



International Journal of Phytoremediation

ISSN: 1522-6514 (Print) 1549-7879 (Online) Journal homepage: https://www.tandfonline.com/loi/bijp20

Evaluation of hyperaccumulation potentials to cadmium (Cd) in six ornamental species (compositae)

Zhouli Liu, Wei Chen & Xingyuan He

To cite this article: Zhouli Liu, Wei Chen & Xingyuan He (2019): Evaluation of hyperaccumulation potentials to cadmium (Cd) in six ornamental species (compositae), International Journal of Phytoremediation, DOI: 10.1080/15226514.2018.1501343

To link to this article: https://doi.org/10.1080/15226514.2018.1501343



Published online: 17 Jan 2019.



🖉 Submit your article to this journal 🗗

Article views: 9



則 View Crossmark data 🗹

Evaluation of hyperaccumulation potentials to cadmium (Cd) in six ornamental species (compositae)

Zhouli Liu^a, Wei Chen^a, and Xingyuan He^{a,b}

^aCAS Key Laboratory of Forest Ecology and Management, Institute of Applied Ecology, Shenyang, China; ^bUniversity of Chinese Academy of Sciences, Beijing, China

ABSTRACT

Phytoremediation is considered as a promising soil remediation technique. In the present study, the growth responses, cadmium (Cd) accumulation and uptake capability of six popular compositae species, namely, *Taraxacum mongolicum* Hand.-Mazz., *Tagetes erecta* L., *Tagetes patula* L., *Zinnia elegans Jacq., Centaurea cyanus* L. and *Gerbera jamesonii Bolus* under Cd stress were investigated. Among the six compositae species, the growth of *T. erecta* L. and *T. patula* L. improved under 10 mg kg⁻¹ Cd exposure in term of the total biomass and height increased along with the increased Cd concentration in soil, and the growth of the two plants had no significant differences at the high Cd concentration (100 mg kg⁻¹), which indicated that they have good tolerance to Cd toxicity. At the same time, the two plants have higher biomass than four other plants. Furthermore, they can accumulate Cd above 100 μ g g⁻¹ dry tissue, which is the threshold value of a Cd-hyperaccumulator, and have higher Cd uptake ability, translocation factor (TF) and bioconcentration factor (BCF) values. According to these traits, it was shown that *T. erecta* L. and *T. patula* L. had strong tolerance and accumulation capability to Cd, therefore they can become potential hyperaccumulators in phytoremediation of Cd-contaminated soils.

KEY WORDS

Accumulation; compositae species; growth; heavy metal; phytoremediation

Taylor & Francis

Check for updates

Taylor & Francis Group

Introduction

Cadmium (Cd) is one of the highly toxic environmental pollutants (Rui et al. 2016; Fu et al. 2018). The contamination of Cd are produced from non-ferrous metals smelting, mine exploitation, industrial products and agricultural activities. Cd is non-essential element for plants, however, it can be absorbed easily by plants and eventually enter the human body via the food chain (Yasin et al. 2018). Therefore, Cdcontaminated soil is current concern globally because of the serious threat to human health.

Nowadays several technologies have been developed to cleanup Cd-contaminated soil. Phytoremediation is considered as a promising soil remediation technique where the plants with hyperaccumulation ability are used to remove, assimilate or adsorb hazardous pollutants in soil (He et al. 2013; Midhat et al. 2017; Madejón et al. 2018; Zeng et al. 2018). It has recently become a popular research topic in screening potential hyperaccumulators for phytoremediation. Cd concentration in shoots (dry weight) of some species is generally $0.05-0.2 \,\mu g g^{-1}$ (Solis-Dominguez et al. 2007), however, hyperaccumulators can accumulate Cd above 0.01% dry weight (100 μ g g⁻¹) (Liu et al. 2009). Nearly 500 species of hyperaccumulators have been documented all over the world, however, only few plant species such as Thlaspi caerulescens, Arabidopsis halleri, Sedum alfredii, Brassica napus L., Lonicera japonica Thunb. and Coronopus

