		
1 °CC	1	$0.12 \pm 0.02$	$0.09 \pm 0.01$	$0.12 \pm 0.02$	$0.09 \pm 0.01^{ns}$
2 °CC	/	$0.23 \pm 0.04$	$0.15 \pm 0.02^*$	$0.3 \pm 0.02$	$0.18 \pm 0.02^{**}$
3 °CC	/	$0.32 \pm 0.06$	$0.5 \pm 0.07^{ns}$	$0.51 \pm 0.05$	$0.63 \pm 0.06^{ns}$
minor CC	/	$0.25 \pm 0.03$	$0.07 \pm 0.01^*$		
total CC	/	$0.76 \pm 0.07$	$0.75 \pm 0.08^{ns}$	$0.93 \pm 0.04$	$0.91 \pm 0.06^{ns}$
$g_{max}$ / total VLA	10 <sup>-4</sup> molm <sup>-1</sup> s <sup>-1</sup>	$1.10 \pm 0.12$	0.42 ± 0.07***	$1.40 \pm 0.10$	0.36 ± 0.04***
SV	no.mm <sup>-1</sup>	106.3 ± 14.6	37.1 ± 6.4***	155.6 ± 11.0	33.5 ± 3.0***

\*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001; ns: P > 0.05.

leaf traits between EMH and TMH by using the 'geiger' package (Pennell et al. 2014) in R. We used the linear descriptive analysis to select the most important variables for separating EMH and TMH by using 'caret' packages in R. SMATR v2.0 software was used to examine the differences in linear relationships between EMH and TMH (Warton et al. 2006).

#### Results

We found that EMH had significantly higher mean leaf area, leaf thickness, stomatal density,  $g_{\rm max}$ , major vein diameter (including 1° VD, 2° VD and 3° VD) and stomatal number per vein length, but had lower mean major VLA, total VLA and the xylem construction cost of leaf veins than TMH (Table 2). The results of phylogenetic ANOVA followed the similar pattern with the results of normal one-way ANOVA; however, the differences in stomatal density and total VLA between EMH and TMH became marginally significant when considering the phylogeny relationships [see Supporting Information—Table S1].

Axis 1 and axis 2 of the PCA explained 42.6 and 21.2 % of the total variance, respectively. Axis 1 was loaded by stomatal density and VLA on the positive side and by leaf thickness and vein diameter on the negative side, whereas Axis 2 was loaded by stomatal number per vein length on the positive side (Fig. 2A). EMH and TMH can be separated from one another along axis 2, with EMH distributed on the positive side of axis 2 and TMH distributed on the negative side of axis 2 (Fig. 2B). There were 11 variables selected for the classification of the two groups with the linear discriminant analysis [see Supporting Information— Table S2]. Within the 11 variables, stomatal number per vein length, leaf thickness, second vein diameter, third vein diameter and the xylem construction cost of the third leaf veins were the top five important variables.

A significant and positive correlation between total VLA and stomatal density was found in both EMH and TMH (Fig. 3A; EMH,  $r^2 = 0.34$ , P < 0.05; TMH,  $r^2 = 0.40$ , P < 0.05), and the linear regression slope for these variables in EMH was significantly greater than that of TMH (Fig. 3A). Similarly, the relationships were also significant for TMH of Poaceae species (Fig. 3B;  $r^2 = 0.66$ , P < 0.05), but not significant for EMH of Poaceae species (Fig. 3B;  $r^2 = 0.72$ , P = 0.07).

Stomatal length was significantly and negatively correlated with stomatal density in both EMH and TMH (Fig. 4A; EMH,  $r^2 = 0.82$ , P < 0.001; TMH,  $r^2 = 0.85$ , P < 0.001), and the intercept of the regression line in EMH was significantly higher than that in TMH. The stomatal length was also significantly and negatively correlated with total VLA in both EMH and TMH (Fig. 4B; EMH,  $r^2 = 0.56$ , P < 0.01; TMH,  $r^2 = 0.34$ , P = 0.03), and the intercept of the regression line in EMH was significantly lower than that in TMH.