didymus have been reported (Escarré et al. 2000; Huguet et al. 2012; Jaffré et al. 2013; Wang et al. 2013; Xiao et al. 2013; Liu et al. 2015; Angelova et al. 2017; Sidhu et al. 2017). Application of hyperaccumulators in practice is limited because of low biomass and slow growth rate of hyperaccumulators or accumulators (Liu et al. 2018). Thus, it is important and necessary to identify new universal hyperaccumulators or accumulators of Cd. The plant species with fast growth, high biomass and ecological value, such as ornamental plants, may provide a good cleanup method for Cd-contaminated soil. Some ornamental plants have also been used for the bioaccumulation and remediation of heavy metal and VOCs in contaminated environment (Liu et al. 2011, 2013; Han et al. 2015; Jelusic and Lestan 2015; Rafiq et al. 2016; Teiri et al. 2018; Yan et al. 2018). However, little information is available on the accumulation potential of Cd in compositae species.

Compositae (Asteraceae) family is one of the largest flowering plant families throughout the world. Compositae is important primarily for its many beautiful ornamentals, such as Taraxacum, Tagetes, Zinnia, Centaurea and Gerbera. The compositae species are widely used as landscape greening and horticultural ornamentation. In the present study, six popular compositae species, namely, *Taraxacum mongolicum* Hand.-Mazz., *Tagetes erecta* L., *Tagetes patula* L., *Zinnia elegans Jacq., Centaurea cyanus* L. and *Gerbera jamesonii Bolus* were

CONTACT Zhouli Liu 😒 liuzhouli@hotmail.com; Wei Chen 🐼 forestry83@hotmail.com 🗈 CAS Key Laboratory of Forest Ecology and Management, Institute of Applied Ecology, Shenyang, Liaoning, 110016, China.

^{© 2018} Taylor & Francis Group, LLC

selected. The aims of the study were: (1) to evaluate the effects of different concentrations Cd (0, 10 and 100 mg kg⁻¹) on the growth of six ornamental plants and (2) to identify the accumulation and translocation characteristics of six ornamental plants to Cd stress. Furthermore, it can provide a reference for screening new hyperaccumulators for phytoremediation of Cd-contaminated soil.

Materials and methods

Plant culture and Cd exposure

The pot-culture experiment was carried out in July 2017 at the Shenyang Botanical Garden of Chinese Academy of Sciences $(41^{\circ}46' \text{ N and } 123^{\circ}26' \text{ E})$, which is in the temperate zone with a semi-humid monsoon climate. The average annual temperature is 7.8 °C, the average annual precipitation is 734.5 mm, and the relative air humidity is 65-75%. During the experiment, minimal and maximal temperatures ranged from 15 °C to 20 °C and 25 °C to 31 °C, respectively. The soil used in the pots was collected from the top soil (0-20 cm) of a garden. Table 1 lists physical and chemical properties of the test soil. The air-dried soil samples were sieved through a 3-mm mesh sieve and placed into plastic pots with (20 cm diameter \times 15 cm height), mixed uniformly with the specified concentration of CdCl₂·2.5H₂O solution. Three Cd concentrations were applied: 0 (CK), 10 and 100 mg kg^{-1} . Seeds of six compositae species (T. mongolicum Hand.-Mazz., T. erecta L., T. patula L., Z. elegans Jacq., C. cyanus L. and G. jamesonii Bolus) were obtained from Liaoning Academy of Agricultural Sciences. Table 2 lists the natural properties of sampled plants. The seeds of each species were superficially sterilized by contact with ethanol solutions (70%, v/v) for 1 min, followed by distilled water under agitation for 10 min. Twenty seeds of each species were transplanted into each pot. Each Cd concentration was repeated three times in separate pots. The plants were harvested 8 weeks later for analysis.

Measurements of plant biomass and Cd content in plant tissues

The harvested plants were rinsed with tap water, and the roots were immersed in 20 mM Na₂-EDTA for 15 min to remove Cd adhered to the root surface (Yang et al. 2004). Then, the plants were separated into leaves, stems and roots. They were then separately rinsed with running tap water and distilled water, wiped with tissues and weighed. They were then dried at 105 °C for 30 min, then at 70 °C until weight was constant for Cd content measurement.