## Discussion

An important finding of this study is that the mean stomatal density of EMH was more than 2-fold that of TMH, whereas the total VLA of TMH was also significantly higher than that of EMH, which lead to the stomatal number per vein length of EMH was nearly 3-fold as that of TMH. For the Poaceae species only, the stomatal number per vein length in EMH was 5-fold as that of TMH. Compared with TMH, EMH appeared to have adapted to aquatic conditions via enlarged vein diameter and leaf thickness rather than increased VLA. However, we did not observe a positive linear correlation between VLA and stomatal density when the data of all species in both groups were pooled,



Figure 2. First two axes of the principal component analysis for the leaf functional traits and loading of the 27 species along the first two PC axes. The trait codes are as in Table 2 and species codes are as in Table 1.

although this relationship was found when the data of EMH and TMH were evaluated separately. Thus, our results showed that the coordination between leaf water supply and demand was environment-specific.

The results of the present study deepen the understanding of the coordination between stomatal density and VLA in emergent aquatic species. Although the positive linear correlation between stomatal density and VLA has been found in many terrestrial species under various conditions (Brodribb and Jordan 2011; Sun et al. 2014; Carins Murphy et al. 2016; Schneider et al. 2017; Zhao et al. 2017), we extended this correlation to the emergent aquatic herbs in this study. The intercept of the linear relationship between the stomatal density and total VLA in EMH was significantly lower than that of TMH, which indicated on a given value of stomatal density, TMH generally had higher VLA. The significant difference in the average stomatal number per vein length between the two groups also showed the difference in leaf water supply and demand coordination. EMH had significantly higher average stomatal density and lower average total VLA than TMH, which might be mainly because EMH had larger vein diameters (Table 2).



Figure 3. Stomatal density in relation to total vein length per area (total VLA) of terrestrial and emergent aquatic monocotyledonous herbs (TMH and EMH, respectively). Each point represents one species: (A) was for all species and (B) was for Poaceae species. The x-axis and y-axis are logarithmic. The correlations were statistically significant for both 14 EMH (total VLA =  $-0.17 + 0.31 \times SD$ ,  $r^2 = 0.34^{\circ}$ ), 13 TMH (total VLA =  $-0.38 + 0.52 \times SD$ ,  $r^2 = 0.40^{\circ}$ ) and 8 TMH<sub>p</sub> (Poaceae species, total VLA =  $-0.65 + 0.65 \times SD$ ,  $r^2 = 0.66^{\circ}$ ), but not significant for all the 27 species ( $r^2 = 0.11$ , P = 0.08) and 5 EMH<sub>p</sub> ( $r^2 = 0.72$ , P = 0.07).\*P < 0.05.



**Figure 4. Stomatal** length in relation to stomatal density (A) and total vein length per area (B) for 13 terrestrial and 14 emergent aquatic monocotyledonous herbs (TMH and EMH, respectively). Each point represents one species. The x-axis and y-axis are logarithmic. Correlations were statistically significant for both EMH and TMH in (A) (EMH, SD =  $5.8-2.3 \times SL$ ,  $r^2 = 0.82^{***}$ ; TMH, SD =  $5.2-2.1 \times SL$ ,  $r^2 = 0.85^{***}$ ) and (B) (EMH, total VLA =  $2.0 - 1.0 \times SL$ ,  $r^2 = 0.56^{**}$ ; TMH, total VLA =  $2.3 - 1.1 \times SL$ ,  $r^2 = 0.34^*$ ). \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001; ns, P > 0.05.