Dried plant materials were weighed and ground. The powders were digested with a concentrated acid mixture of $HNO_3/HClO_4$ (3:1, v/v). The Cd concentration in plant tissues was determined with an Optima3000 ICP-AES instrument (Perkin-Elmer, USA).

Table 1. Physical and chemical properties of the test soil.

Soil type	Meadow burozem				
рН	7.25 ± 0.03				
Organic matter (OM) (%)	4.09 ± 0.02				
Cation exchange capacity (CEC) (cmol kg^{-1})	19.53 ± 0.08				
Total N (g kg ^{-1})	3.65 ± 0.03				
Total P (g kg ^{-1})	2.11 ± 0.01				
Total K $(g kg^{-1})$	16.98 ± 0.06				
Cd (mg kg^{-1})	0.15 ± 0.02				

Data analysis

The translocation factor (TF) indicated the ability of plants to translocate heavy metals from the roots to the shoots (Mattina et al. 2003). It was calculated as

$$TF = \frac{The metal concentration in shoots}{The metal concentration in roots}$$

The bioconcentration factor (BCF) was described as (Saraswat and Rai 2018):

 $BGF = \frac{The metal concentration in roots}{The metal concentration in medium}$

Heavy-metal uptake was calculated using the following formula as (Sharma and Agrawal 2006)

Uptake(
$$\mu$$
g plant⁻¹d⁻¹) = $\frac{M_2 W_2 - M_1 W_1}{T_2 - T_1}$

where M_1 and M_2 are metal concentrations in the plant tissue and W_1 and W_2 are the plant biomass at time T_1 (initial sampling) and T_2 (final sampling).

Statistical analyses

All measurements were replicated three times. Means and standard deviations (SD) were calculated by the Microsoft Office Excel 2010 for all the data. One-way analysis of variance was carried out with SPSS 17.0. The significant difference was set between treatments at p < .05 or p < .01. Multiple comparison was also made by the least significant difference (LSD) test.

Results and discussion

Differences in Cd tolerance among the six compositae species

Heavy metal Cd is a non-essential element but has some influence on plant growth. As shown as Figure 1, after 8 weeks exposure to 10 mg kg⁻¹ Cd, the total biomass dry weight of the four plants (*T. mongolicum* Hand.-Mazz., *Z. elegans Jacq., C. cyanus* L. and *G. jamesonii Bolus*) had no significant differences compared with the control, by contrast, the total biomass dry weight of *T. erecta* L. and *T. patula* L. increased significantly (p < .01), indicating that a low Cd concentrations might have a stimulating effect on plant growth. The same phenomenon has been found by Kinraide (1993), Calabrese and Baldwin (2003), Scebba et al. (2006) and Jia et al. (2015), which is also proposed as hormesis by de la Rosa et al. (2004). With the concentration of Cd increasing, chlorosis on the leaves of *C. cyanus* L. and





Figure 1. Effects of Cd soil concentrations on total biomass dry weight of six compositae species. Top legend – concentration of Cd. Values represent mean \pm SD.

G. jamesonii Bolus was observed, and the total biomass dry weight of the two plants had a significant decrease compared with the control (p < .01). However, the total biomass dry weight of the four plants (*T. mongolicum* Hand.-Mazz., *Z. elegans Jacq., T. erecta* L. and *T. patula* L.) had no significant differences compared with the control.

As shown as Figure 2, after 8 weeks exposure to 10 mg kg⁻¹ Cd, the height of the three plants (*T. mongolicum* Hand.-Mazz., *Z. elegans Jacq., C. cyanus* L.) had no significant differences compared with the control, and the height of *T. erecta* L. and *T. patula* L. increased significantly (p < .01), which is also consistent with the increased total biomass dry weight of the two plants. The phenomenon is also termed as hormesis by other researchers (Kovalchuk et al. 2003; de la Rosa et al. 2004; Aina et al. 2007; Seth et al. 2008; Sidhu et al. 2017), which could result from an overcompensation response of cells and organisms to toxic chemicals or the defense mechanisms induced by oxygen free radicals. The underlying mechanism study needs to be further investigated. By contrast, the height of *G. jamesonii*

Bolus had a significant decrease under 10 mg kg^{-1} Cd exposure compared with the control. When the concentration of Cd was up to 100 mg kg^{-1} in soil, the height of the four plants (*T. mongolicum* Hand.-Mazz., *Z. elegans Jacq., C. cyanus* L. and *G. jamesonii* Bolus), decreased significantly compared with the control (p < .01). However, the height of *T. erecta* L. and *T. patula* L. showed no significant differences compared with the control.