A comparison of the specific changes in coordination between leaf water supply and demand of different plants could help explain the changes in leaf water-use strategies. Vein density and diameter determine the water transport efficiency of the leaves and reflects transpirational characteristics (Boyce 2009; Sack and Scoffoni 2013). In aquatic environments, a low VLA reduces mesophyll displacement inside leaves (Carins Murphy et al. 2014). Higher major vein diameter might results in larger vessels within these veins, thereby providing greater maximum hydraulic conductivity in EMH species. EMH species evolved higher stomatal density to match this greater flow and theoretical  $g_{\rm max}$ . Furthermore, these leaves are rarely exposed to substrate water deficit so evolving very large major vein diameters and vessels would not be maladaptive because these veins would be rarely exposed to embolism or cell collapse. On the contrary, TMH usually endure more drought stress in the dry season than EMH. Consequently, TMH invest more energy to build denser veins with a smaller diameter in their leaves. Actually, species in drier areas do have higher VLA (Sack and Scoffoni 2013). In this study, we also found that the TMH have significantly higher VLA than EMH species, which might be because of TMH species are more prone to xylem embolism induced by drought. Increasing vein density may provide increasingly redundant pathways for water flow (Scoffoni et al. 2017; Scoffoni and Sack 2017).

The emergent aquatic environment also deeply influenced the stomatal traits of EMH, as the average stomatal density of them was more than 2-fold that of TMH, but the stomatal length of both groups was not significantly different. Monocots have distinctly lower leaf vein densities than other angiosperm subclades (Roddy et al. 2013; de Boer et al. 2016), which indicated that monocots may experience less evolutionary pressure to increase leaf gas exchange capacity despite having both leaf sides available to allocate to stomata (Rudall et al. 2017; Haworth et al. 2018). Alternatively, monocots do experience selection for increased gas exchange capacity, but because of C4 photosynthetic pathway, the scaling of VLA and the maximal photosynthesis is different than in C3 plants (Sack and Scoffoni 2013). Hence, the competitive advantage of spatially optimal location of leaf epidermal area to stomata could be negated by specific growth conditions in relation to leaf hydraulics and leaf morphology (de Boer et al. 2016). In this study, when the data of both the groups were pooled, the stomatal length and total VLA were still significantly and negatively correlated (Fig. 4B;  $r^2 = 0.34$ , P < 0.01), although the intercept of the regression line in EMH was significantly higher than that in TMH. Previous studies had also reported this relationship in other species (Zhang et al. 2012; Zhao et al. 2016).

# Conclusions

The emergent aquatic herbs exhibited considerable differences in their water-related functional traits when compared with terrestrial herbs, with the former having greater water transport capacity and stomatal conductance potential. Although a correlation between stomatal density and total vein density was found in each group, this correlation became non-significant when the data from both the groups were pooled. Our results showed that different water conditions modified the coordination between leaf veins and stomatal traits of emergent aquatic and terrestrial plants. The present study also provided new evidence that supporting the hypothesis of a leaf water supply and demand hypothesis.

# **Supporting Information**

The following supporting information is available in the online version of this article—

Table S1. The results of the one-way ANOVA test and phylogenetic ANOVA.

Table S2. The results of the Linear Discriminant Analysis (LDA) analysis.

Figure S1. Phylogeny tree of the 27 studied species.

# **Sources of Funding**

This study was supported by the National Natural Science Foundation of China (Project No. 41630752, 31800330 and 31870591) and by the Natural Science Foundation of Guangdong Province (2018A030310386). The funders had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript.

# **Contributions by the Authors**

P.Z. and W.Z. designed this experiment and revised the manuscript; W.Z. and G.L. carried out the experiment; W.Z. and P.F. did the data analysis; W.Z. drafted the manuscript; P.Z. and P.F. edited the manuscript. All authors read and approved the final manuscript.

## Acknowledgements

We would like to thank Dr Qinggong Mao for identification of plant specimens, Prof. Qing Ye for the experimental platform service and Dr Jin Zhao for her help in phylogenetic ANOVA analysis.

## **Conflicts of Interests**

The authors declare there are no competing interests.