Based on these growth traits, it demonstrated that *T.* erecta L. and *T. patula* L. had high tolerance to as much as 100 mg kg⁻¹ Cd, which is in agreement with no obvious changes on the leaves of the two plants. The growth of the two plants was improved at the low Cd concentration (10 mg kg^{-1}) in term of the total biomass and height increased along with the increased Cd concentration in soil. Moreover, among the six compositae species, *T. erecta* L. and *T. patula* L. have higher biomass than four other plants.



Figure 2. Effects of Cd soil concentrations on height of six compositae species. Top legend – concentration of Cd. Values represent mean \pm SD.

This indicates that *T. erecta* L. and *T. patula* L. have good potential in phytoremediation of Cd-contaminated soils, since the tolerance to the toxicity of heavy metal may give an important index to recognize a hyperaccumulator or accumulator (Ernst and Nelissen 2000; Yang et al. 2004; Liu et al. 2009).

Differences in Cd accumulation among the six compositae species

After 8 weeks of Cd-exposure, the effects of Cd soil concentrations on Cd concentration in shoots and roots of six compositae species are shown in Figure 3. The concentrations of accumulated Cd in shoots and roots of the six compositae species all increased significantly with increasing Cd concentrations in soil (p < .01). When the concentration of Cd was up to 100 mg kg^{-1} in soil, the concentrations of accumulated Cd in shoots of T. erecta L., T. patula L. and Z. elegans Jacq. reached 166.07 ± 5.13 , 231.72 ± 6.87 and $109.89 \pm 5.82 \,\mu g$ g^{-1} dry weight (DW), respectively, which is above the threshold value defined for Cd-hyperaccumulator (100 μ g g⁻¹ DW) (Baker and Brooks 1989; Sun et al. 2008). However, the concentrations of accumulated Cd in shoots of T. mongolicum Hand.-Mazz., C. cyanus L. and G. jamesonii Bolus were all less than $100 \,\mu g g^{-1}$ DW. At the same level of Cd concentration (100 mg kg^{-1}), the concentrations of accumulated Cd in roots of T. mongolicum Hand.-Mazz., T. erecta L., T. patula L. and Z. elegans Jacq. reached 109.13 ± 3.29, 177.11 ± 6.65, 202.34 ± 3.67 and $129.18 \pm 4.82 \,\mu g g^{-1}$ DW, respectively. By contrast, the concentrations of accumulated Cd in roots of C. cyanus L. and G. jamesonii Bolus were 85.22 ± 3.91 and $87.79 \pm 6.11 \ \mu g \ g^{-1} \ DW.$

The elevated translocation factor (TF) was used to evaluate the ability of the plant to tolerate and translocate Cd from root to shoots (Wei et al. 2016). As shown as Table 3, when the concentration of Cd was 10 mg kg^{-1} in soil, the



Figure 3. Effects of Cd soil concentrations on Cd concentration in shoots and root of six compositae species. Top legend – concentration of Cd. Values represent mean ± SD.

Table 3. Effects of Cd soil concentrations on Cd uptake, TF and BCF of six compositae species.

Plant species	Cd uptake (μ g plant ⁻¹ d ⁻¹)			TF			BCF		
	0	10	100	0	10	100	0	10	100
Taraxacum mongolicum HandMazz.	0.11 ± 0.01	1.13 ± 0.02	2.11 ± 0.07		0.92	0.86		4.68	1.09
Tagetes erecta L.	0.56 ± 0.03	6.79 ± 0.08	27.45 ± 0.16		0.39	0.93		5.37	1.77
Tagetes patula L.	0.29 ± 0.01	3.92 ± 0.05	17.36 ± 0.13		0.87	1.15		3.99	2.02
Zinnia elegans Jacq.	0.17 ± 0.02	1.98 ± 0.01	4.92 ± 0.09		1.08	0.85		4.02	1.29
Centaurea cyanus L.	0.08 ± 0.01	0.59 ± 0.03	1.16 ± 0.02		1.68	0.82		2.12	0.85
Gerbera jamesonii Bolus	0.21 ± 0.03	1.09 ± 0.02	3.21 ± 0.04		0.81	0.79		2.63	0.88

Notes: Values represent mean \pm SD. TF: the translocation factor; BCF: the bioconcentration factor.

TF values of T. mongolicum Hand.-Mazz., Z. elegans Jacq. and C. cyanus L. were higher than 0.90. When the concentration of Cd was up to 100 mg kg⁻¹ in soil, the TF values of T. erecta L. and T. patula L. increased significantly and reached 0.93 and 1.15. The bioconcentration factor (BCF) was also used to measure the metal accumulation potential of plants with respect to the metal concentration in soil (Saraswat and Rai 2018). When the concentration of Cd was 10 mg kg^{-1} in soil, the BFC values of *T. mongolicum* Hand.-Mazz., T. erecta L., T. patula L. and Z. elegans Jacq. were higher than 4.00. When the concentration of Cd was up to 100 mg kg^{-1} in soil, the BCF values of *T. erecta* L. and *T.* patula L. still reached 1.77 and 2.02. The results above suggested that the two plants (T. erecta L. and T. patula L.) had stronger tolerance to high concentration Cd and had better potential in translocating Cd more efficiently.

Cd uptake in six compositae species varied with increasing Cd concentrations in the soil (Table 3). When the concentration of Cd was 10 mg kg^{-1} in soil, Cd uptake of T. erecta L. and T. patula L. were 6.79 ± 0.08 and $3.92 \pm 0.05 \,\mu g$ $plant^{-1}d^{-1}$, which is higher than four other plants. When the concentration of Cd was up to 100 mg kg^{-1} in the soil, Cd uptake of increased significantly and reached 27.45 ± 0.16 and $17.36 \pm 0.13 \,\mu g$ plant⁻¹d⁻¹, and Cd uptake of the four other plants were all less than 5.00 μ g plant⁻¹d⁻¹. There was a positive correlation between Cd uptake and Cd accumulation in T. erecta L. and T. patula L., indicating the two plants may accumulate larger amounts of Cd when exposed to higher Cd concentrations in the soil. Based on higher Cd uptake, TF and BCF values, the higher concentrations of accumulated Cd in shoots and roots of T. erecta L. and T. patula L. indicating that the two plants have the potential for phytoremediation of Cd-contaminated soils.

Conclusions

In the present study, exposure to 10 mg kg^{-1} Cd, the total biomass and height of *T. erecta* L. and *T. patula* L. all increased significantly, indicating that low Cd concentrations might have a stimulating effect on plant growth. When the concentration of Cd was up to 100 mg kg^{-1} , the total biomass and height of the two plants had no significant differences compared with the control, which is in agreement with the observation of no obvious changes on the leaves of the two plants. At the same time, the two plants have higher biomass than four other plants among the six compositae species. Moreover, *T. erecta* L. and *T. patula* L. can accumulate Cd above 100 µg g^{-1} DW, which is the threshold value

of Cd-hyperaccumulator, and have higher Cd uptake ability, TF and BCF values. According to these traits, it is shown T. *erecta* L. and T. *patula* L. can become potential hyperaccumulators in phytoremediation of Cd-contaminated soils. On the one hand, as popular ornamental plants, T. *erecta* L. and T. *patula* L. have the double advantages of beautifying the environment and purifying the soil. On the other hand, coming from the same families and genera, the two plants have the similar capability of Cd hyperaccumulation, and our results might suggest that most of the genera plants possess the same hyperaccumulation characteristics to Cd or not? The present study will also provide an important reference for understanding Cd tolerant strategies in hyperaccumulator cells.