### **Literature Cited**

- Baresch A, Crifò C, Boyce CK. 2019. Competition for epidermal space in the evolution of leaves with high physiological rates. *The New Phytologist* 221:628–639.
- Boyce CK, Brodribb TJ, Feild TS, Zwieniecki MA. 2009. Angiosperm leaf vein evolution was physiologically and environmentally transformative. Proceedings. Biological Sciences **276**:1771–1776.
- Brodribb TJ, Feild TS, Jordan GJ. 2007. Leaf maximum photosynthetic rate and venation are linked by hydraulics. Plant Physiology 144:1890–1898.
- Brodribb TJ, Jordan GJ. 2011. Water supply and demand remain balanced during leaf acclimation of Nothofagus cunninghamii trees. The New Phytologist **192**:437–448.
- Brodribb TJ, Jordan GJ, Carpenter RJ. 2013. Unified changes in cell size permit coordinated leaf evolution. *The New Phytologist* **199**:559–570.
- Carins Murphy MR, Jordan GJ, Brodribb TJ. 2014. Acclimation to humidity modifies the link between leaf size and the density of veins and stomata. Plant, Cell & Environment **37**:124–131.
- Carins Murphy MR, Jordan GJ, Brodribb TJ. 2016. Cell expansion not cell differentiation predominantly co-ordinates veins and stomata within and among herbs and woody angiosperms grown under sun and shade. Annals of Botany **118**:1127–1138.
- Christin PA, Osborne CP, Chatelet DS, Columbus JT, Besnard G, Hodkinson TR, Garrison LM, Vorontsova MS, Edwards EJ. 2013. Anatomical enablers and the evolution of C4 photosynthesis in grasses. Proceedings of the National Academy of Sciences of the United States of America 110:1381–1386.
- Conklin PA, Strable J, Li S, Scanlon MJ. 2019. On the mechanisms of development in monocot and eudicot leaves. The New Phytologist 221:706–724.
- de Boer HJ, Drake PL, Wendt E, Price CA, Schulze ED, Turner NC, Nicolle D, Veneklaas EJ. 2016. Apparent overinvestment in leaf venation relaxes leaf morphological constraints on photosynthesis in arid habitats. Plant Physiology 172:2286–2299.
- Fanourakis D, Carvalho SM, Almeida DP, Heuvelink E. 2011. Avoiding high relative air humidity during critical stages of leaf ontogeny is decisive for stomatal functioning. Physiologia Plantarum 142:274–286.
- Feild TS, Brodribb TJ. 2013. Hydraulic tuning of vein cell microstructure in the evolution of angiosperm venation networks. The New Phytologist 199:720–726.
- Franks PJ, Beerling DJ. 2009. Maximum leaf conductance driven by  $CO_2$  effects on stomatal size and density over geologic time. Proceedings of the National Academy of Sciences of the United States of America **106**:10343–10347.
- Franks PJ, Farquhar GD. 2001. The effect of exogenous abscisic acid on stomatal development, stomatal mechanics, and leaf gas exchange in Tradescantia virginiana. Plant Physiology **125**:935–942.
- Golub VB, Losev GA, Mirkin BM. 1991. Aquatic and hygrophytic vegetation of the lower Volga valley. Phytocoenologia **20**:1–64.
- Haworth M, Scutt CP, Douthe C, Marino G, Gomes MT, Loreto F, Flexas J, Centritto M. 2018. Allocation of the epidermis to stomata relates to stomatal physiological control: stomatal factors involved in the evolutionary diversification of the angiosperms and development of amphistomaty. Environmental and Experimental Botany 151: 55–63.
- Hetherington AM, Woodward FI. 2003. The role of stomata in sensing and driving environmental change. Nature **424**:901–908.
- Hovenden MJ, Vander Schoor JK, Osanai Y. 2012. Relative humidity has dramatic impacts on leaf morphology but little effect on stomatal index or density in Nothofagus cunninghamii (nothofagaceae). Australian Journal of Botany 60:700–706.
- Lacoul P, Freedman B. 2006. Environmental influences on aquatic plants in freshwater ecosystems. Environmental Reviews 14:89–136.
- Les DH, Schneider EL. 1995. The Nymphaeales, Alismatidae, and the theory of an aquatic monocotyledon origin. In: Rudall PJ, Cribb PJ, Cutler DF, Humphries CJ, eds. Monocotyledons: systematics and evolution. Kew, UK: Royal Botanic Gardens Press, 23–42.
- Lundgren MR, Dunning LT, Olofsson JK, Moreno-Villena JJ, Bouvier JW, Sage TL, Khoshravesh R, Sultmanis S, Stata M, Ripley BS, Vorontsova MS, Besnard G, Adams C, Cuff N, Mapaura A, Bianconi ME, Long CM,