Funding

This work was supported by the National Key Research and Development Program [2016YFC0500300] of China Project.

References

- Aina R, Labra M, Fumagalli P, Vannini C, Marsoni M, Cucchi U, Bracale M, Sgorbati S, Citterio S. 2007. Thiol-peptide level and proteomic changes in response to cadmium toxicity in *Oryza sativa* L. roots. Environ Exp Bot. 59(3):381–392. doi:10.1016/j.envexpbot. 2006.04.010.
- Angelova VR, Ivanova RI, Todorov JM, Ivanov KI. 2017. Potential of rapeseed (*Brassica napus* L.) for phytoremediation of soils contaminated with heavy metals. J Environ Prot Ecol. 18(2):468–478.
- Baker AJM, Brooks RR. 1989. Terrestrial higher plants which hyperaccumulate metallic elements – a review of their distribution, ecology and phytochemistry. Biorecovery. 1:81–126.
- Calabrese EJ, Baldwin LA. 2003. Toxicology rethinks its central belief: hormesis demands. A reappraisal of the way risks are assessed. Nature. 421(6924):691-692. doi:10.1038/421691a.
- de la Rosa G, Peralta-Videa JR, Montes M, Parsons JG, Cano-Aguilera I, Gardea-Torresdey JL. 2004. Cadmium uptake and translocation in tumbleweed (*Salsola kali*), a potential Cd-hyperaccumulator desert plant species: ICP/OES and XAS studies. Chemosphere. 55(9):1159–1168. doi:10.1016/j.chemosphere.2004.01.028.
- Ernst WHO, Nelissen HJM. 2000. Life-cycle phases of a zinc- and cadmium-resistant ecotype of *Silene vulgaris* in risk assessment of polymetallic mine soils. Environ Pollut. 107(3):329–338. doi:10.1016/ S0269-7491(99)00174-8.
- Escarré J, Lefèbvre C, Gruber W, Leblanc M, Lepart J, Rivière Y, Delay B. 2000. Zinc and cadmium hyperaccumulation by thlaspi caerulescens from metalliferous and nonmetalliferous sites in the Mediterranean area: implications for phytoremediation. New Phytol. 145(3):429–437. doi:10.1046/j.1469-8137.2000.00599.x.
- Fu H, Yu H, Li T, Zhang X. 2018. Influence of cadmium stress on root exudates of high cadmium accumulating rice line (*Oryza sativa* L.). Ecotox Environ Safe. 150:168–175. doi:10.1016/j.ecoenv.2017.12.014.

- Han Y, Chen G, Chen Y, Shen Z. 2015. Cadmium toxicity and alleviating effects of exogenous salicylic acid in iris hexagona. Bull Environ Contam Toxicol. 95(6):796–802. doi:10.1007/s00128-015-1640-3.
- He J, Ma C, Ma Y, Li H, Kang J, Liu T, Polle A, Peng C, Luo ZB. 2013. Cadmium tolerance in six poplar species. Environ Sci Pollut Res. 20(1):163–174. doi:10.1007/s11356-012-1008-8.
- Huguet S, Bert V, Laboudigue A, Barthès V, Isaure MP, Llorens I, Schat H, Sarret G. 2012. Cd speciation and localization in the hyperaccumulator arabidopsis halleri. Environ Exp Bot. 82(5):54–65. doi:10.1016/j.envexpbot.2012.03.011.
- Jaffré T, Pillon Y, Thomine S, Merlot S. 2013. The metal hyperaccumulators from new caledonia can broaden our understanding of nickel accumulation in plants. Front Plant Sci. 4(9):279. doi:10.3389/ fpls.2013.00279.
- Jelusic M, Lestan D. 2015. Remediation and reclamation of soils heavily contaminated with toxic metals as a substrate for greening with ornamental plants and grasses. Chemosphere. 138:1001–1007. doi:10.1016/j.chemosphere.2014.12.047.
- Jia L, Liu Z, Chen W, Ye Y, Yu S, He X. 2015. Hormesis effects induced by cadmium on growth and photosynthetic performance in a hyperaccumulator, *Lonicera japonica* Thunb. J Plant Growth Regul. 34(1):13–21. doi:10.1007/s00344-014-9433-1.
- Kinraide TB. 1993. Aluminum enhancement of plant growth in acid rooting media. A case of reciprocal alleviation of toxicity by two toxic cations. Physiol Plantarum. 88(4):619–625. doi:10.1111/j.1399-3054.1993.tb01380.x.
- Kovalchuk I, Filkowski J, Smith K, Kovalchuk O. 2003. Reactive oxygen species stimulate homologous recombination in plants. Plant Cell Environ. 26(9):1531–1539. doi:10.1046/j.1365-3040.2003.01076.x.
- Liu Z, Chen W, He X. 2015. Influence of Cd²⁺ on growth and chlorophyll fluorescence in a hyperaccumulator – *Lonicera japonica* Thunb. J Plant Growth Regul. 34(3):672–676. doi:10.1007/s00344-015-9483-z.
- Liu Z, He X, Chen W. 2011. Effects of cadmium hyperaccumulation on the concentrations of four trace elements in *Lonicera japonica* Thunb. Ecotoxicology. 20(4):698–705. doi:10.1007/s10646-011-0609-1.
- Liu Z, He X, Chen W, Yuan F, Yan K, Tao D. 2009. Accumulation and tolerance characteristics of cadmium in a potential hyperaccumulator – *Lonicera japonica* Thunb. J Hazard Mater. 169(1–3): 170–175. doi:10.1016/j.jhazmat.2009.03.090.
- Liu Z, He X, Chen W, Zhao M. 2013. Ecotoxicological responses of three ornamental herb species to cadmium. Environ Toxicol Chem. 32(8):1746–1751. doi:10.1002/etc.2237.
- Liu JW, Xin X, Zhou QX. 2018. Phytoremediation of contaminated soils using ornamental plants. Environ Rev. 26(1):43–54. doi:10.1139/er-2017-0022.
- Madejón P, Domínguez MT, Madejón E, Cabrera F, Marañón T, Murillo JM. 2018. Soil-plant relationships and contamination by trace elements: A review of twenty years of experimentation and monitoring after the Aznalcóllar (SW Spain) mine accident. Sci Total Environ. 625:50–63. doi:10.1016/j.scitotenv.2017.12.277.
- Mattina MI, Lannucci-Berger W, Musante C, White JC. 2003. Concurrent plant uptake of heavy metals and persistent organic pollutants from soil. Environ Pollut. 124(3):375–378. doi:10.1016/ S0269-7491(03)00060-5.
- Midhat L, Ouazzani N, Esshaimi M, Ouhammou A, Mandi L. 2017. Assessment of heavy metals accumulation by spontaneous vegetation: screening for new accumulator plant species grown in Kettara mine-Marrakech, Southern Morocco. Int J Phytoremediat. 19(2):191–198. doi:10.1080/15226514.2016.1207604.
- Rafiq M, Farooq U, Athar M, Salman M, Aslam M, Raza HMH. 2016. Gardenia jasminoides: an ornamental plant for the biosorption of lead and cadmium ions. Desalin Water Treat. 57(22):10432–10442. doi:10.1080/19443994.2015.1035341.