Christin PA, Osborne CP. 2019. C4 anatomy can evolve via a single developmental change. Ecology Letters **22**:302–312.

- Mckown AD, Dengler NG. 2010. Vein patterning and evolution in C4 plants. Botany **88**:775–786.
- McKown AD, Cochard H, Sack L. 2010. Decoding leaf hydraulics with a spatially explicit model: principles of venation architecture and implications for its evolution. *The American Naturalist* **175**:447–460.
- Meinzer FC, Grantz DA. 1991. Coordination of stomatal, hydraulic, and canopy boundary layer properties: do stomata balance conductances by measuring transpiration? Physiologia Plantarum 832:324–329.
- Napp-Zinn K. 1988. Handbuch der Pflanzenanatomie, Bd. VIII, 2B, Teil 2. Berlin, Stuttgart: Bornträger.
- Niklas KJ. 1999. A mechanical perspective on foliage leaf form and function. The New Phytologist 143:19–31.
- Ogburn RM, Edwards EJ. 2013. Repeated origin of three-dimensional leaf venation releases constraints on the evolution of succulence in plants. *Current Biology: CB* 23:722–726.
- Passioura JB. 2002. Soil conditions and plant growth'. Plant, Cell & Environment 25:311–318.
- Pennell MW, Eastman JM, Slater GJ, Brown JW, Uyeda JC, FitzJohn RG, Alfaro ME, Harmon LJ. 2014. Geiger v2.0: an expanded suite of methods for fitting macroevolutionary models to phylogenetic trees. Bioinformatics (Oxford, England) 30:2216–2218.
- Pyykko M. 1979. Morphology and anatomy of leaves from some woody plants in a humid tropical forest of Venezuelan Guayana. Acta bot. fenn1 12: 1–41.
- R Core Team. 2020. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org/
- Roddy AB, Guilliams CM, Lilittham T, Farmer J, Wormser V, Pham T, Fine PV, Feild TS, Dawson TE. 2013. Uncorrelated evolution of leaf and petal venation patterns across the angiosperm phylogeny. *Journal of Experimental Botany* 64:4081–4088.
- Roddy AB, Jiang GF, Cao K, Simonin KA, Brodersen CR. 2019. Hydraulic traits are more diverse in flowers than in leaves. *The New Phytologist* 223:193–203.
- Roddy AB, Simonin KA, McCulloh KA, Brodersen CR, Dawson TE. 2018. Water relations of *Calycanthus* flowers: hydraulic conductance, capacitance, and embolism resistance. *Plant, Cell & Environment* **41**:2250–2262.
- Roddy AB, Théroux-Rancourt G, Abbo T, Benedetti JW, Brodersen CR, Castro M, Castro S, Gilbride AB, Jensen B, Jiang GF, Perkins JA, Perkins SD, Loureiro J, Syed Z, Thompson RA, Kuebbing SE, Simonin KA. 2020. The scaling of genome size and cell size limits maximum rates of photosynthesis with implications for ecological strategies. International Journal of Plant Science 181:75–87.
- Rudall PJ, Chen ED, Cullen E. 2017. Evolution and development of monocot stomata. American Journal of Botany **104**:1122–1141.
- Sack L, Scoffoni C. 2013. Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. The New Phytologist 198:983–1000.