- Rui H, Chen C, Zhang X, Shen Z, Zhang F. 2016. Cd-induced oxidative stress and lignification in the roots of two *Vicia sativa* L. varieties with different Cd tolerances. J Hazard Mater. 301:304–313. doi:10.1016/j.jhazmat.2015.08.052.
- Saraswat S, Rai JPN. 2018. Aquatic macrophytes mediated remediation of toxic metals from moderately contaminated industrial effluent. Int J Phytoremediat. 20(9):876–884. doi:10.1080/15226514.2018. 1438359
- Scebba F, Arduini I, Ercoli L, Sebastiani L. 2006. Cadmium effects on growth and antioxidant enzymes activities in *Miscanthus sinensis*. Biol Plant. 50(4):688–692. doi:10.1007/s10535-006-0107-0.
- Seth CS, Chaturvedi PK, Misra V. 2008. The role of phytochelatins and antioxidants in tolerance to Cd accumulation in *Brassica juncea* L. Ecotoxicol Environ Saf. 71(1):76–85. doi:10.1016/j.ecoenv.2007. 10.030.
- Sharma RK, Agrawal M. 2006. Single and combined effects of cadmium and zinc on carrots: uptake and bioaccumulation. J Plant Nutr. 29(10):1791–1804. doi:10.1080/01904160600899246.
- Sidhu GPS, Singh HP, Batish DR, Kohli RK. 2017. Tolerance and hyperaccumulation of cadmium by a wild, unpalatable herb *Coronopus didymus* (L.) Sm.(Brassicaceae). Ecotoxicol Environ Saf. 135:209–215. doi:10.1016/j.ecoenv.2016.10.001.
- Solis-Dominguez FA, Gonzalez-Chavez MC, Carrillo-Gonzalez R, Rodriguez-Vazquez R. 2007. Accumulation and localization of cadmium in *Echinochloa polystachya* grown within a hydroponic system. J Hazard Mater. 141(3):630–636. doi:10.1016/j.jhazmat. 2006.07.014.
- Sun YB, Zhou QX, Diao CY. 2008. Effects of cadmium and arsenic on growth and metal accumulation of Cd-hyperaccumulator Solanum nigrum L. Bioresource Technol. 99(5):1103–1110. doi:10.1016/ j.biortech.2007.02.035.
- Teiri H, Pourzamani H, Hajizadeh Y. 2018. Phytoremediation of VOCs from indoor air by ornamental potted plants: a pilot study using a palm species, under the controlled environment. Chemosphere. 197:375–381. doi:10.1016/j.chemosphere.2018.01.078.
- Wang K, Huang H, Zhu Z, Li T, He Z, Yang X, Alva A. 2013. Phytoextraction of metals and rhizoremediation of pahs in cocontaminated soil by co-planting of sedum alfredii with ryegrass (*Lolium perenne*) or castor (*Ricinus communis*). Int J Phytoremediat. 15(3):283–298. doi:10.1080/15226514.2012.694501.
- Wei R, Guo Q, Wen H, Liu C, Yang J, Peters M, Hu J, Zhu G, Zhang H, Tian L, et al. 2016. Fractionation of stable cadmium isotopes in the cadmium tolerant *Ricinus communis* and hyperaccumulator *Solanum nigrum*. Sci Rep. 6(1):24309. doi:10.1038/srep24309.
- Xiao W, Wang H, Li T, Zhu Z, Zhang J, He Z, Yang X. 2013. Bioremediation of Cd and carbendazim co-contaminated soil by Cdhyperaccumulator sedum alfredii associated with carbendazimdegrading bacterial strains. Environ Sci Pollut Res. 20(1):380–389. doi:10.1007/s11356-012-0902-4.
- Yan YY, Wang JJ, Lan XY, Wang QM, Xu FL. 2018. Comparisons of cadmium bioaccumulation potentials and resistance physiology of *Microsorum pteropus* and *Echinodorus grisebachii*. Environ Sci Pollut R.1–8. doi:10.1007/s11356-018-1486-4.
- Yang XE, Long XX, Ye HB, He ZL, Calvert DV, Stoffella PJ. 2004. Cadmium tolerance and hyperaccumulation in a new Zn-hyperaccumulating plant species (*Sedum alfredii* Hance). Plant Soil. 259(1/ 2):181–189. doi:10.1023/B:PLSO.0000020956.24027.f2.
- Yasin NA, Zaheer MM, Khan WU, Ahmad SR, Ahmad A, Ali A, Akram W. 2018. The beneficial role of potassium in Cd-induced stress alleviation and growth improvement in *Gladiolus grandiflora* L. Int J Phytoremediat. 20(3):274–283. doi:10.1080/15226514.2017. 1374337.
- Zeng P, Guo Z, Cao X, Xiao X, Liu Y, Shi L. 2018. Phytostabilization potential of ornamental plants grown in soil contaminated with cadmium. Int J Phytoremediat. 20(4):311–320. doi:10.1080/ 15226514.2017.1381939.