- Sack L, Scoffoni C, McKown AD, Frole K, Rawls M, Havran JC, Tran H, Tran T. 2012. Developmentally based scaling of leaf venation architecture explains global ecological patterns. Nature Communications 3:837.
- Schneider JV, Habersetzer J, Rabenstein R, Wesenberg J, Wesche K, Zizka G. 2017. Water supply and demand remain coordinated during breakdown of the global scaling relationship between leaf size and major vein density. *The New Phytologist* **214**:473–486.
- Scoffoni C, Albuquerque C, Brodersen CR, Townes SV, John GP, Cochard H, Buckley TN, McElrone AJ, Sack L. 2017. Leaf vein xylem conduit diameter influences susceptibility to embolism and hydraulic decline. *The New Phytologist* 213:1076–1092.
- Scoffoni C, Sack L, Ort D. 2017. The causes and consequences of leaf hydraulic decline with dehydration. *Journal of Experimental Botany* 68:4479–4496.
- Simonin KA, Roddy AB. 2018. Genome downsizing, physiological novelty, and the global dominance of flowering plants. PLoS Biology 16:e2003706.
- Sporck MJ, Sack L. 2010. Adaptive radiation of leaf venation and the meaning of disjunct veins in the C<sub>4</sub> Hawaiian Euphorbia. Abstract presented to the Botanical Society of America. Providence, Rhode Island: Botanical Society of America.
- Sun M, Yang SJ, Zhang JL, Bartlett M, Zhang SB. 2014. Correlated evolution in traits influencing leaf water balance in *Dendrobium* (Orchidaceae). *Plant Ecology* 215:1255–1267.
- Ueno O, Kawano Y, Wakayama M, Takeda T. 2006. Leaf vascular systems in  $\rm c_3$  and  $\rm c_4$  grasses: a two-dimensional analysis. Annals of Botany 97:611–621.
- Uhl D, Mosbrugger V. 1999. Leaf venation density as a climate and environmental proxy: a critical review and new data. *Palaeogeography*, *Palaeoclimatology* **149**:15–26.
- Warton DI, Wright IJ, Falster DS, Westoby M. 2006. Bivariate line-fitting methods for allometry. Biological Reviews 81:259–291.
- Zanne AE, Tank DC, Cornwell WK, Eastman JM, Smith SA, FitzJohn RG, McGlinn DJ, O'Meara BC, Moles AT, Reich PB, Royer DL, Soltis DE, Stevens PF, Westoby M, Wright IJ, Aarssen L, Bertin RI, Calaminus A, Govaerts R, Hemmings F, Leishman MR, Oleksyn J, Soltis PS, Swenson NG, Warman L, Beaulieu JM. 2014. Three keys to the radiation of angiosperms into freezing environments. Nature 506:89–92.
- Zhang SB, Dai Y, Hao GY, Li JW, Fu XW, Zhang JL. 2015. Differentiation of water-related traits in terrestrial and epiphytic Cymbidium species. Frontiers in Plant Science 6:260.
- Zhang SB, Guan ZJ, Sun M, Zhang JJ, Cao KF, Hu H. 2012. Evolutionary association of stomatal traits with leaf vein density in *Paphiopedilum*, Orchidaceae. *PLoS One* **7**:e40080.
- Zhao WL, Chen YJ, Brodribb TJ, Cao KF. 2016. Weak co-ordination between vein and stomatal densities in 105 angiosperm tree species along altitudinal gradients in Southwest China. Functional Plant Biology: FPB 43:1126–1133.
- Zhao WL, Siddiq Z, Fu PL, Zhang JL, Cao KF. 2017. Stable stomatal number per minor vein length indicates the coordination between leaf water supply and demand in three leguminous species. *Scientific Reports* 7:2211.